

# Surface Patterning and Polymer Chemistry

## Lesson 9

MSE 304

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# Plan of the Course: Fundamentals, Characterization, and Applications

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1: Intro to Surfaces & Interfaces

2: Surfaces in the Real World - Adsorption

3: Surface Energetics & Interfacial Phenomena

4: Atomic Structure of Real Surfaces

5: Solid-Solid Interfaces

6: From Ideal Planes to Real Materials (Recap)

7: Characterization of Surfaces & Interfaces

8: Surface Chemistry

9: Surface Patterning and Polymer Chemistry

10: Characterization of Molecular Assembly

11: Electronic Properties of Surfaces

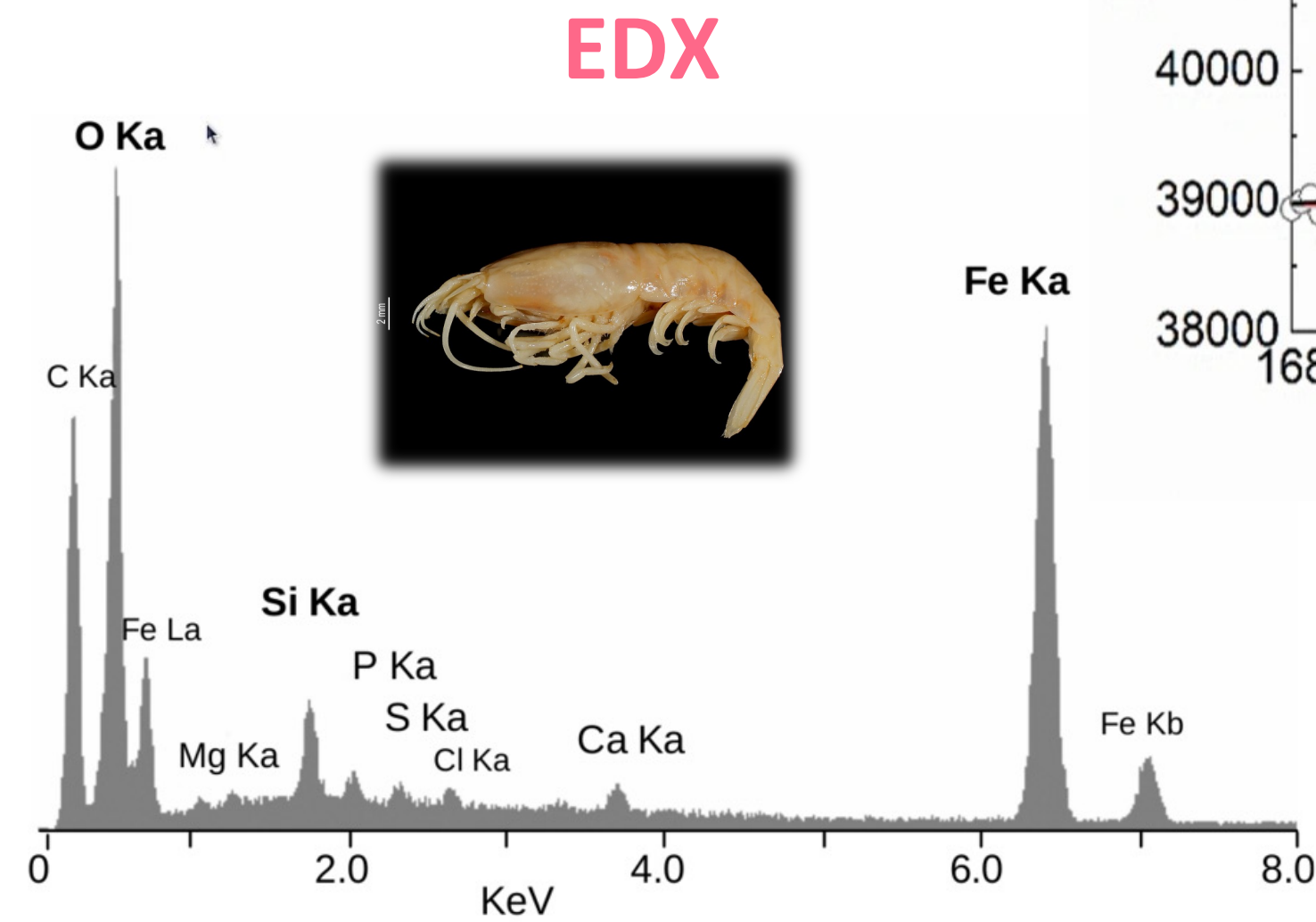
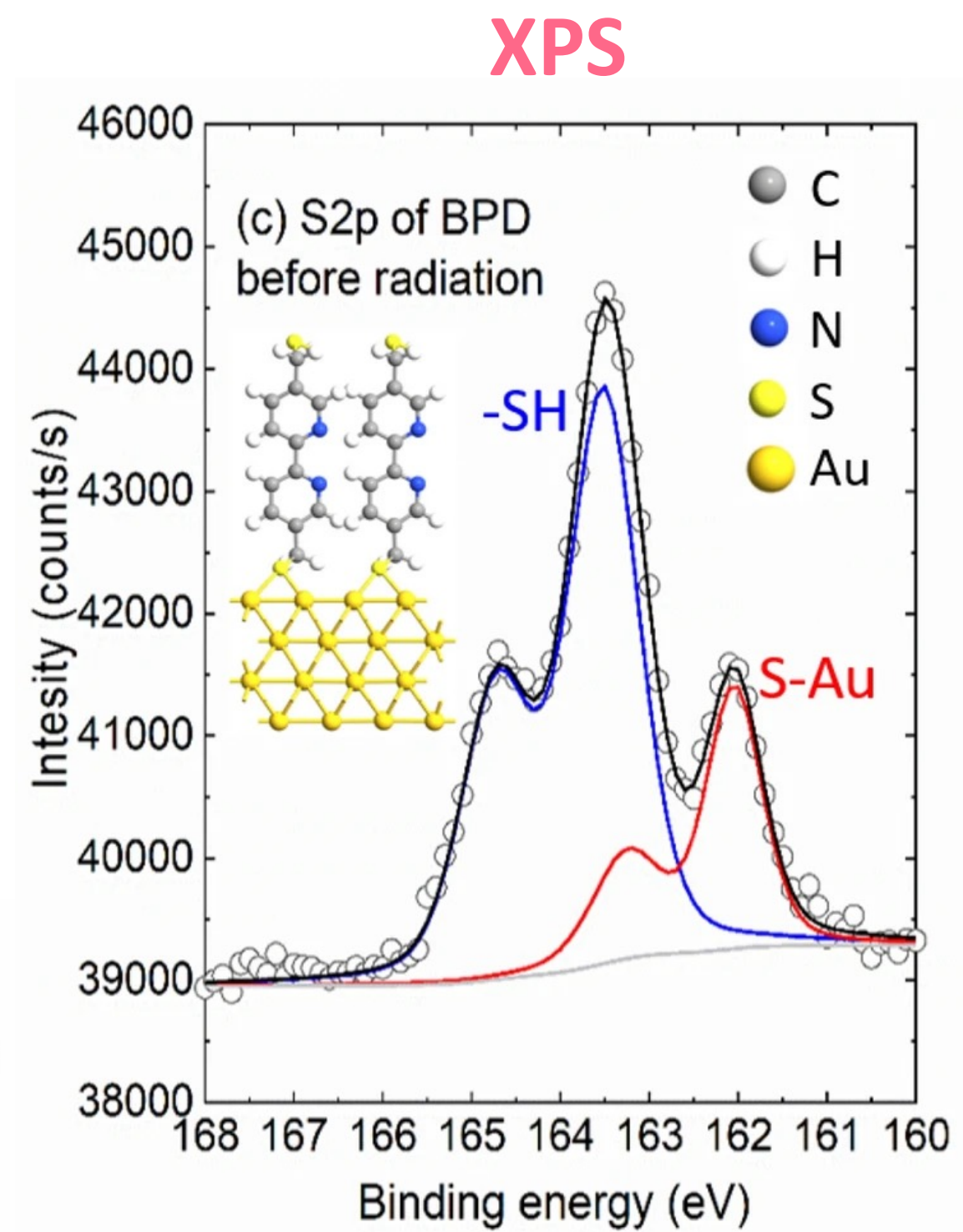
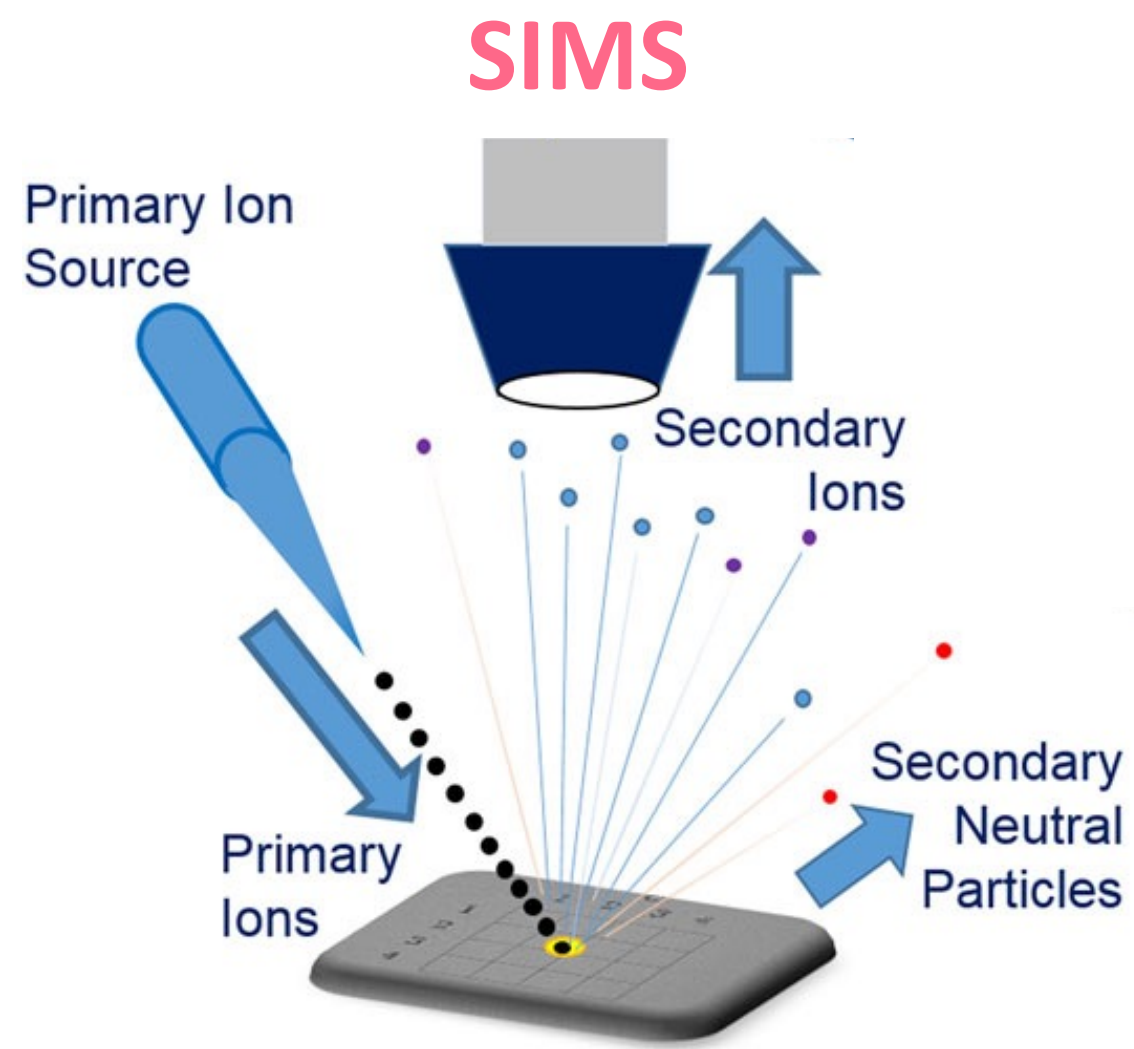
12: Biosensor Fundamentals

13: Biosensing applications

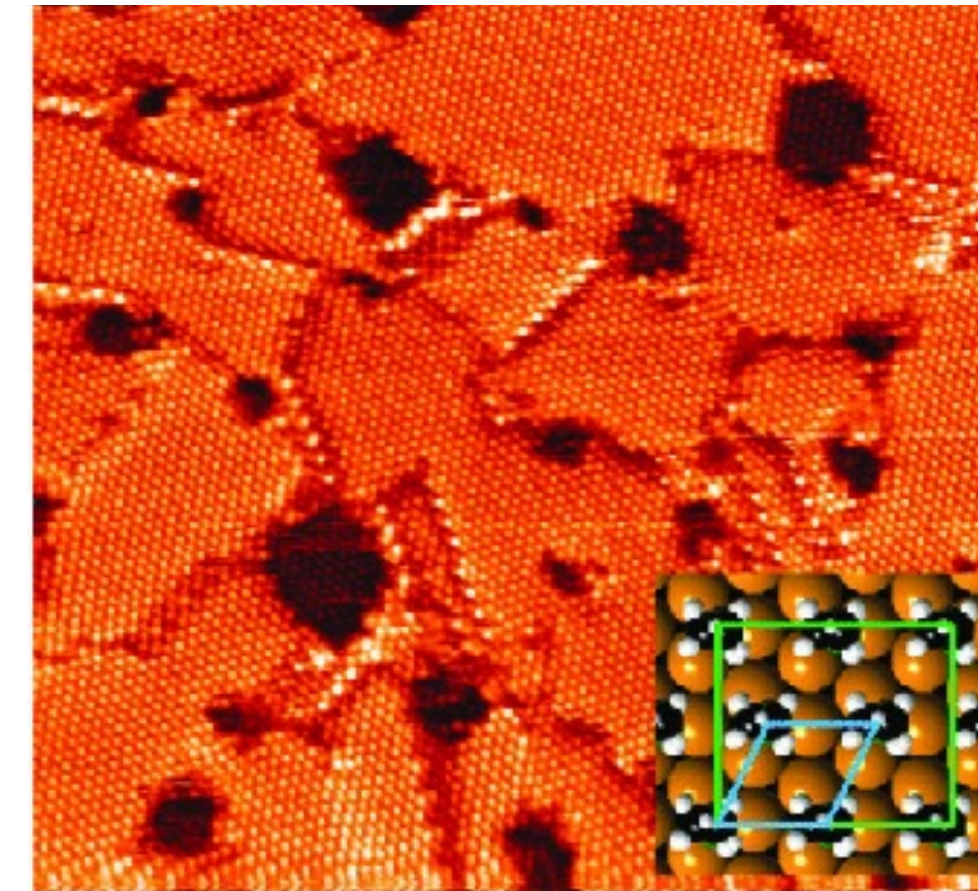
14: Chemistry of Semiconductor Surfaces and Beyond



# Recap from Lesson 6/7: Surface Chemistry Characterization

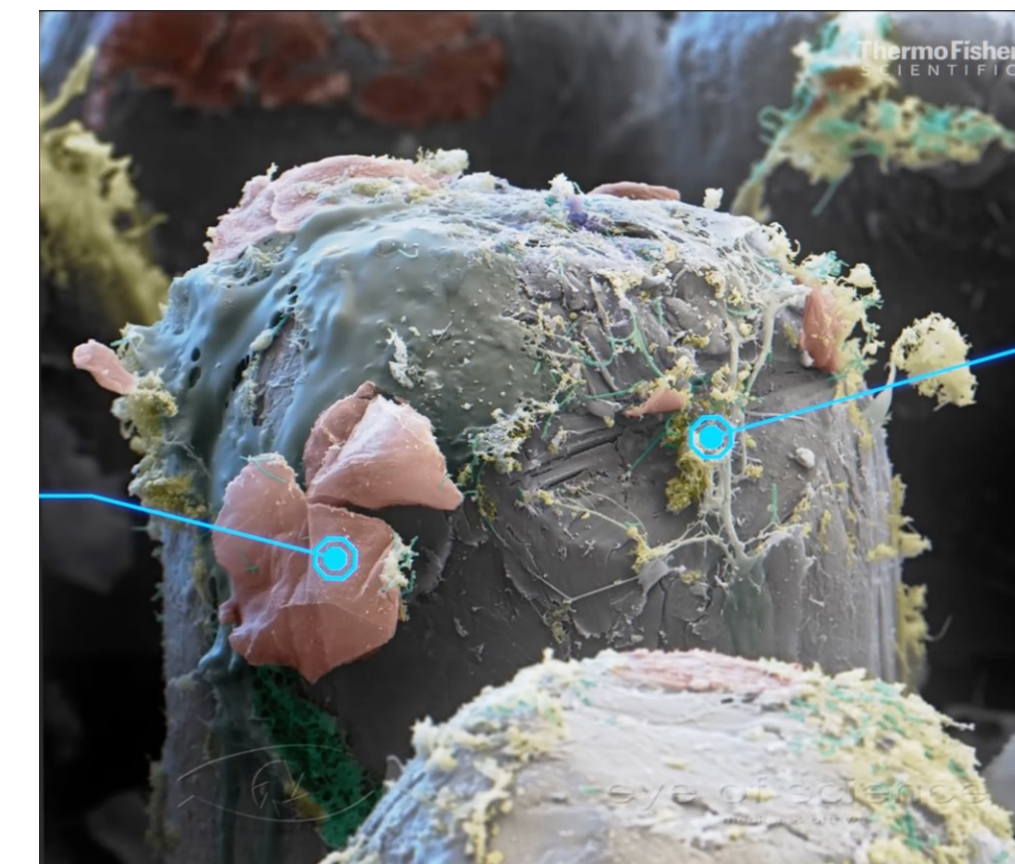


### STM

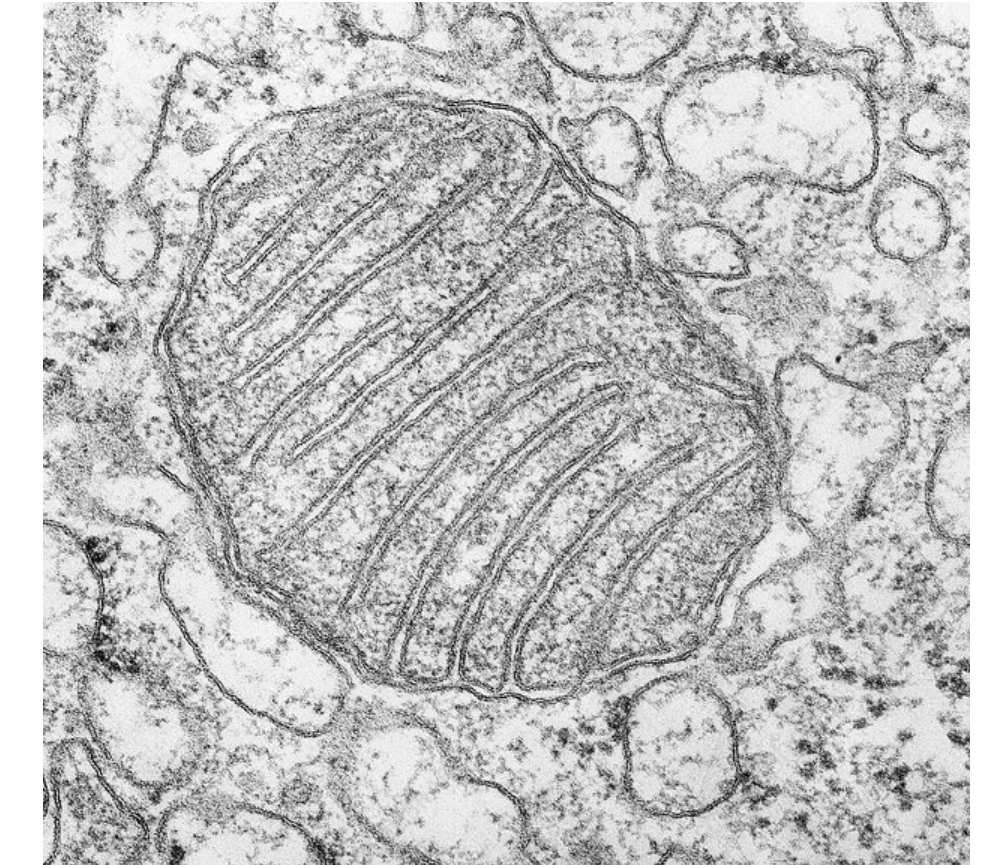


5 nm

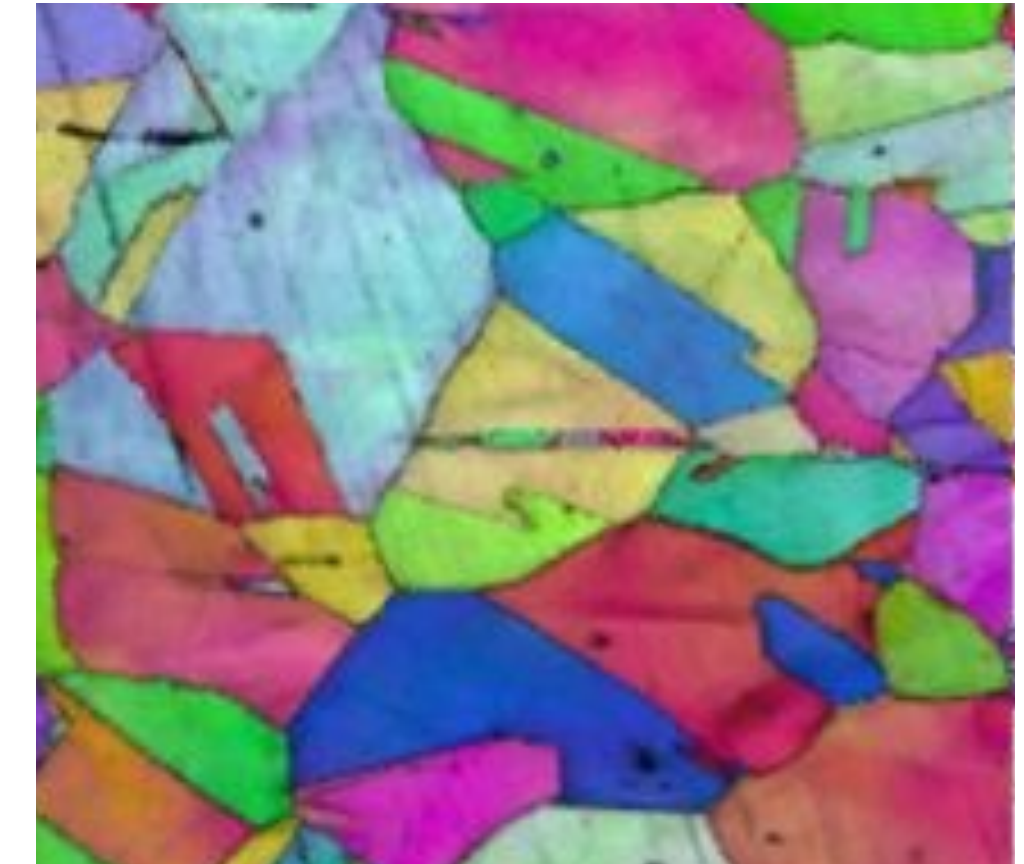
### SEM



### TEM



### EBSD



# Recap from Lesson 6: Surface Characterization Techniques

What techniques enable **quantitative analysis of elements?**

Quantitative analysis is the determination of how much of a substance is present

It gives numerical results (% , concentrations, thickness, counts), not just identification

Category	Techniques	What They Provide
Quantitative / Semi-Quantitative	XPS, EDX, SIMS (with caveats)	Elemental or molecular amounts, sometimes depth
Qualitative (Structural / Imaging)	STM, TEM, SEM, EBSD	Shape, structure, lattice, atomic arrangement — but not how much

# Elemental Quantification Reliability Ranking

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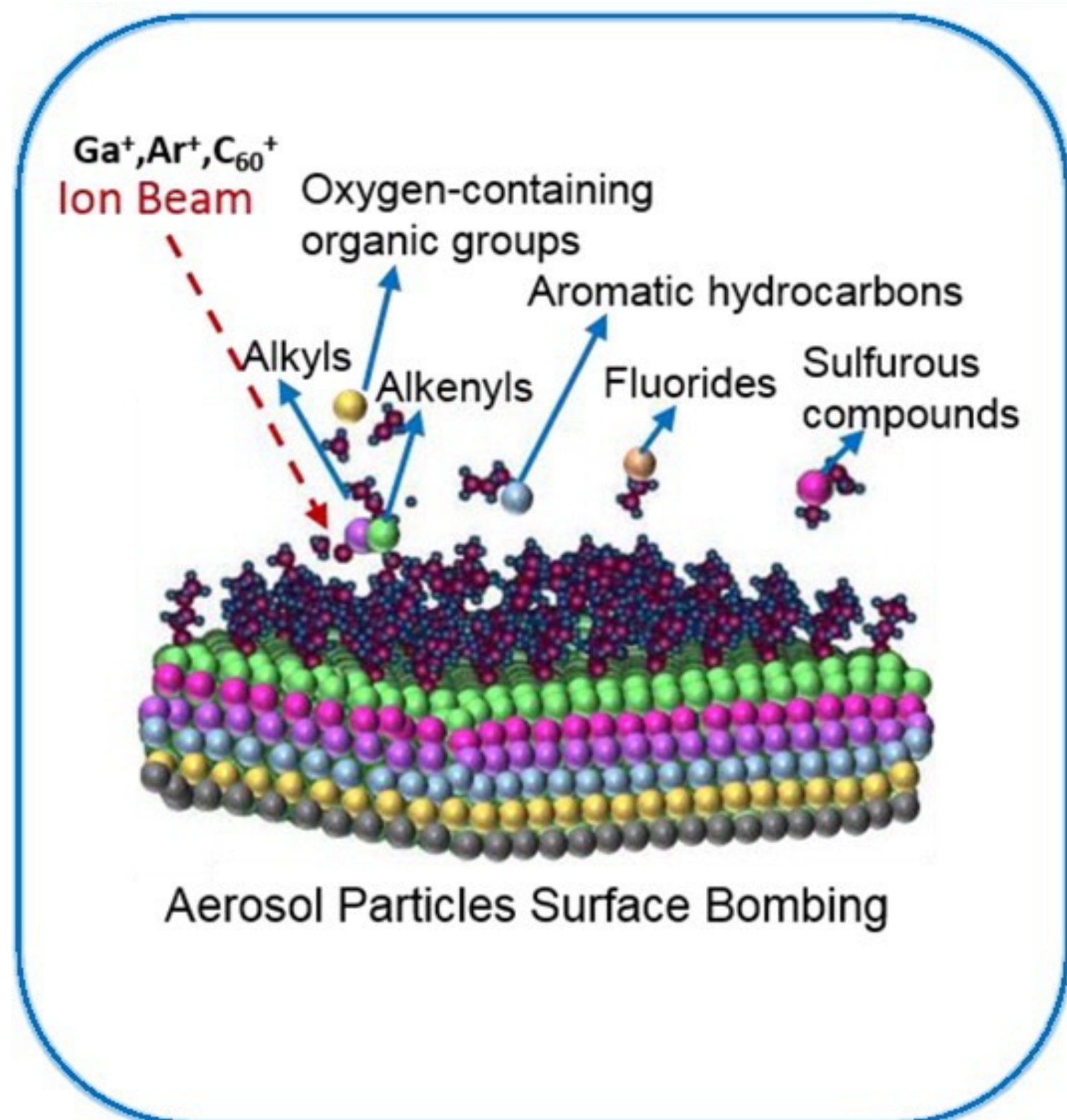
**XPS** = quantitative surface chemistry

**EDX** = rough elemental composition

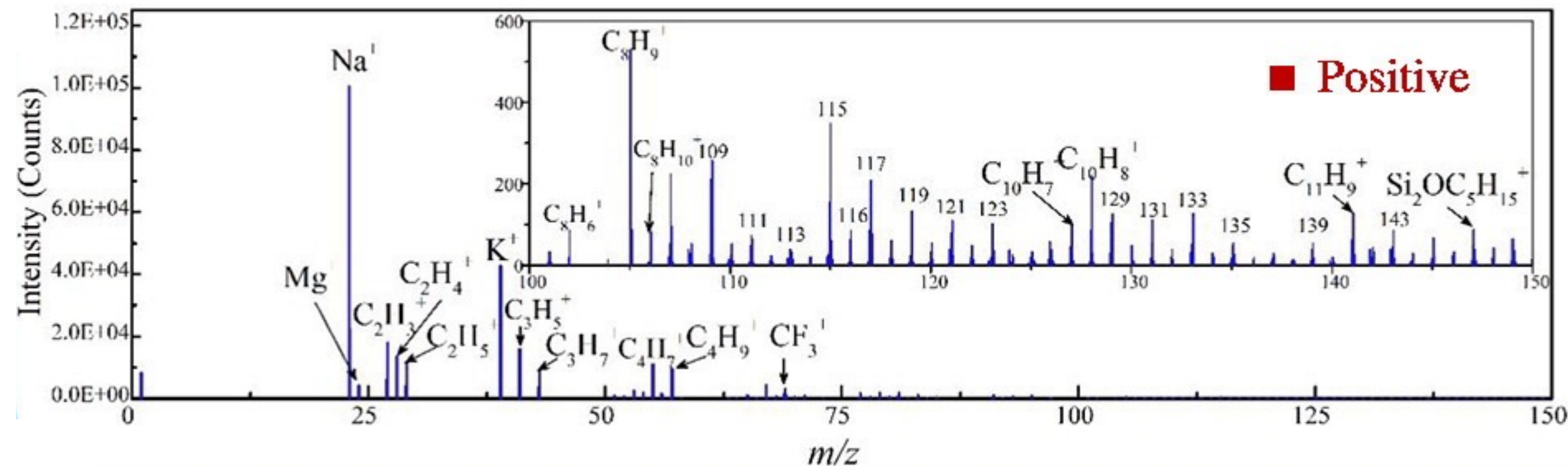
**SIMS** = relative composition and depth trends

Technique	Quality	Strength	Limitation	Best Use
XPS	Most reliable for surface atomic %	Chemical states + surface composition (~5–10 nm)	Needs flat, uniform samples	SAMs, oxides, thin coatings
EDX (in SEM/TEM)	Semi-quantitative (~±5–10%)	Fast, easy elemental survey	Poor for light elements; sees deeper (~0.5–2 μm)	Rough composition of bulk
SIMS	Relative quantification only	Extremely sensitive to trace organics	Need standards for absolute amounts	Surface contamination, polymer chemistry

# SIMS Requires a “Standard” for Quantification



Li *et al.* | J. Environ. Sci. | 2018



SIMS measures intensity of ion fragments, not actual amount of material

The intensity depends on:

- How easily that element/fragment ionizes
- Surrounding chemical environment
- Instrument settings

So **raw SIMS signal  $\neq$  concentration**  $\rightarrow$  need a reference sample (**standard**)

- Known composition
- Same environment

# Quantitative Analysis of Height

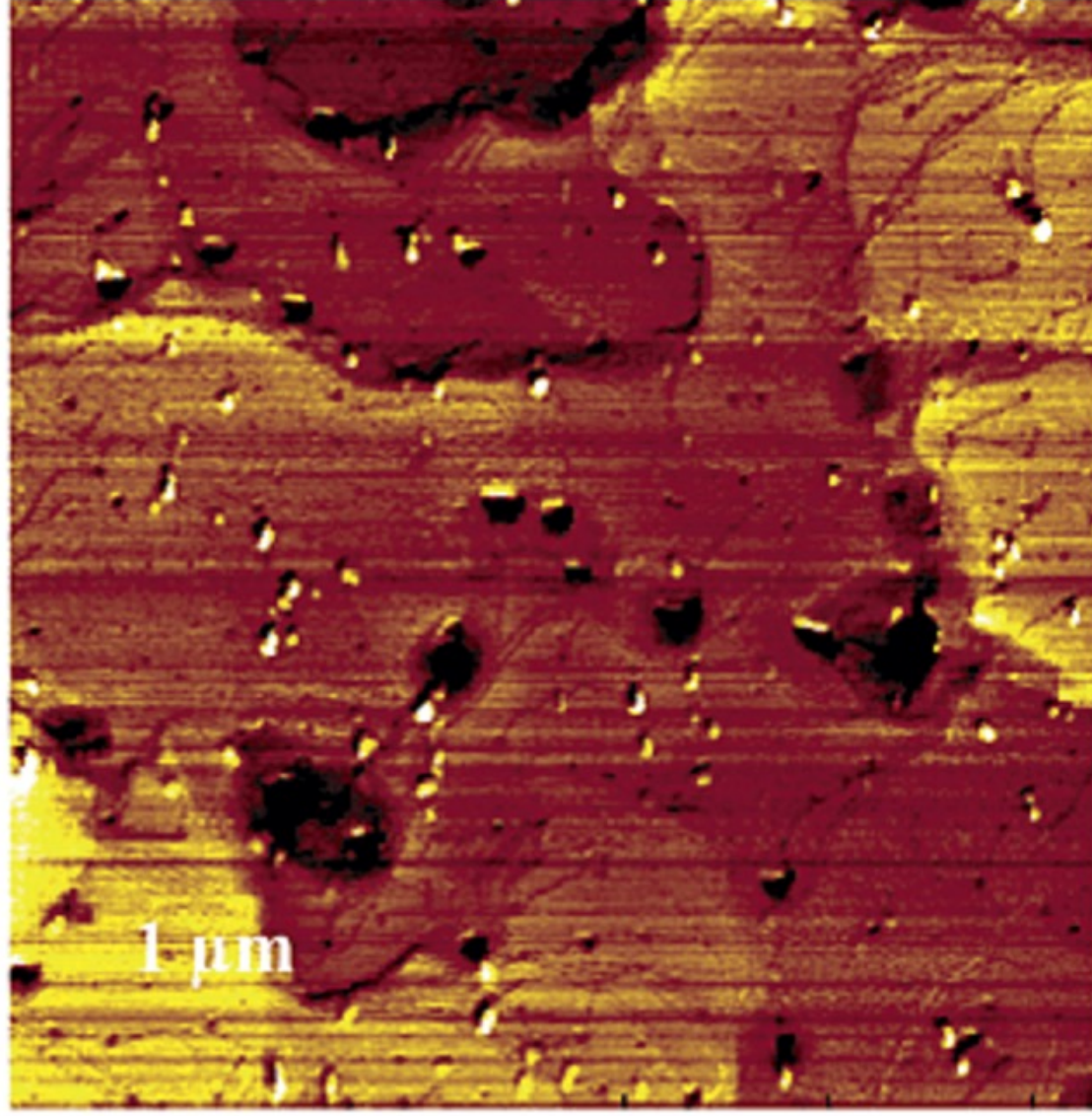
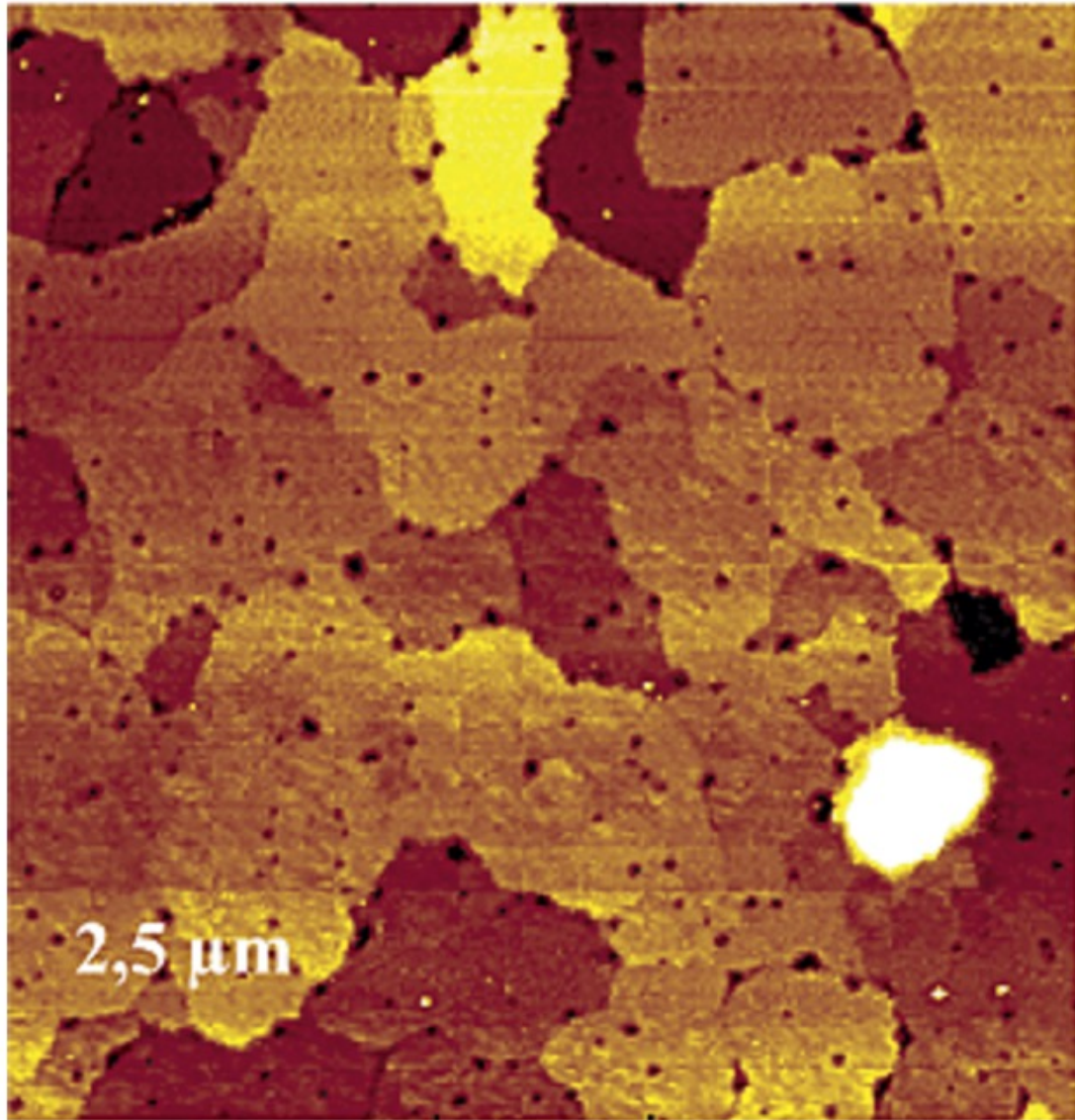
What techniques enable **quantitative analysis of height/thickness**?

Technique	Quality of height measure	How it works	Typical thickness range
STM	Direct, very accurate	Measures tunneling current change as tip follows surface	Sub-nanometer to a few nm (atomic steps)
TEM (cross-section)	Direct in thin sample	Image sliced cross-section and measure	1–200 nm layer thickness very precisely
SEM (cross-section)	Indirect, from image	Tilt sample + measure thickness from magnified image	~50 nm to many microns

XPS and SIMS can only estimate thickness based on models/calibration

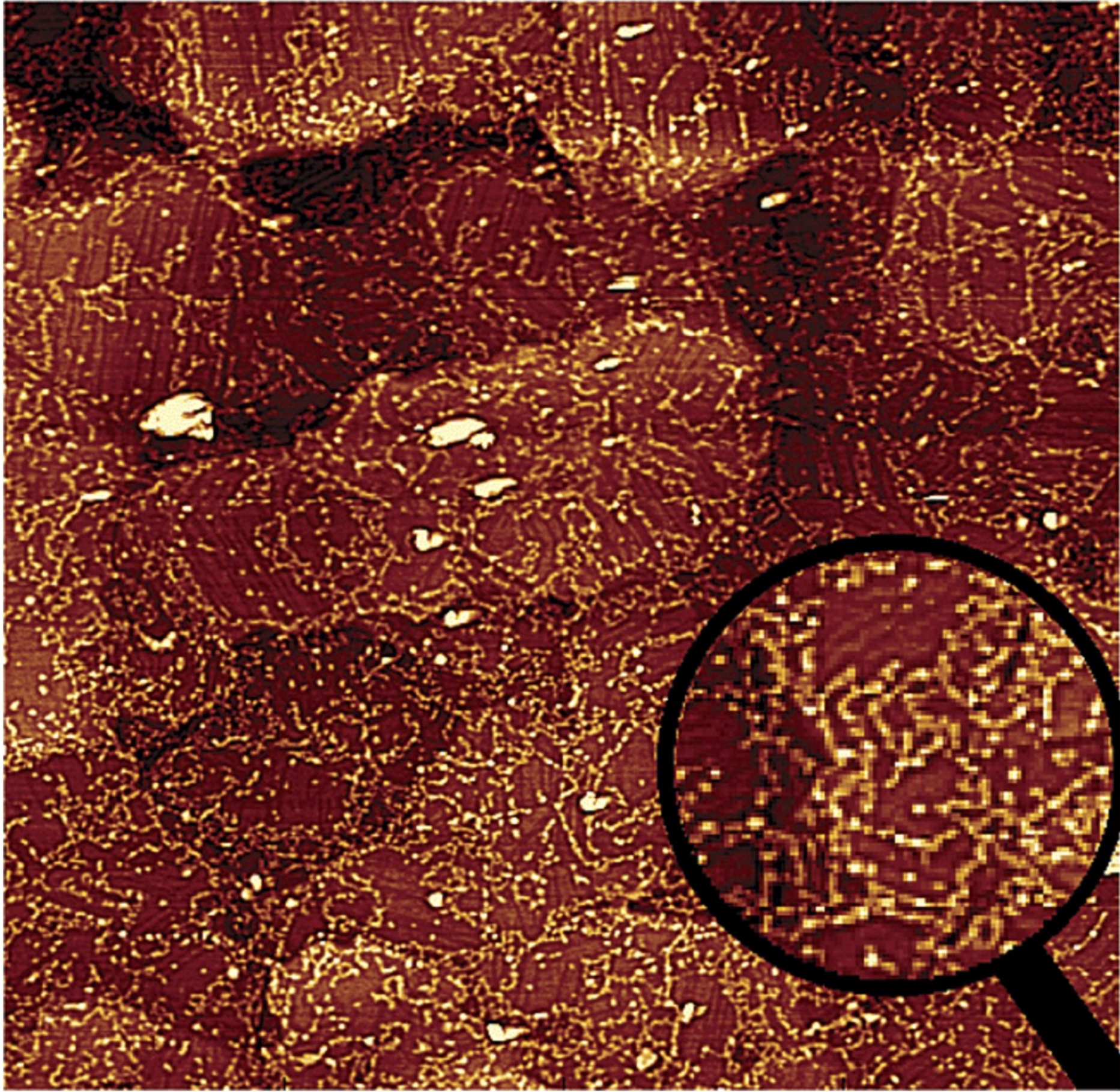
We will cover **atomic force microscopy (AFM)** later – go-to method for measuring nanoscale height

# Upcoming Tool Next Week: Atomic Force Microscopy

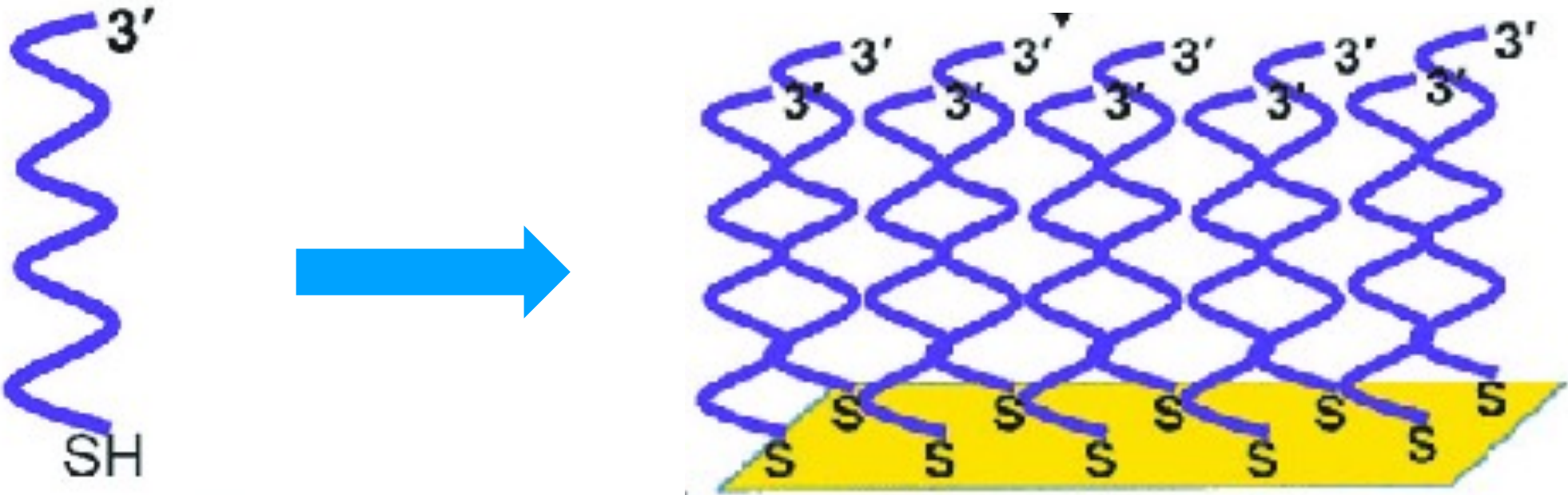


AFM image of Au (111) surface (terraces observed)

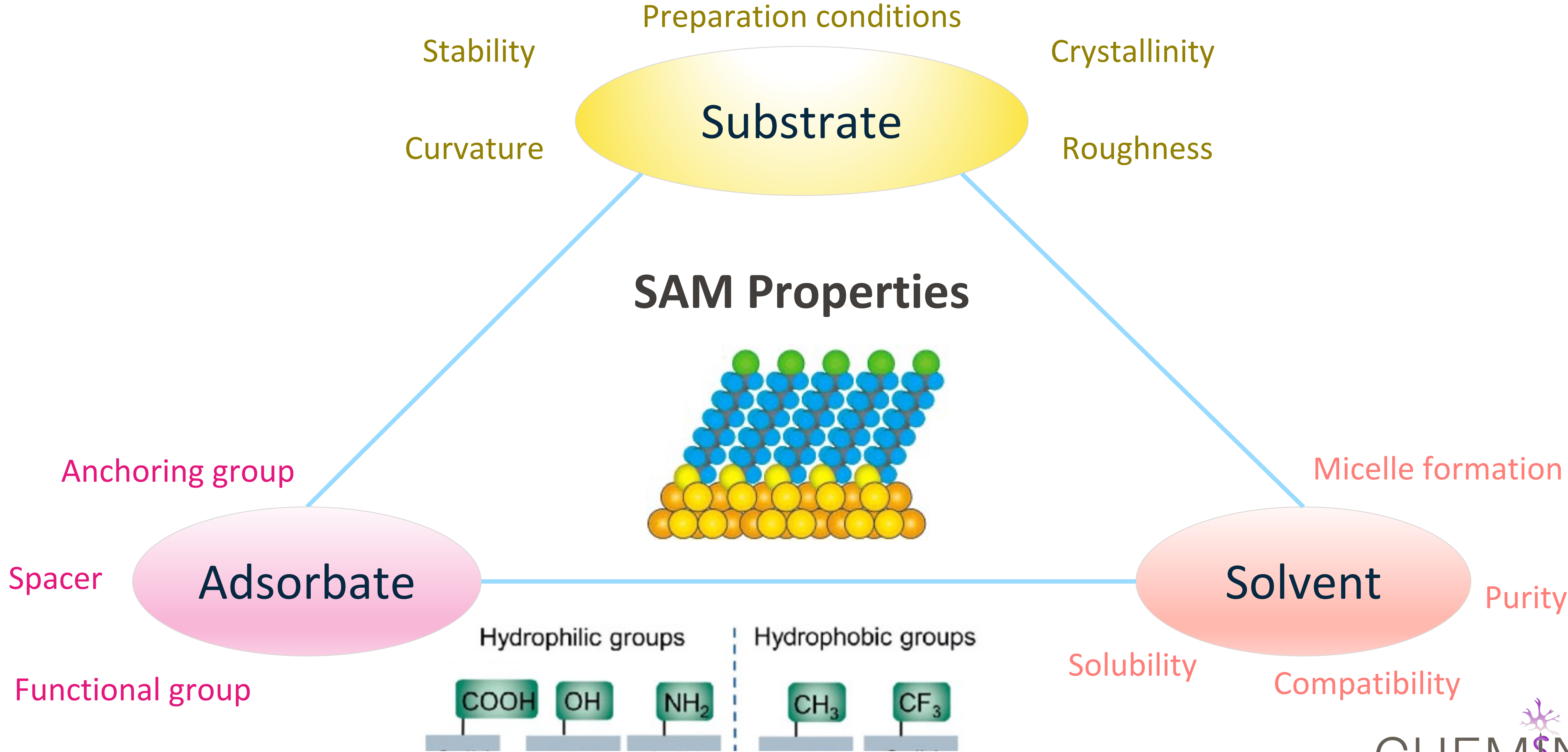
Surface exposed to a solution of oligonucleotides (25-base thiolated DNA)



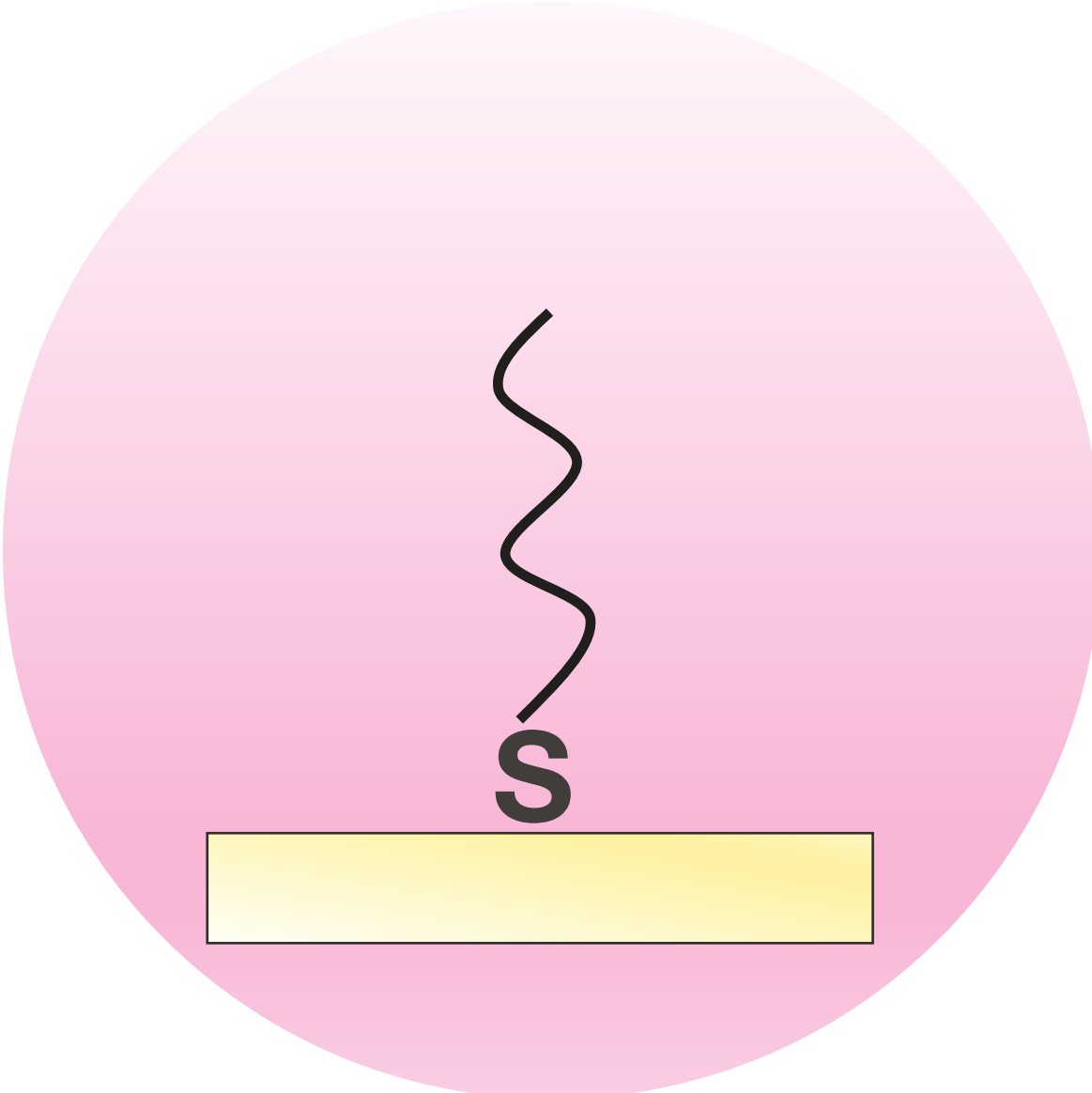
2 μm



# Recap from Lesson 8: Factors Governing Self Assembly

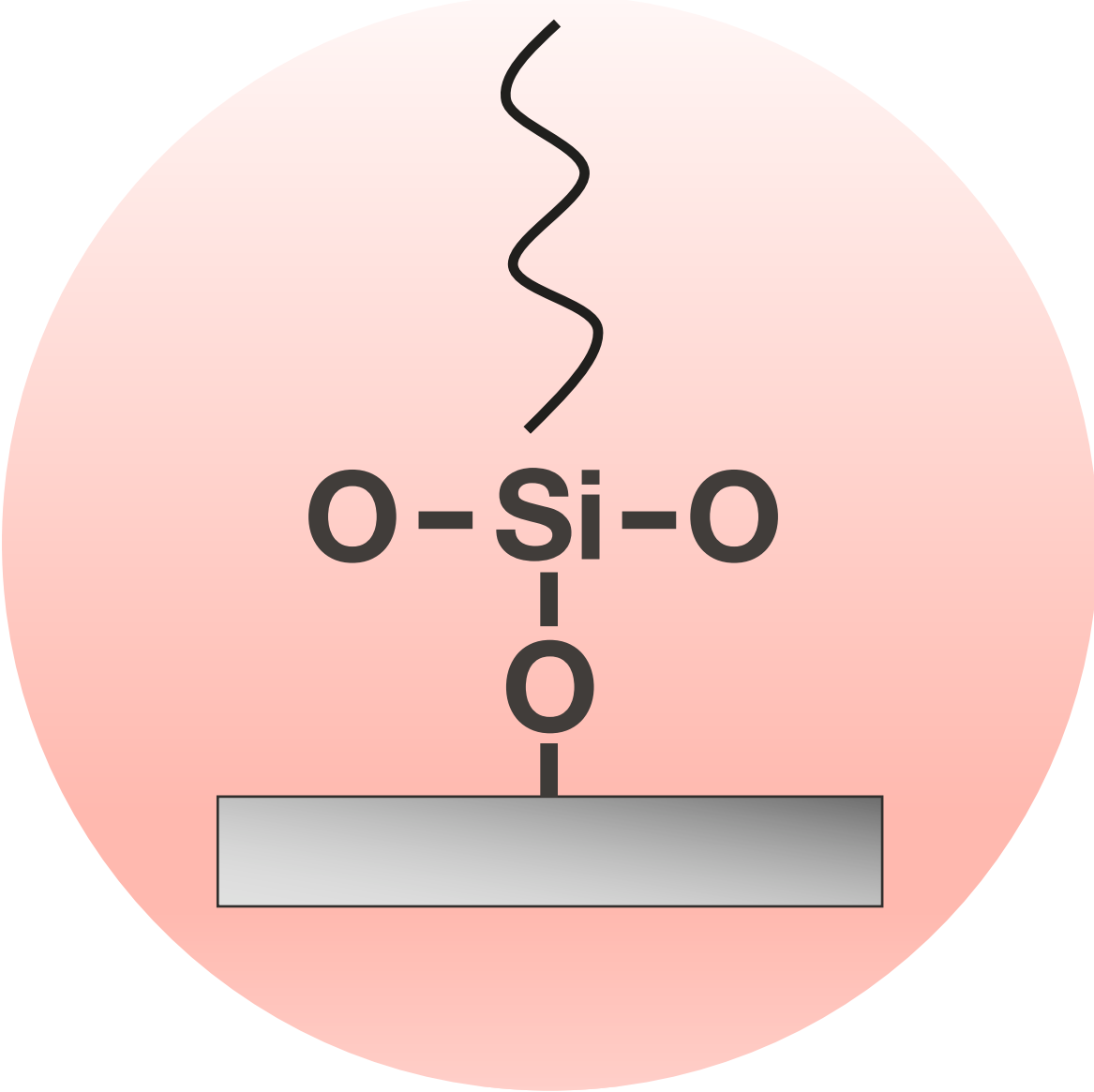


# Recap from Lesson 8: Surface Chemistry Strategies

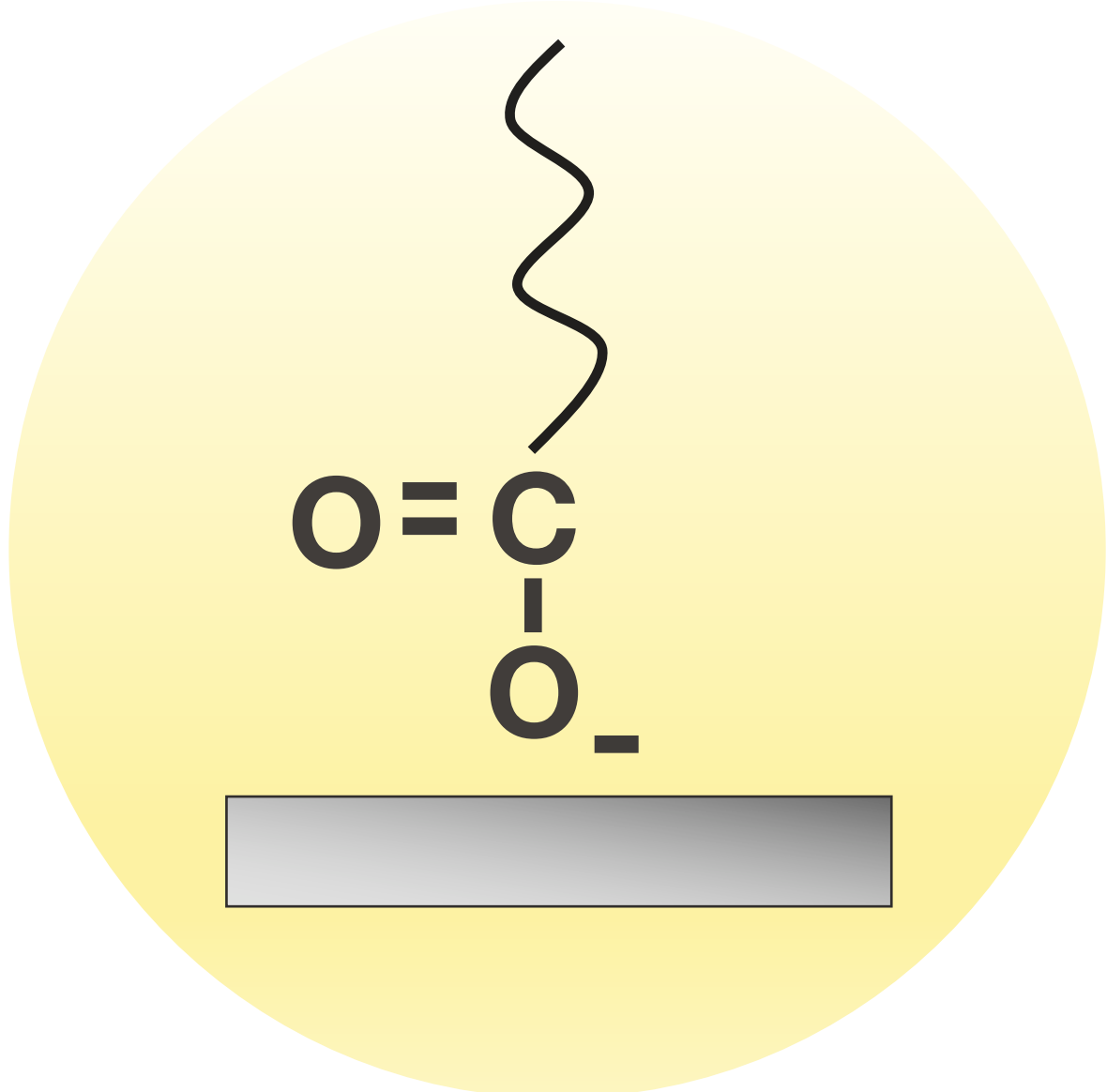


**Thiol-based chemistry**  
**Metallic surfaces**

**Surface chemistry**



**Silane, Phosphonates**  
**Metal oxide surfaces**



**Carboxylic acids**  
**Metal oxide surfaces**

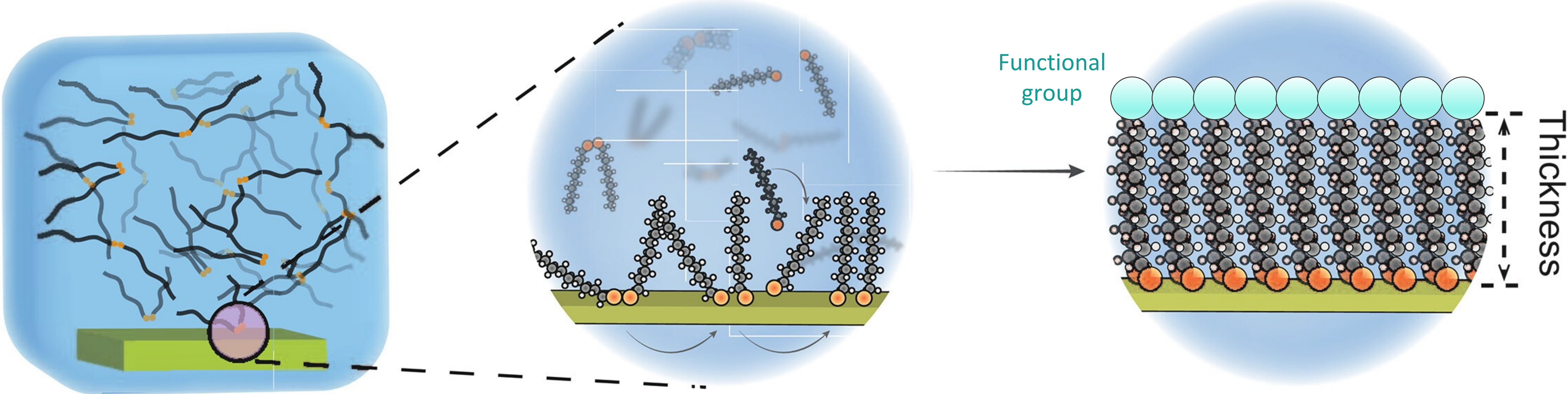
# Outline of Lesson 9

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- Coupling chemistries to surface functional groups
- *Why where* we immobilize matters – the need for patterning
- Four patterning methods for microscale – nanoscale patterning of surfaces
- Motivate the need for polymer-based surface chemistries – protein fouling on surfaces
- How do proteins stick to surfaces of different properties – thermodynamics
- Polymer grafting to tackle nonspecific binding



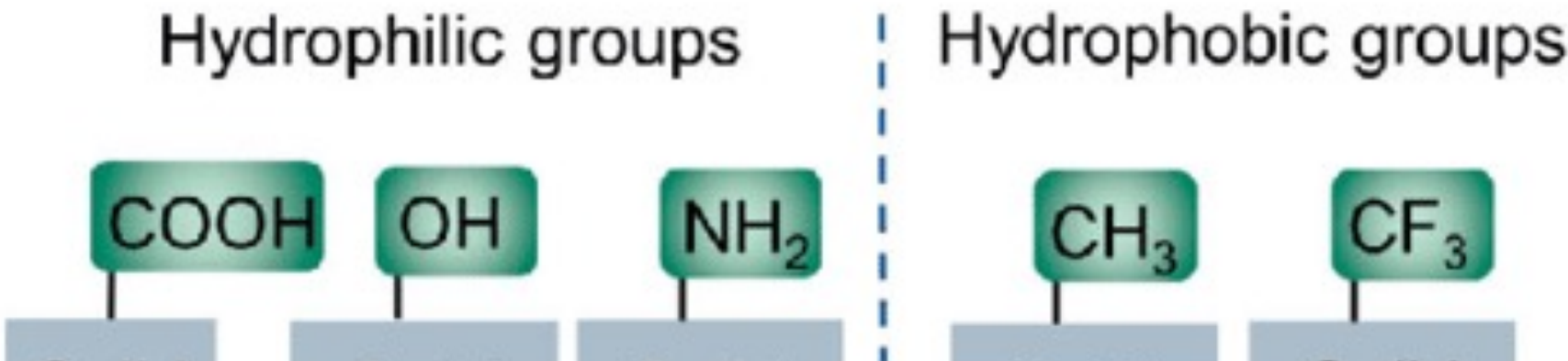
# Assembled SAMs Serve As Interface for Molecular Immobilization



Adsorption

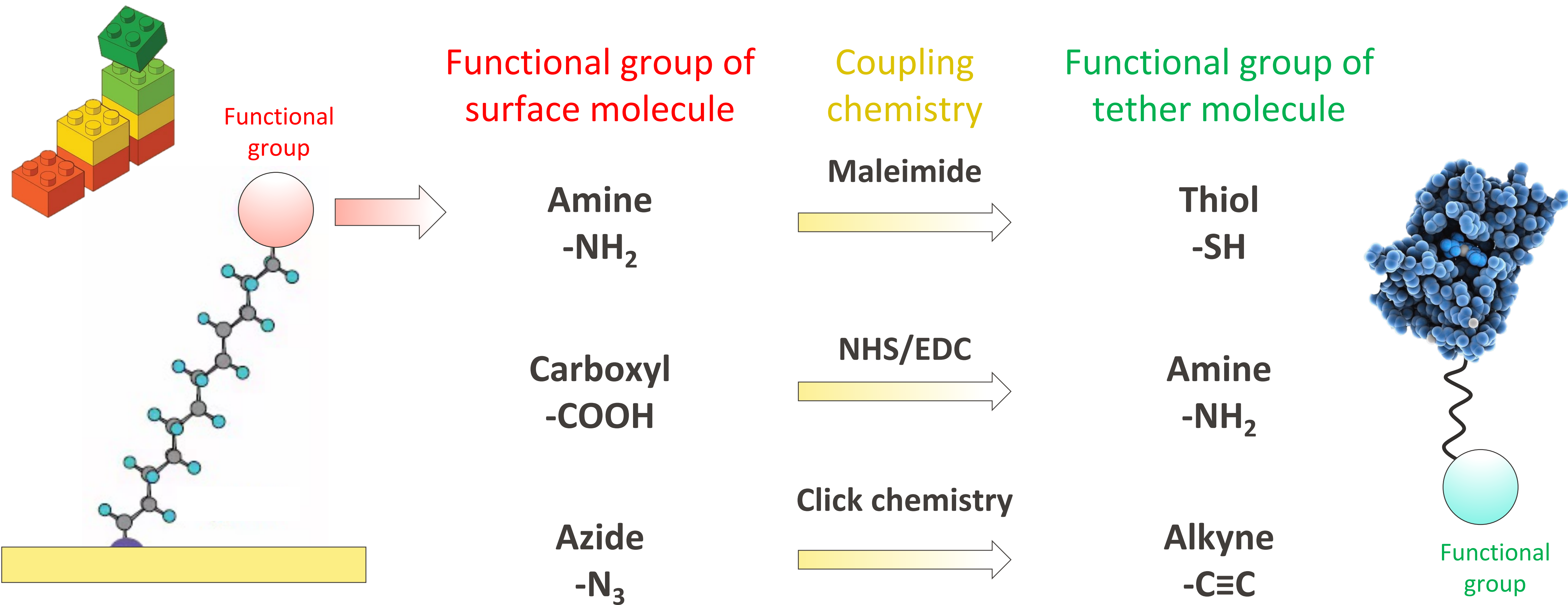
Self-assembled

Functional groups can tune surface properties:

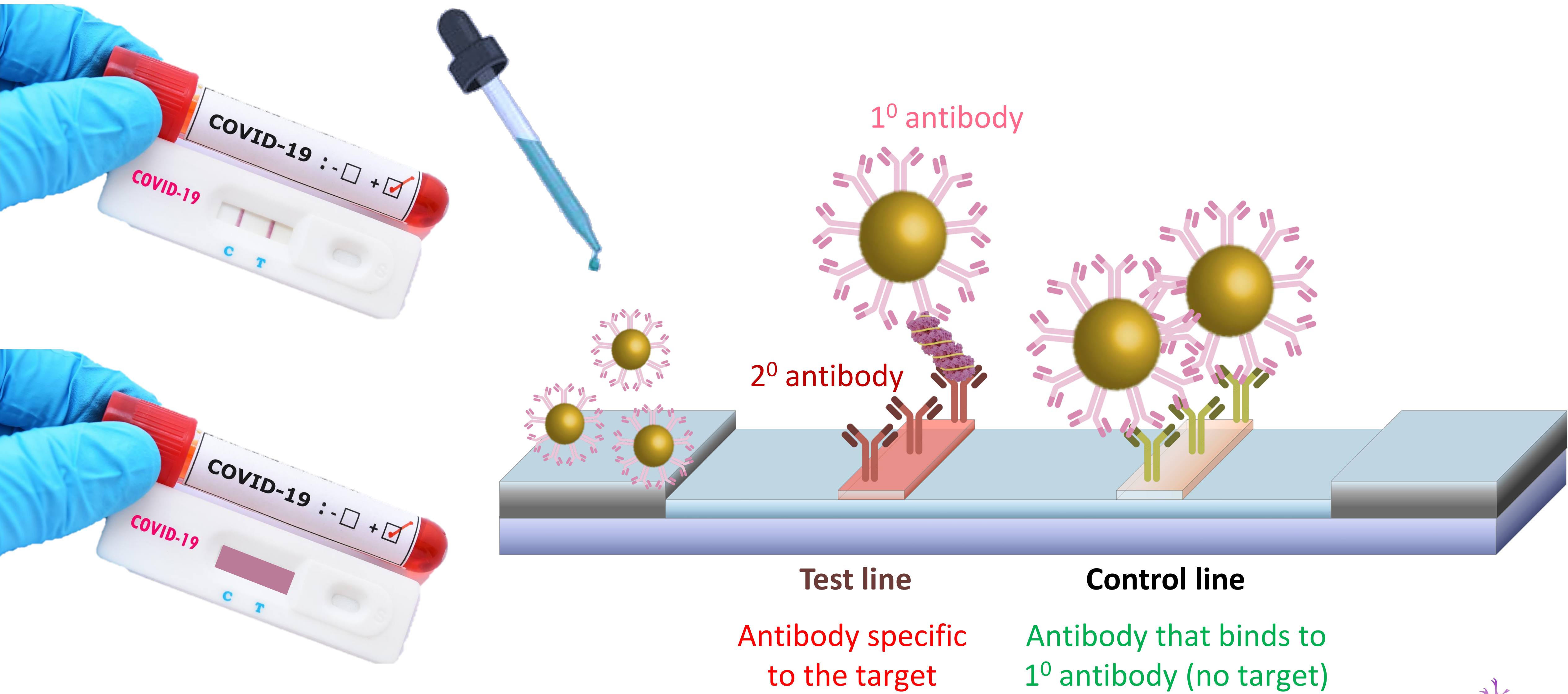


In addition, they enable coupling of molecules to surfaces

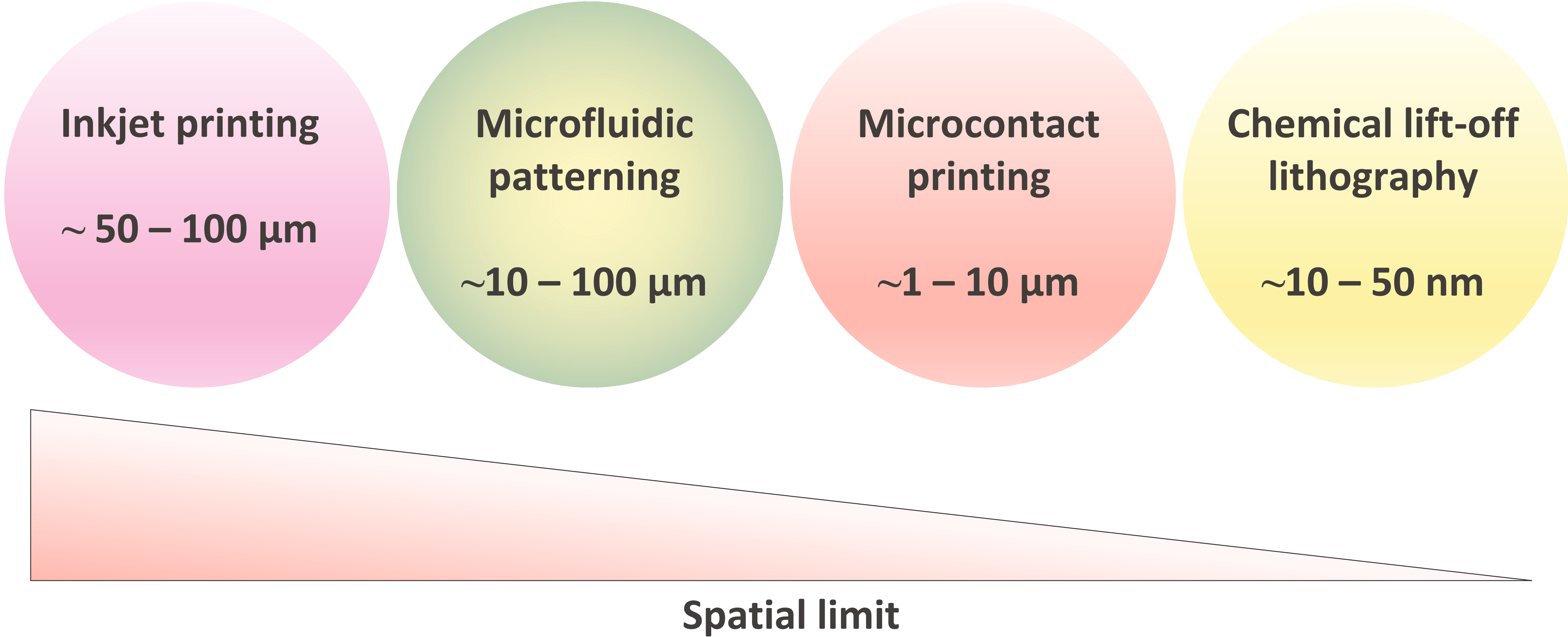
# Assembled SAMs Serve As Interface for Molecular Immobilization



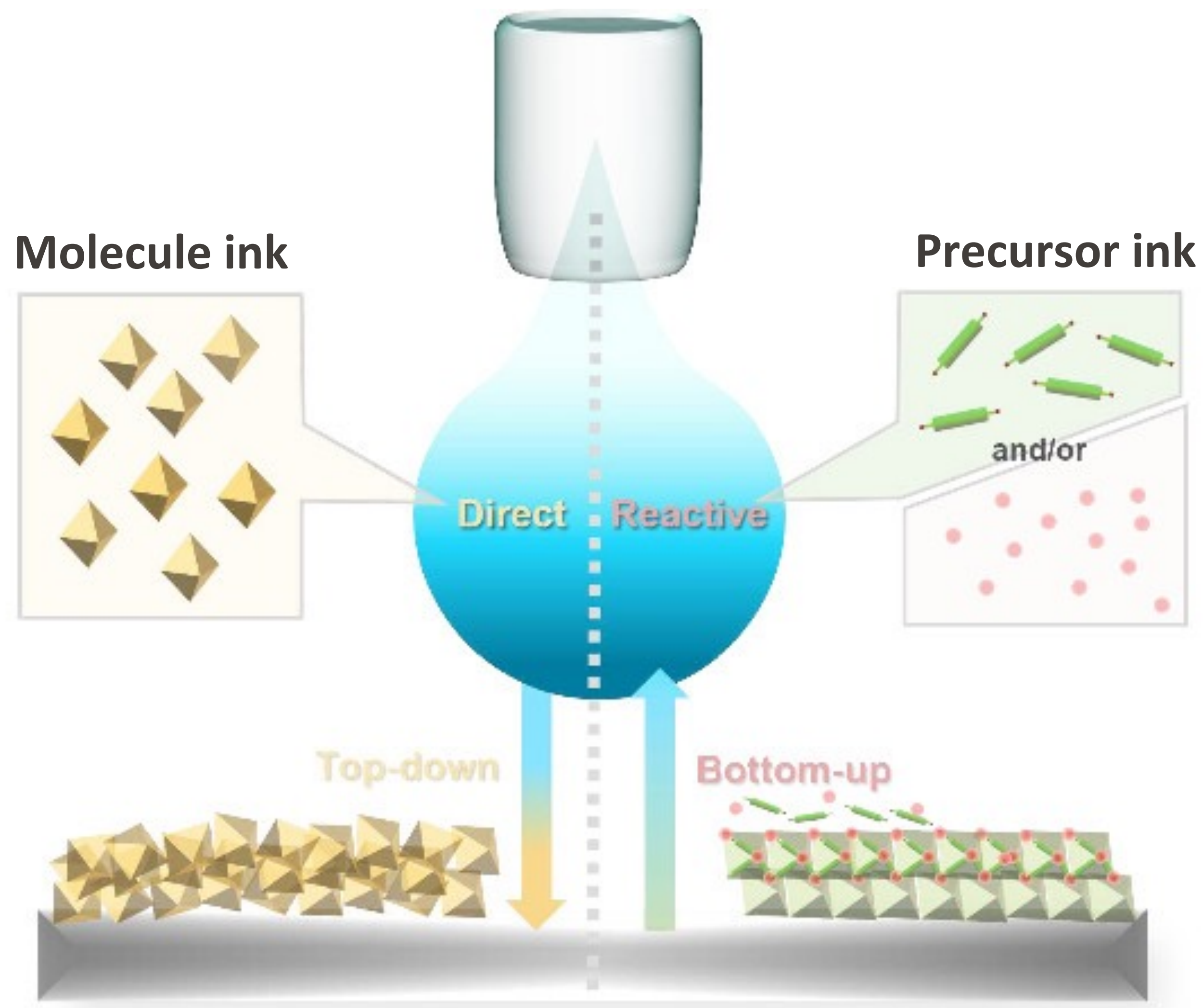
# When We Tether Biomolecules, *Where* We Immobilize Matters



# Molecules Stick Everywhere – Patterning for Spatial Control



# Inkjet Printing Large Areas with Controlled Volume Droplets

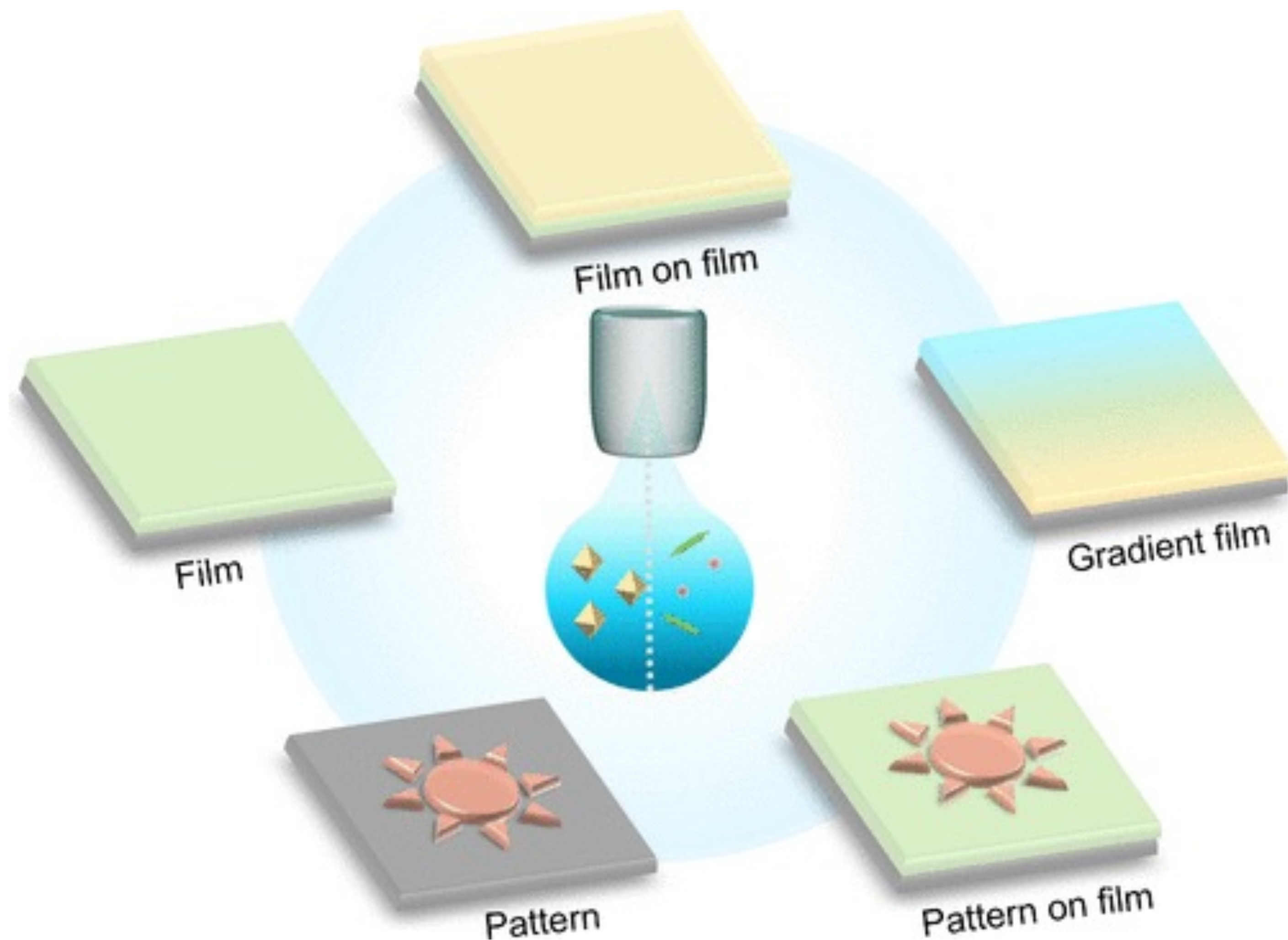


- Nozzle size
- Droplet velocity

- Viscosity
- Surface tension
- Particle size
- Density

- Wetting ability
- Adhesion

# Diversity of Inkjet Patterning on Surfaces

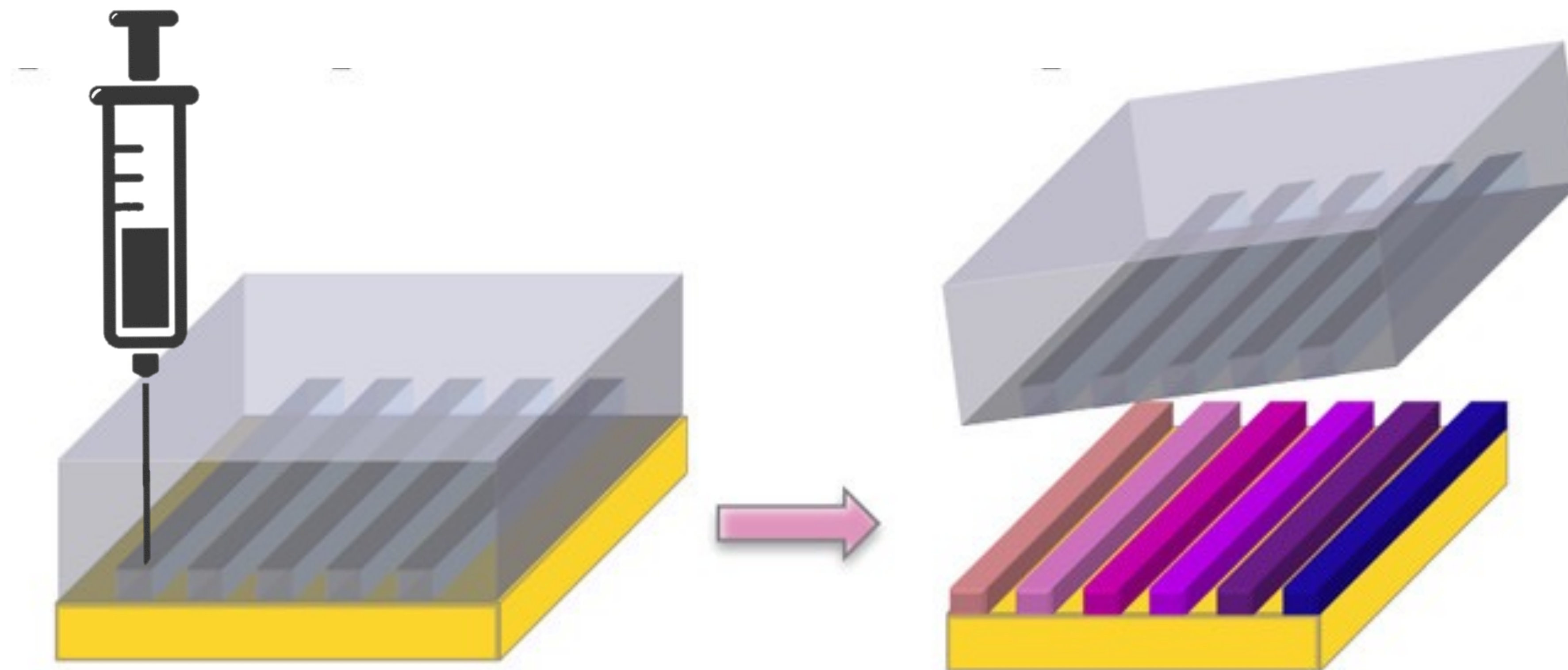


Multiple chemistries can be printed

High-throughput, flexible, and  
computer controlled

Appropriate for larger patterns  
(~50 – 100  $\mu\text{m}$  for alkanethiols)

# Microfluidic Patterning (Polymer Channels)

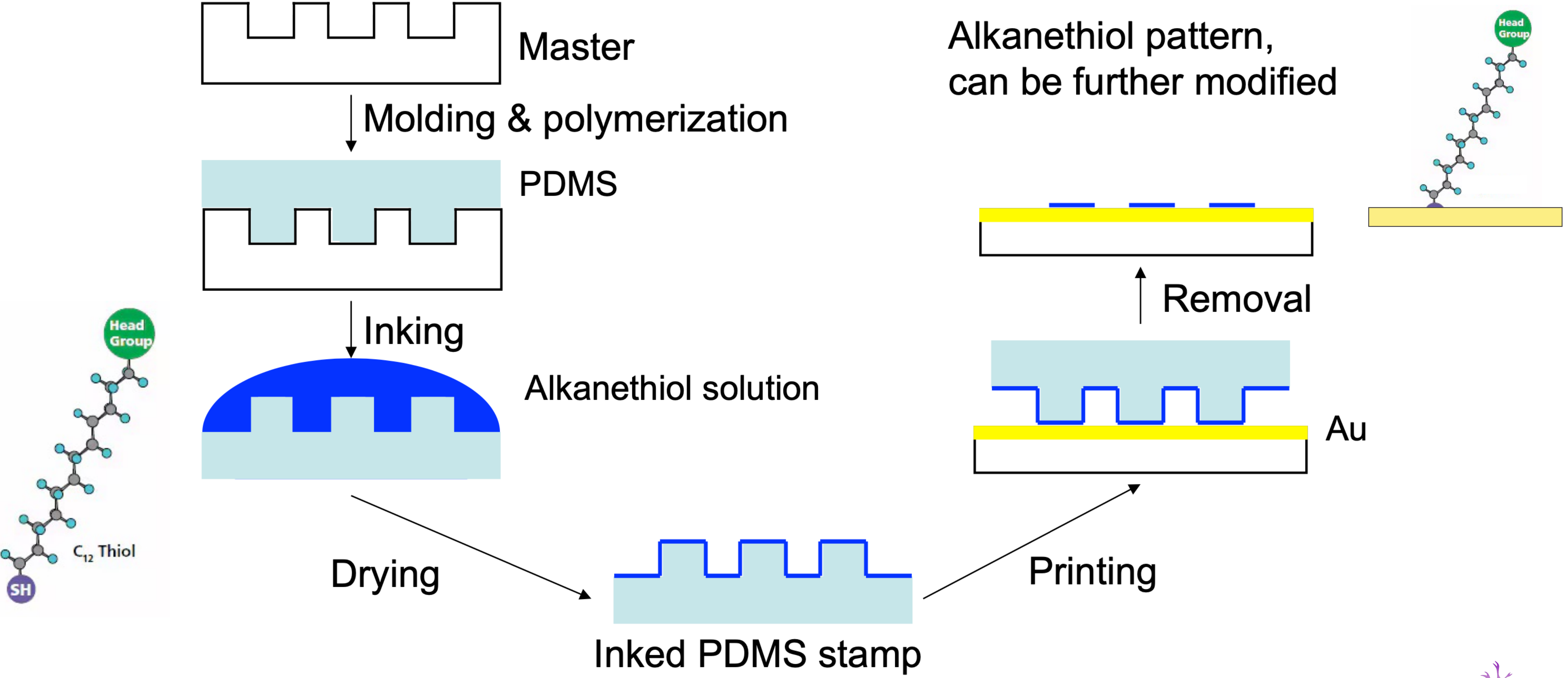


Inject different molecules in each channel

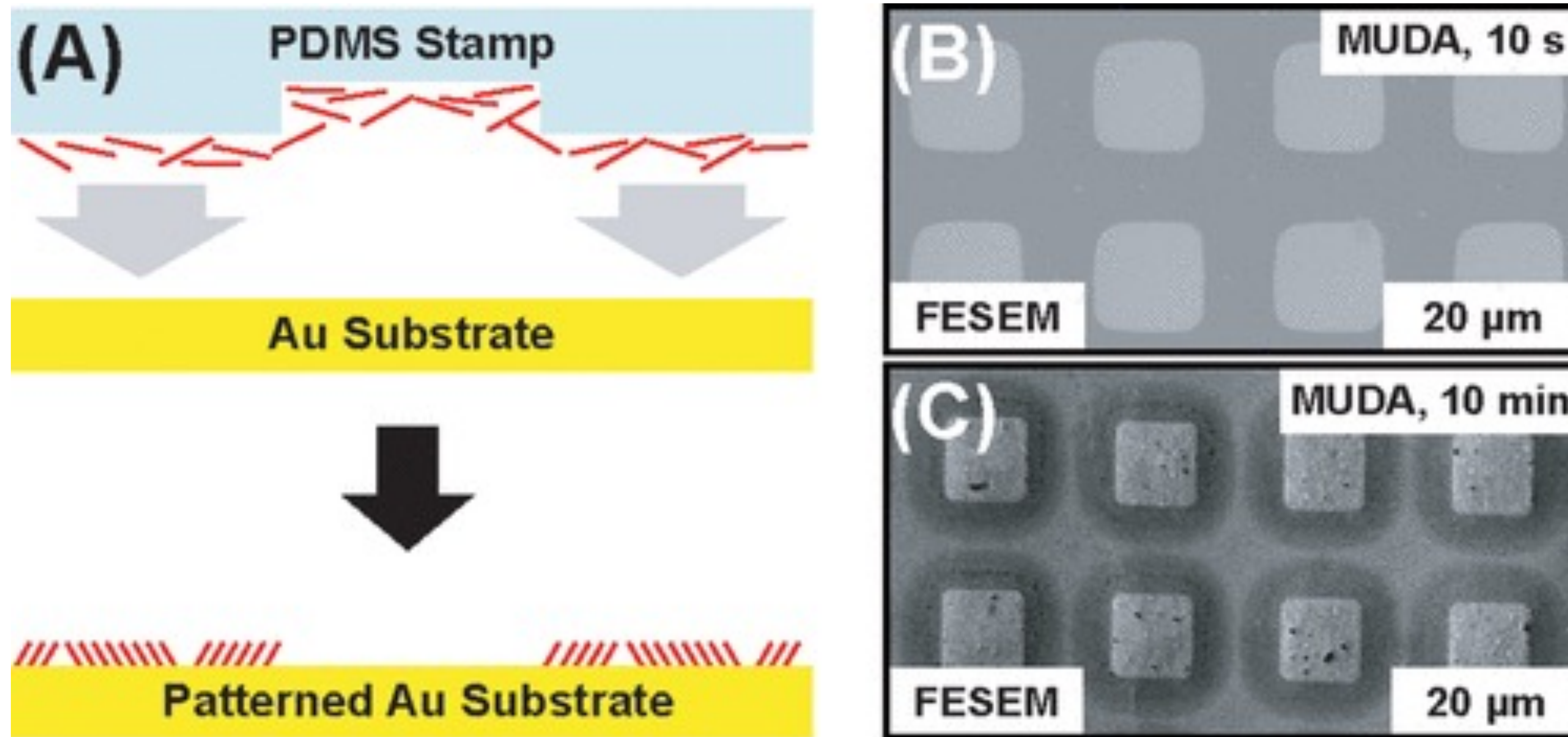
Pattern different molecules

Spatial resolution:  $\sim 10\text{--}100\ \mu\text{m}$ , limited by channel width and molecular diffusion

# Microcontact Printing – Transferring Molecules to Surfaces



# Microcontact Printing – Resolution Limit of ~100 nm

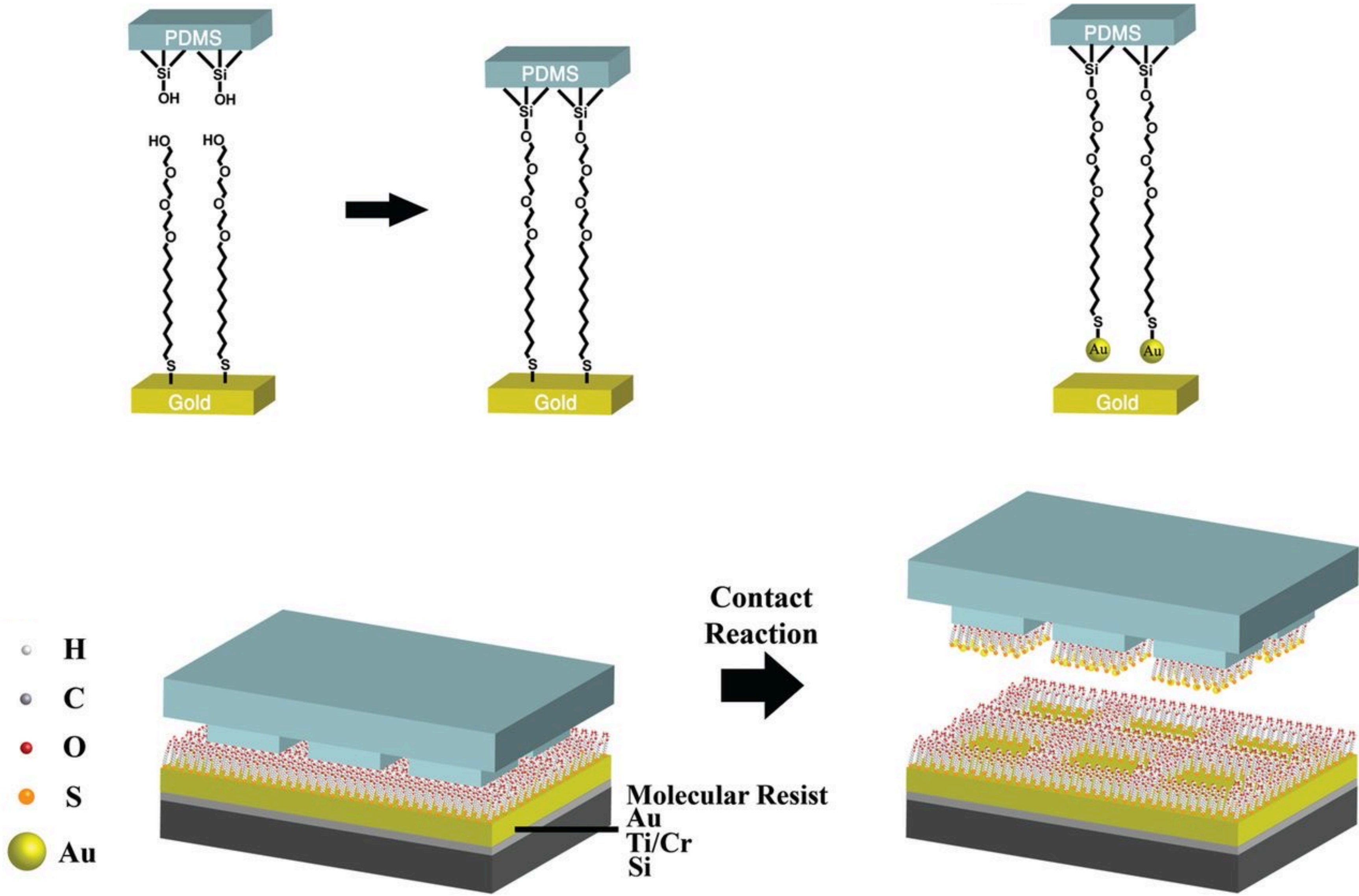


Success of printing limited by chemistries and compatibilities of inks, stamps, and substrates

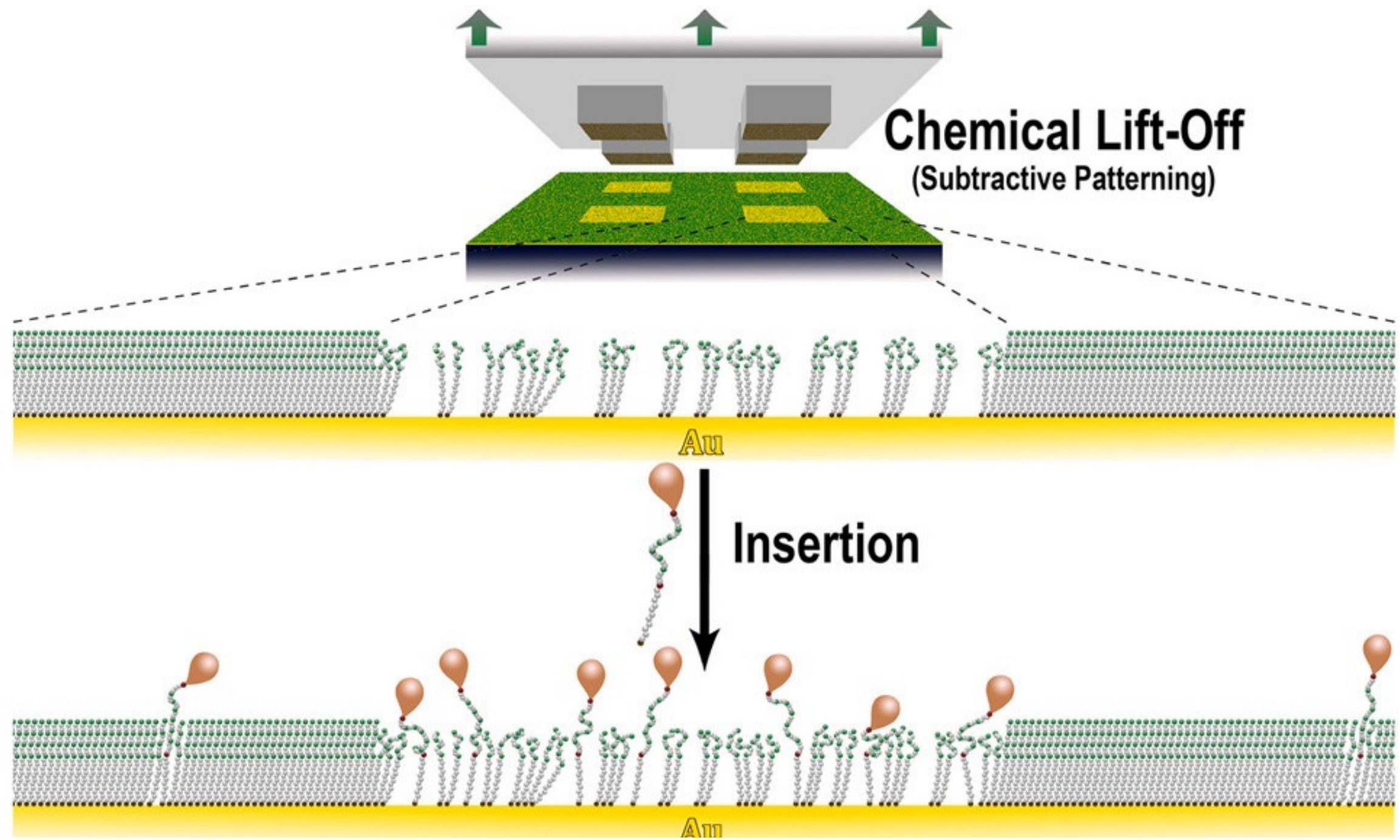
Lateral diffusion of inked molecules reduce pattern fidelity (sharpness of pattern without blurring)

Realistic resolution ~1–10 μm

# Chemical Lift-Off Lithography (CLL) – Patterning Smaller Features



# Chemical Lift-Off Lithography (CLL) – Patterning Smaller Features



PDMS stamp with “UCLA” characters as positive features and “CNSI” as negative features

Fluorescent molecule inserted into patterned regions



250 μm



In the gaps formed from lifting off molecules, new molecules can be inserted patterning

Realistic resolution: ~ 50-200 nm

Best possible limit: ~ 10–20 nm

# Key Takeaways

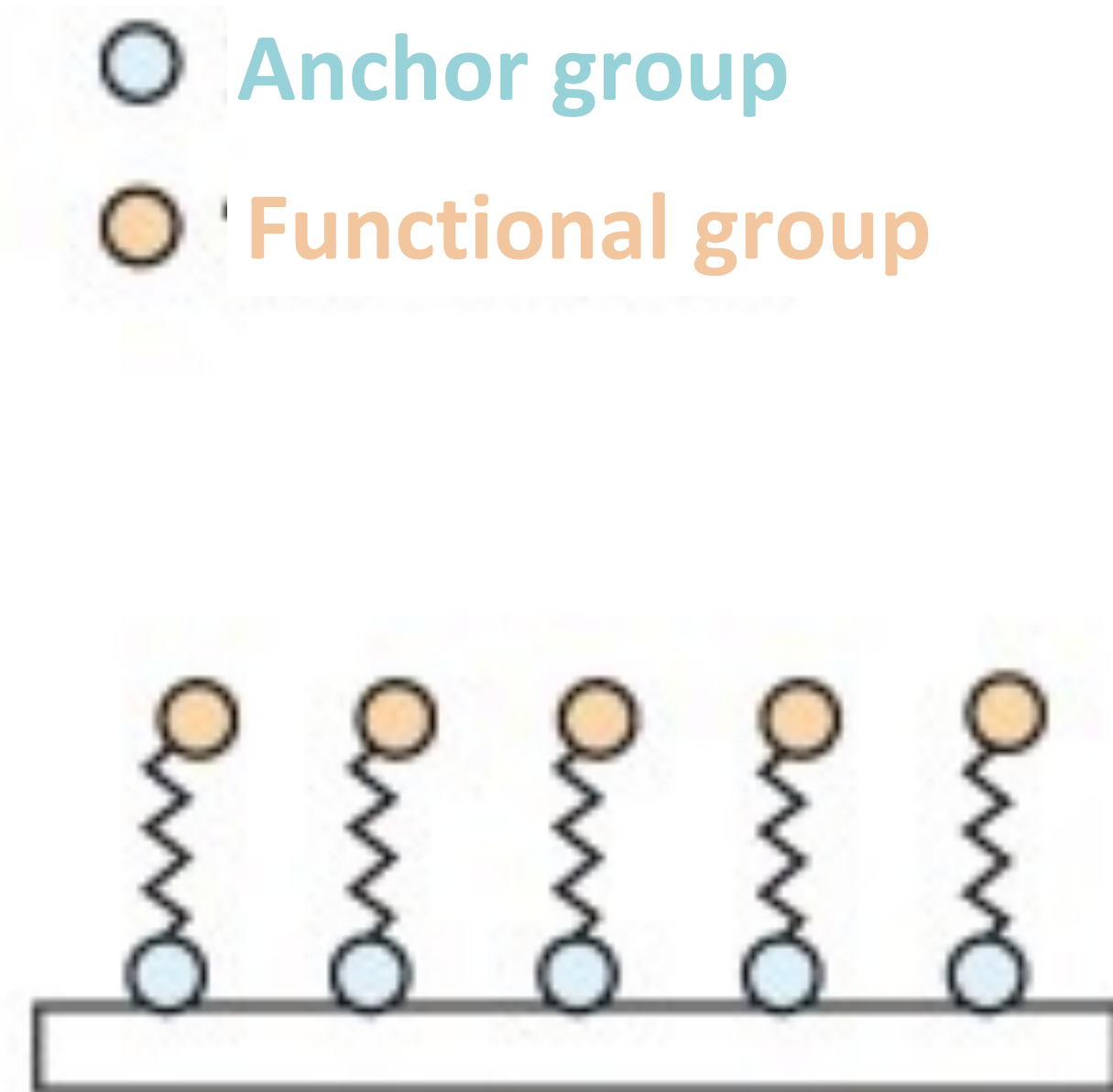
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- Typically, multiple characterization techniques are used for surface analysis
- Once assembled, the functional groups of SAMs can be coupled to molecules
  - Especially for biosensors, *where* we pattern molecules, matters
- There are various techniques to pattern SAMs from the microscale to nanoscale (we covered four examples)

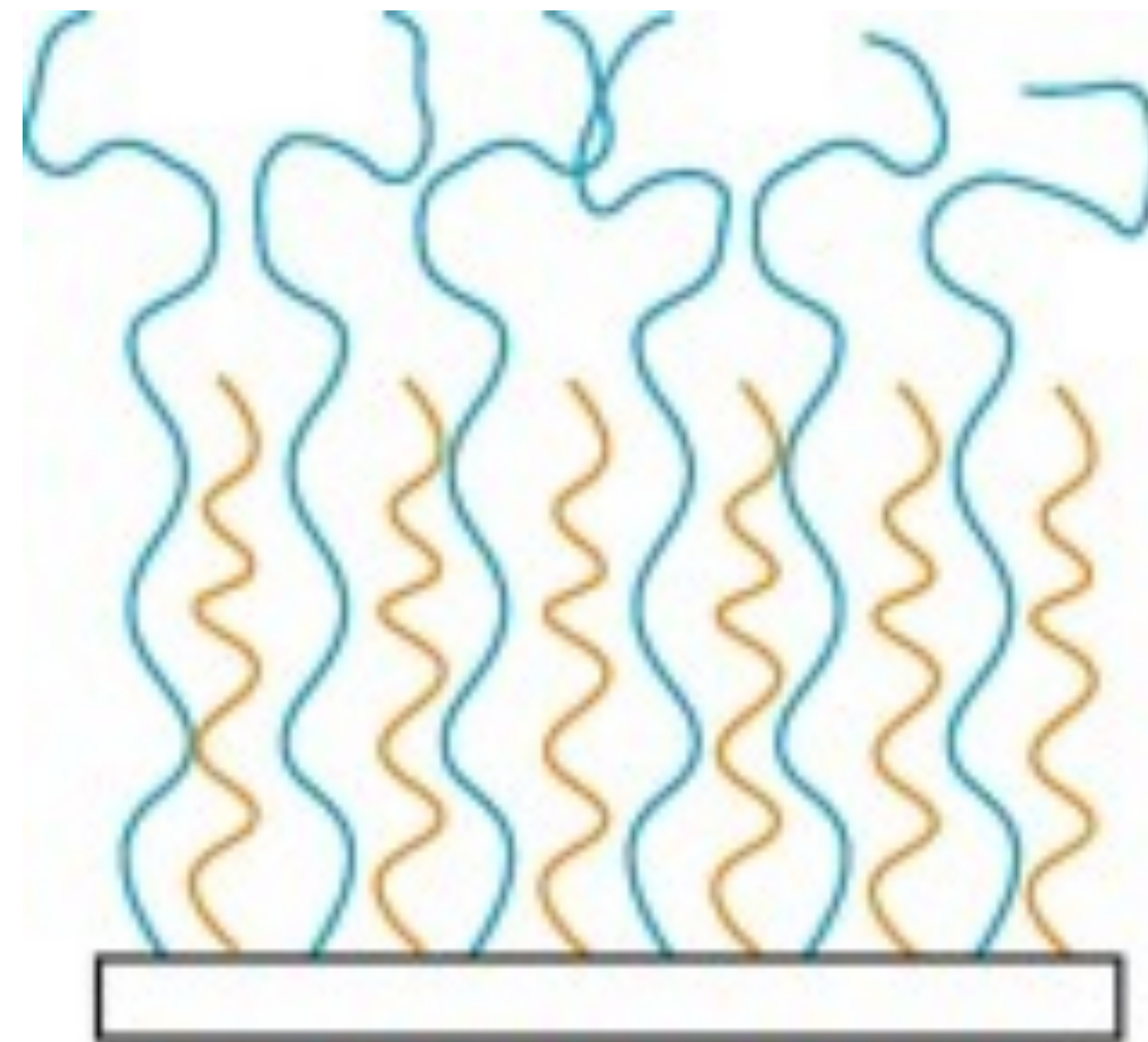
# Why Move Beyond SAMs?

SAMs are thin + ordered, but limited – stability issues, limited thickness/hydration

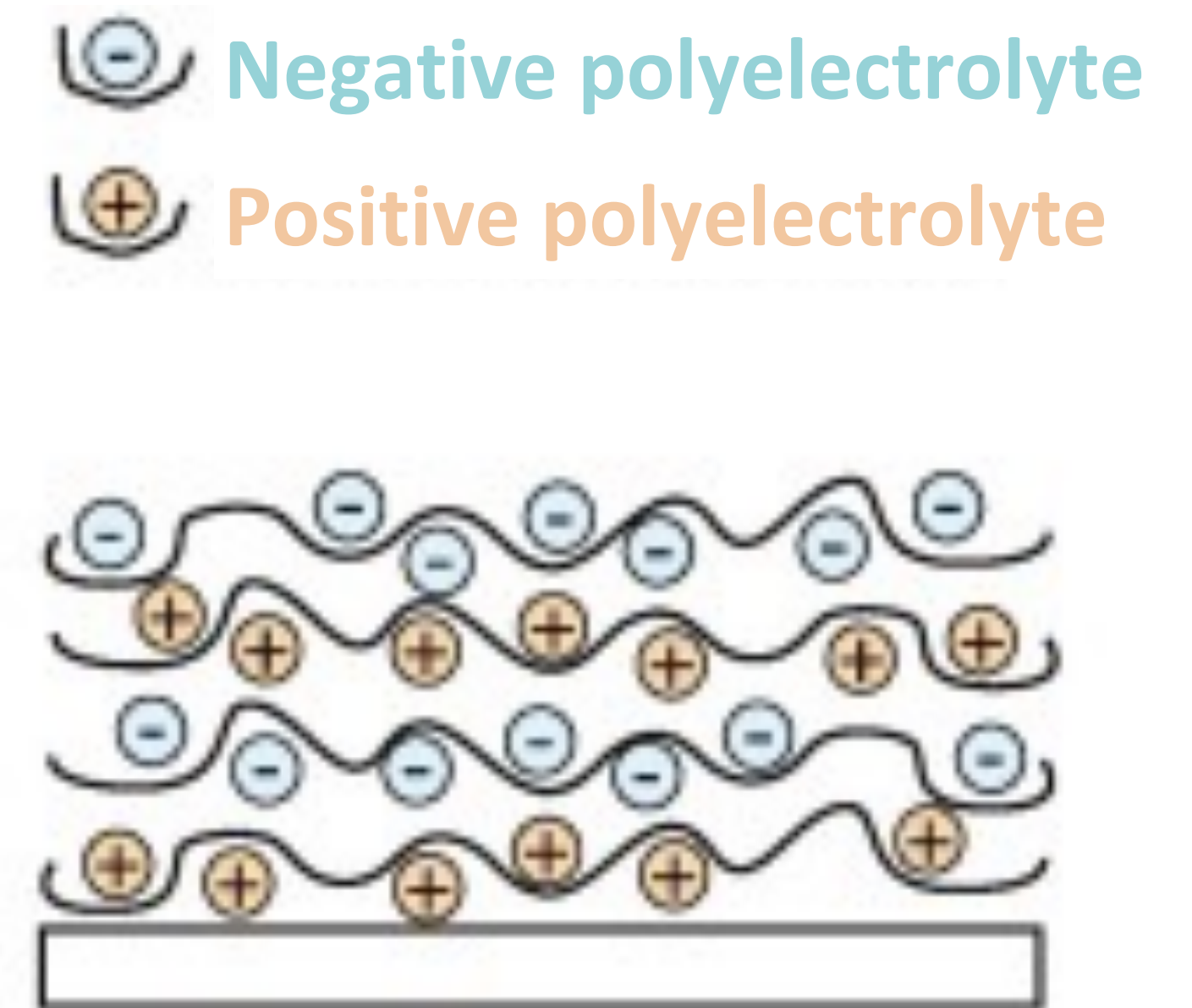
Need for thicker, tunable, hydrated surfaces especially at biological interfaces



SAM



Polymer brush (binary)



Polyelectrolyte multilayer

- Anchor group
- Functional group

- ⊖ Negative polyelectrolyte
- ⊕ Positive polyelectrolyte

# Applications Driving Polymer-Coated Surfaces

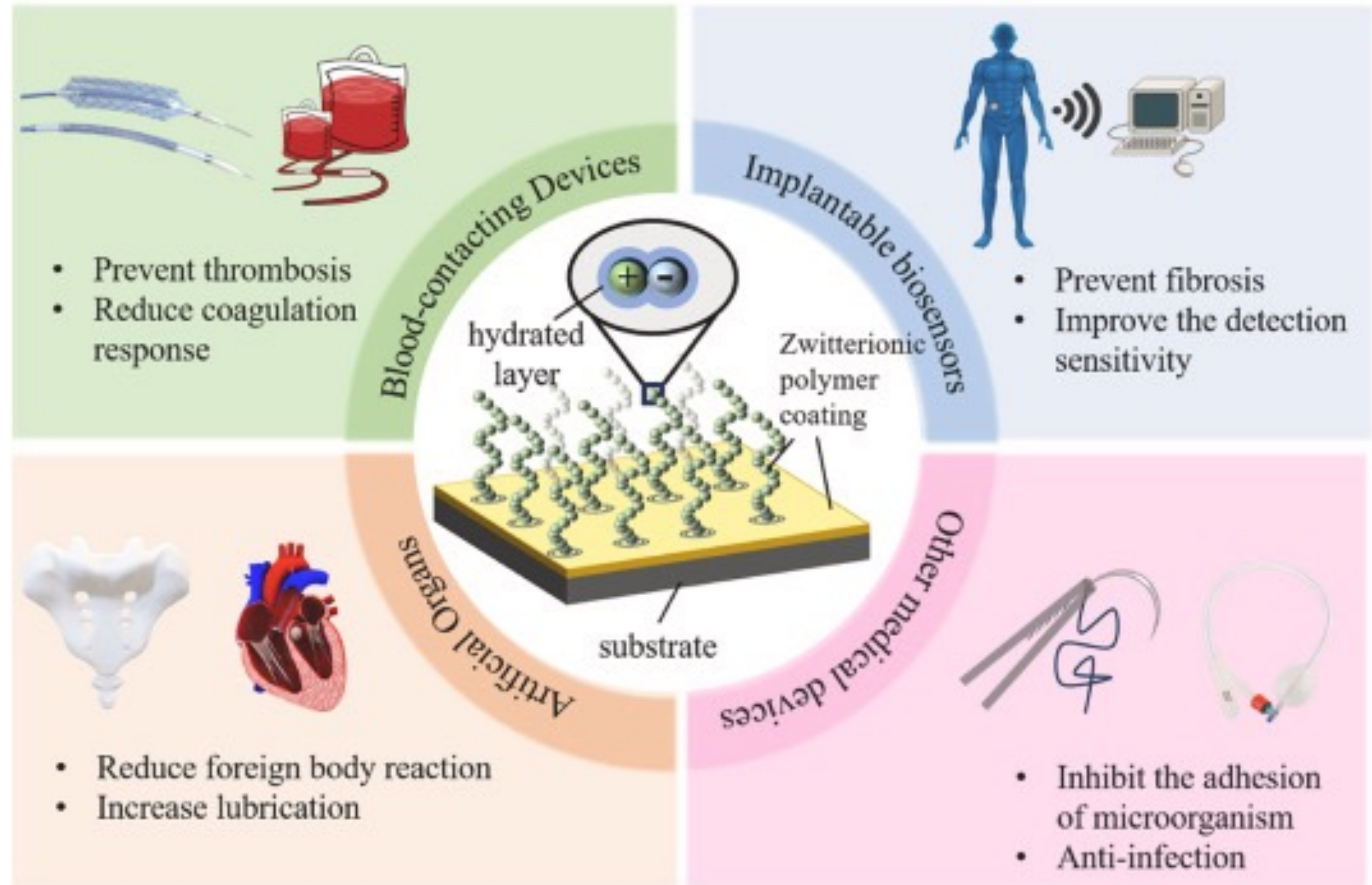
Biosensors

Medical implants

Microfluidics

Lab-on-chip

Antifouling coatings



# Where are Polymer-Coated Layers Most Useful in Surface Science?

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Nonspecific protein adsorption in complex environments/biofluids

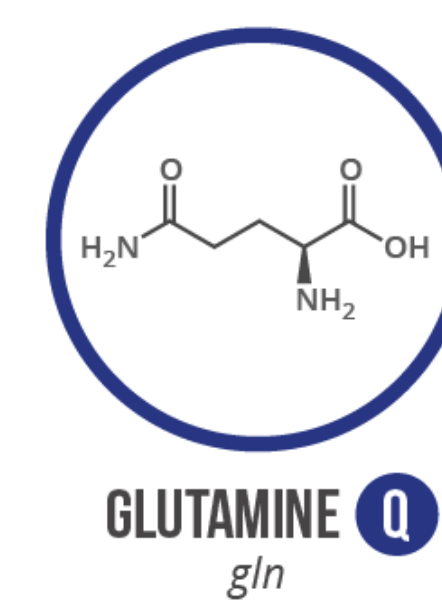
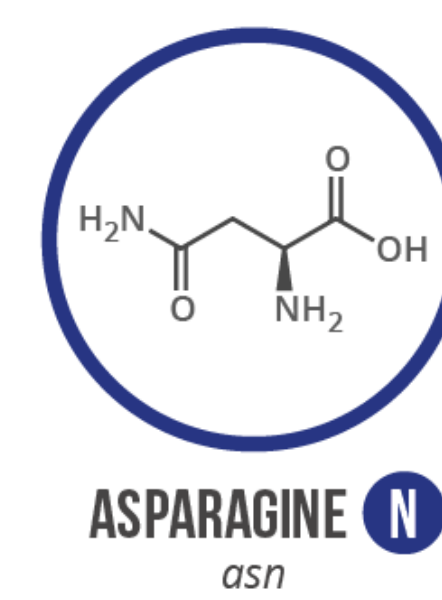
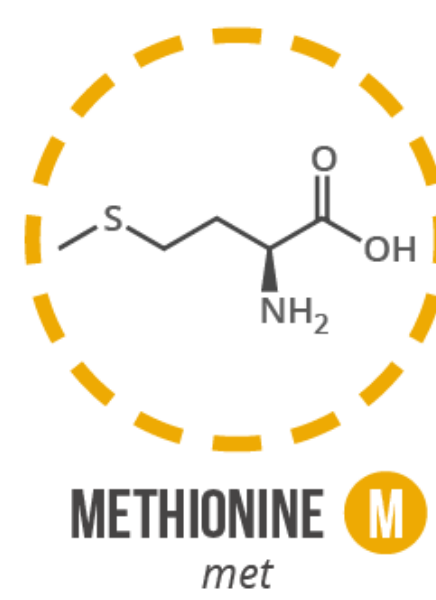
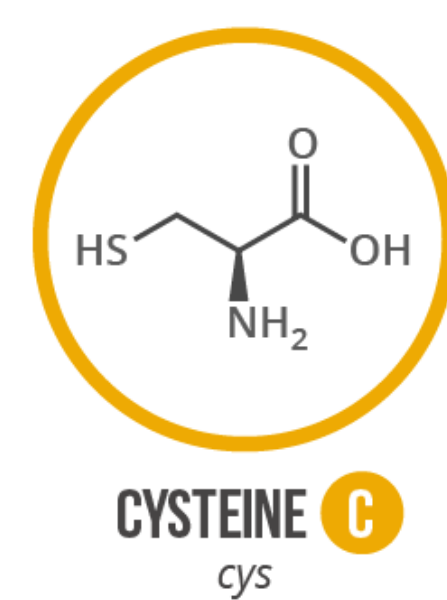
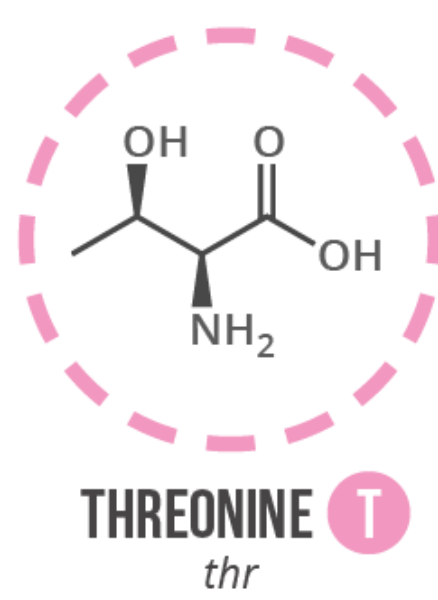
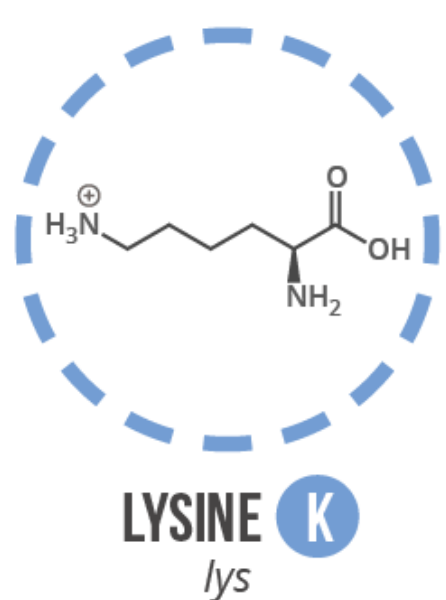
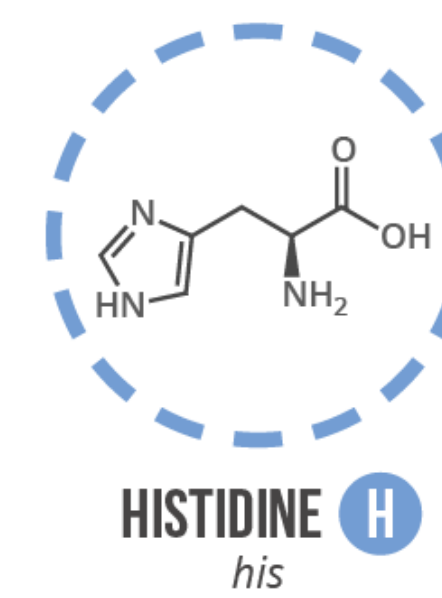
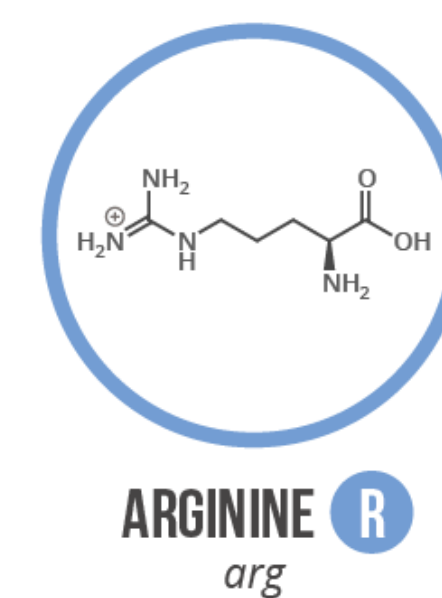
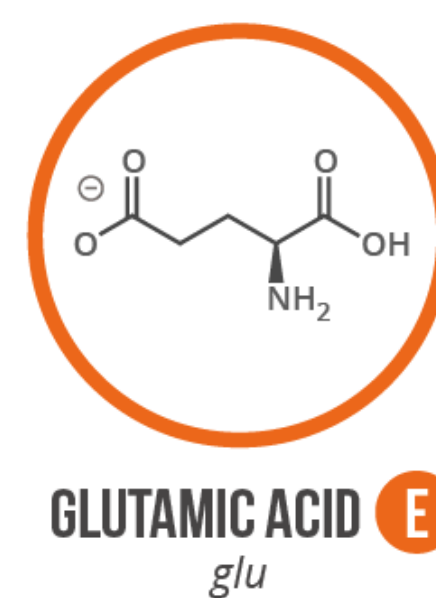
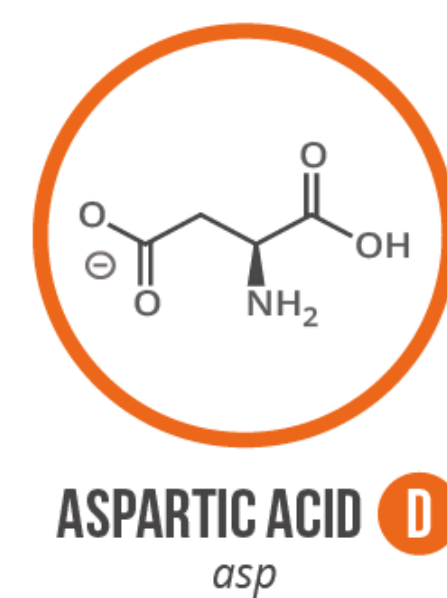
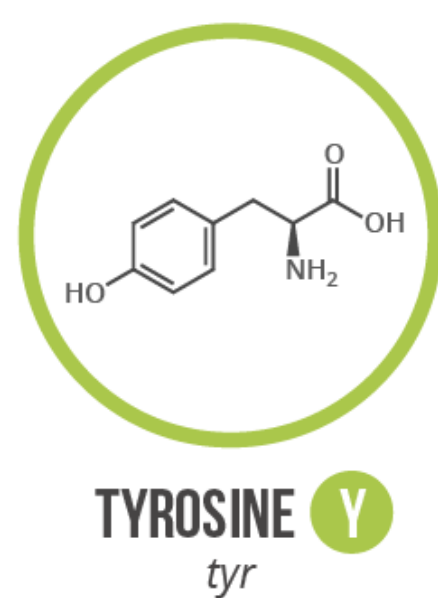
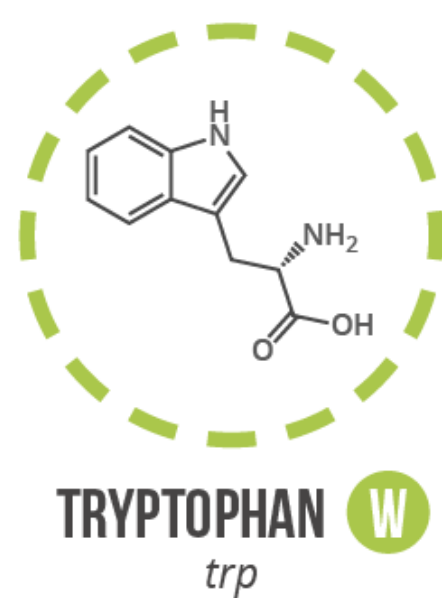
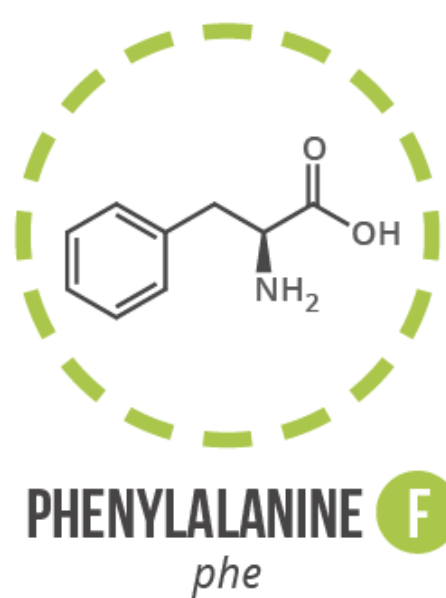
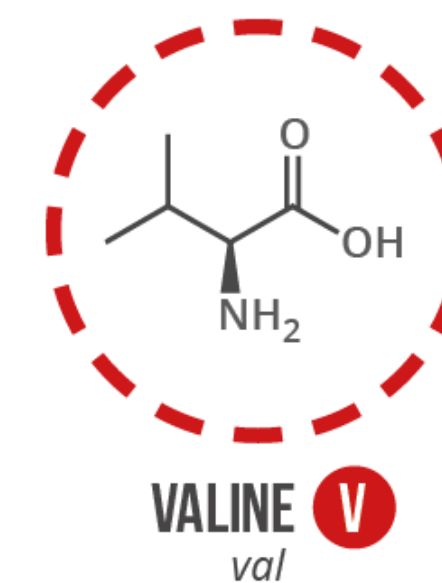
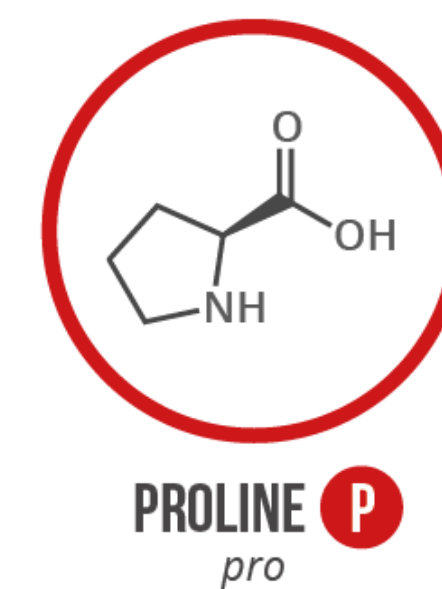
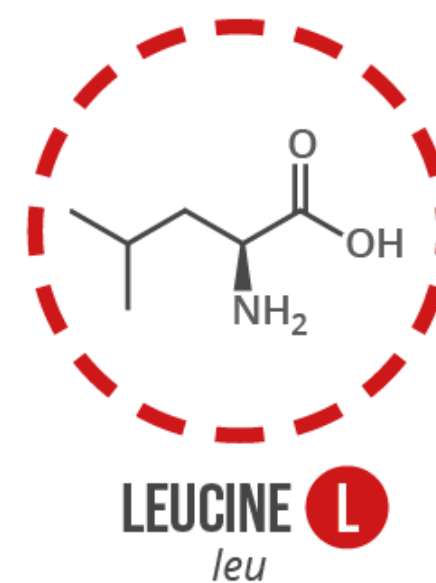
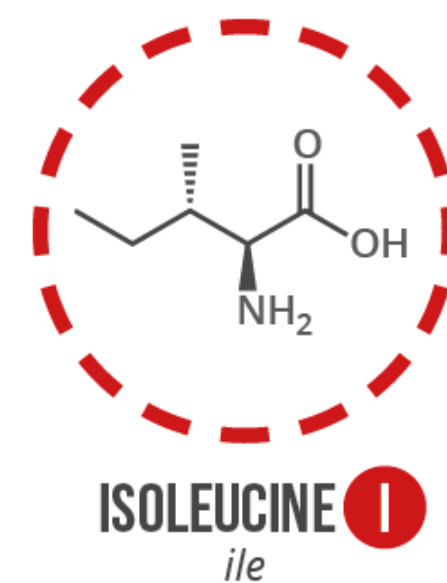
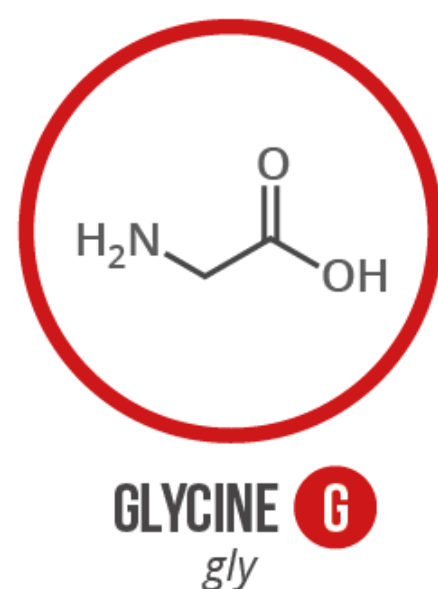
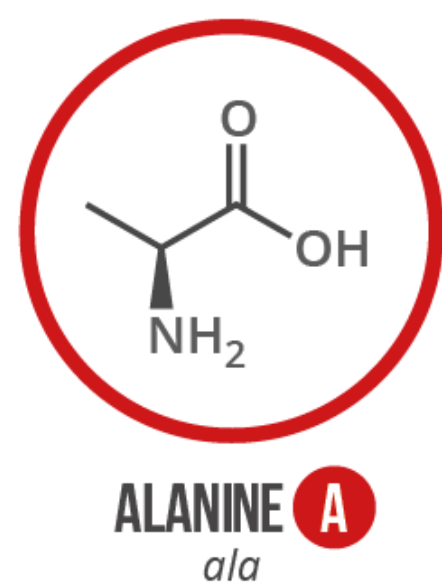
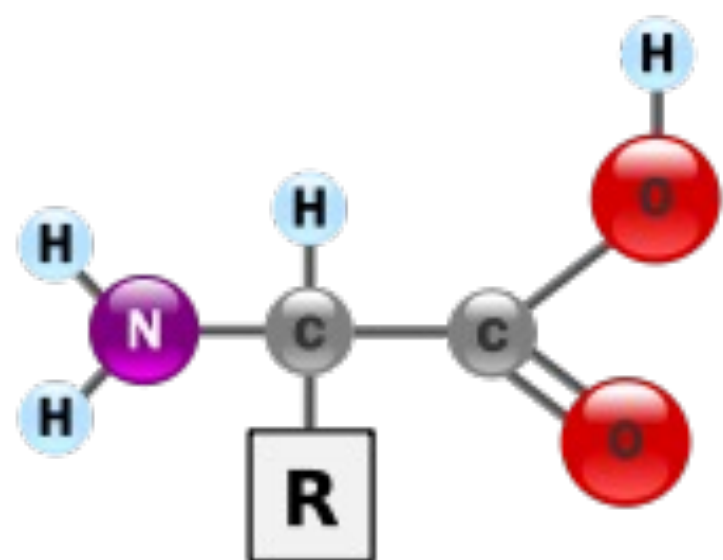


**Surface**

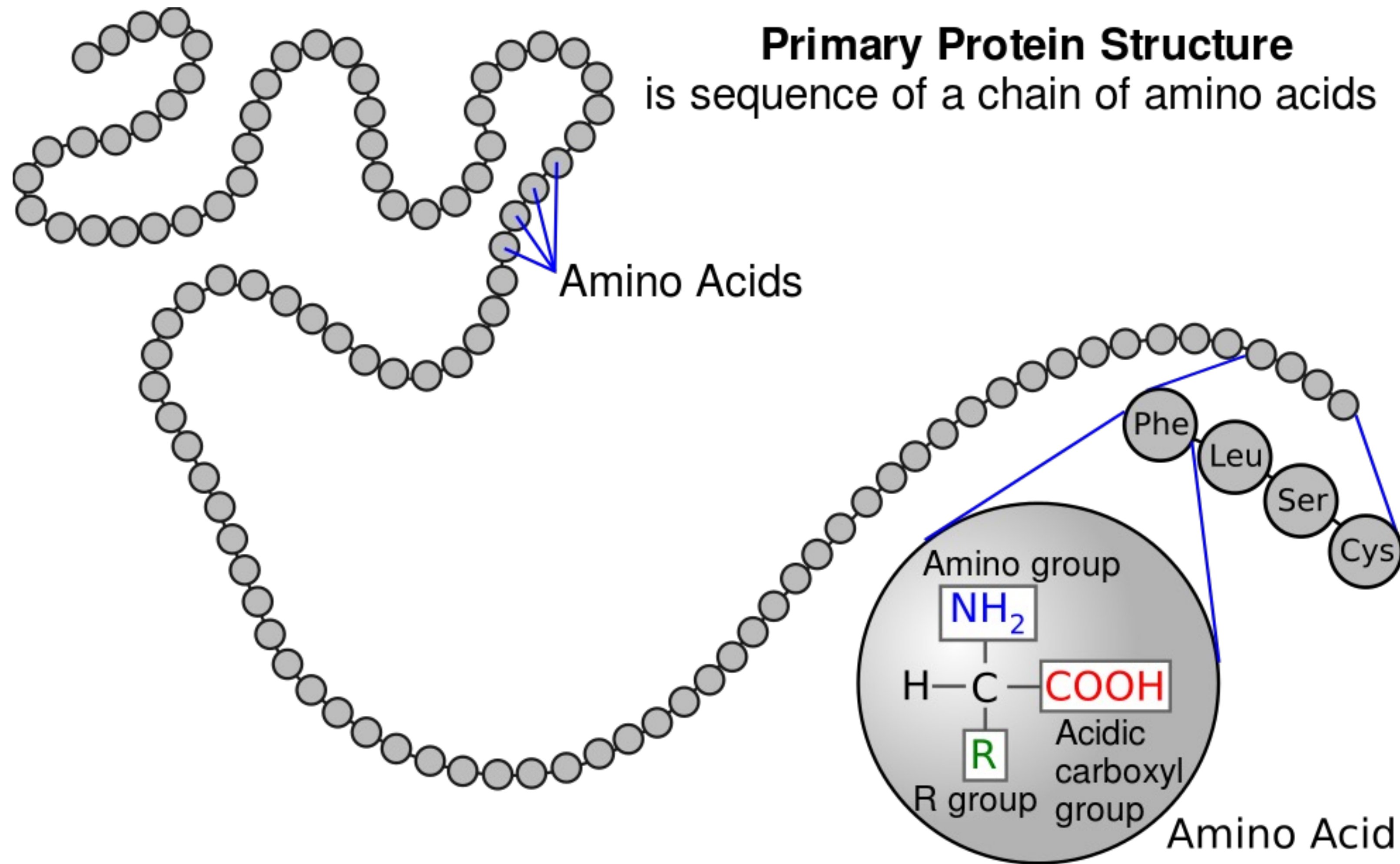
What are proteins and why do they stick everywhere?

# Amino Acids are the Monomer of Proteins

**Chart Key:** ● ALIPHATIC ● AROMATIC ● ACIDIC ● BASIC ● HYDROXYLIC ● SULFUR-CONTAINING ● AMIDIC ○ NON-ESSENTIAL ○ ESSENTIAL



# Amino Acids Connect to Form Polymers (Polypeptides/Proteins)

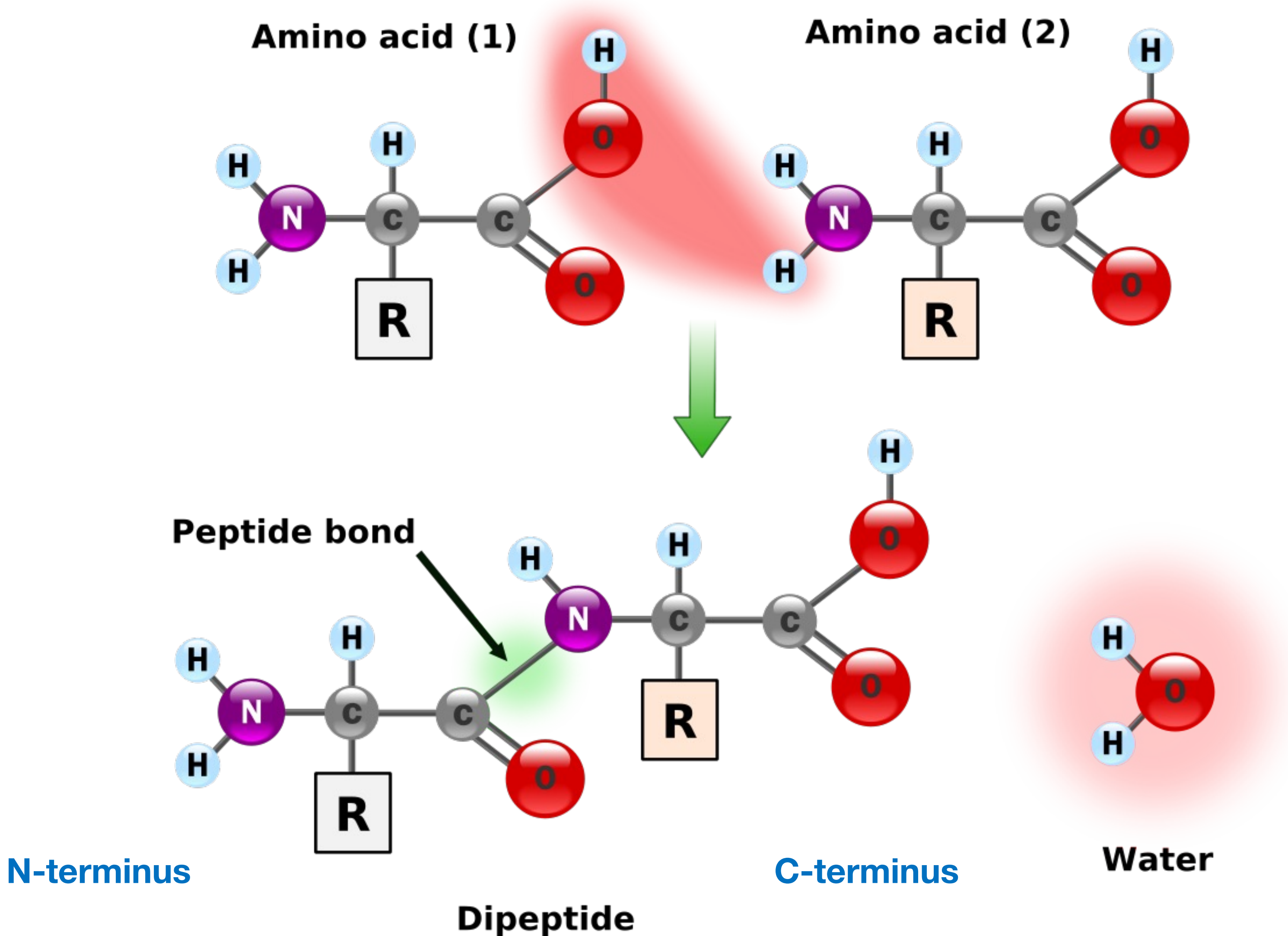


The order is very important for function

There is an enormous number of possible combinations of amino acids: even with only 40 units, there are  $20^{40}$  ( $\approx 10^{52}$ ) possibilities

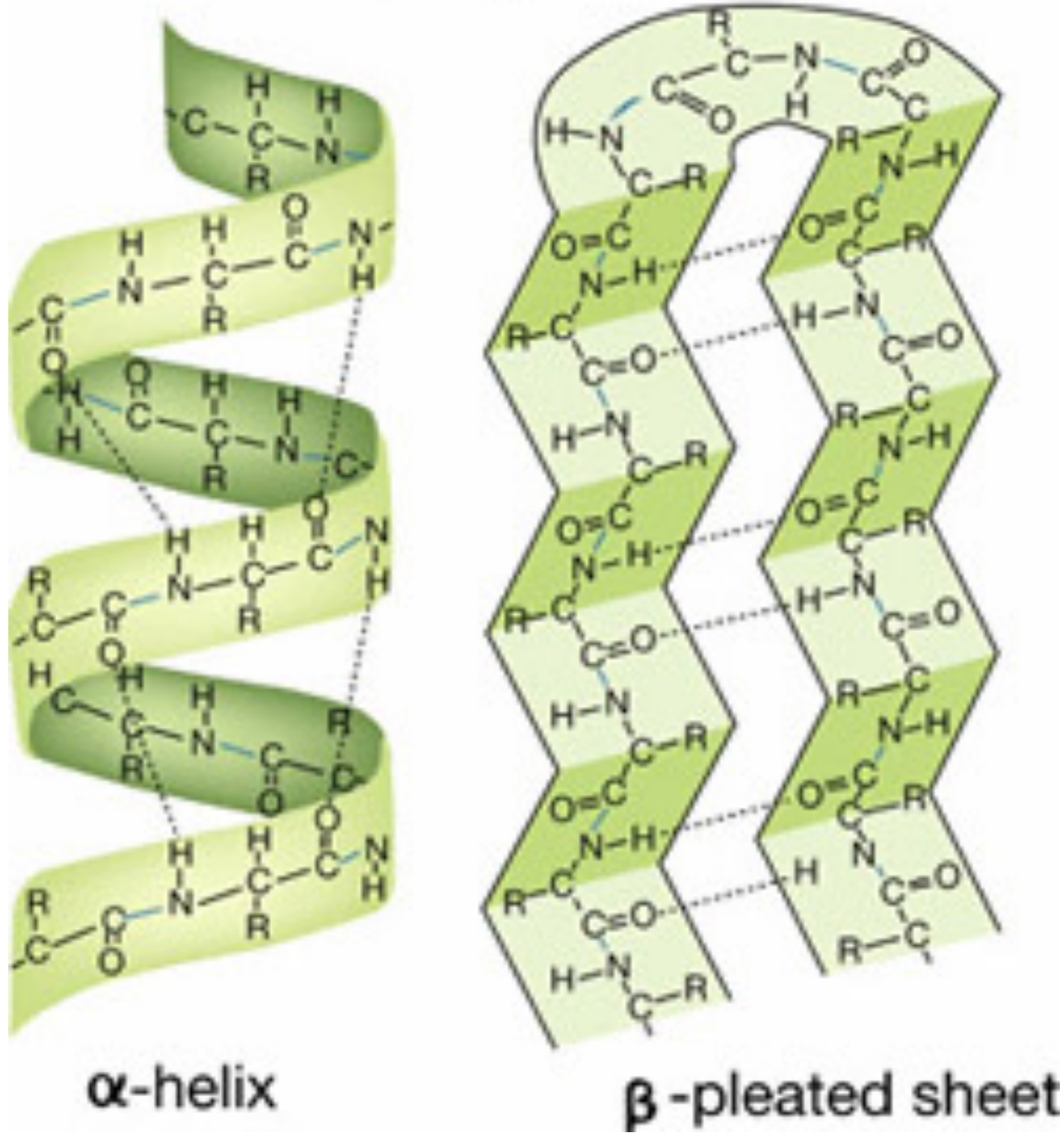
# Connection Between Amino Acids = Peptide Bond

Amino acids connect together to form polymers (polypeptides or proteins) of 40 to 10,000+ amino acid units



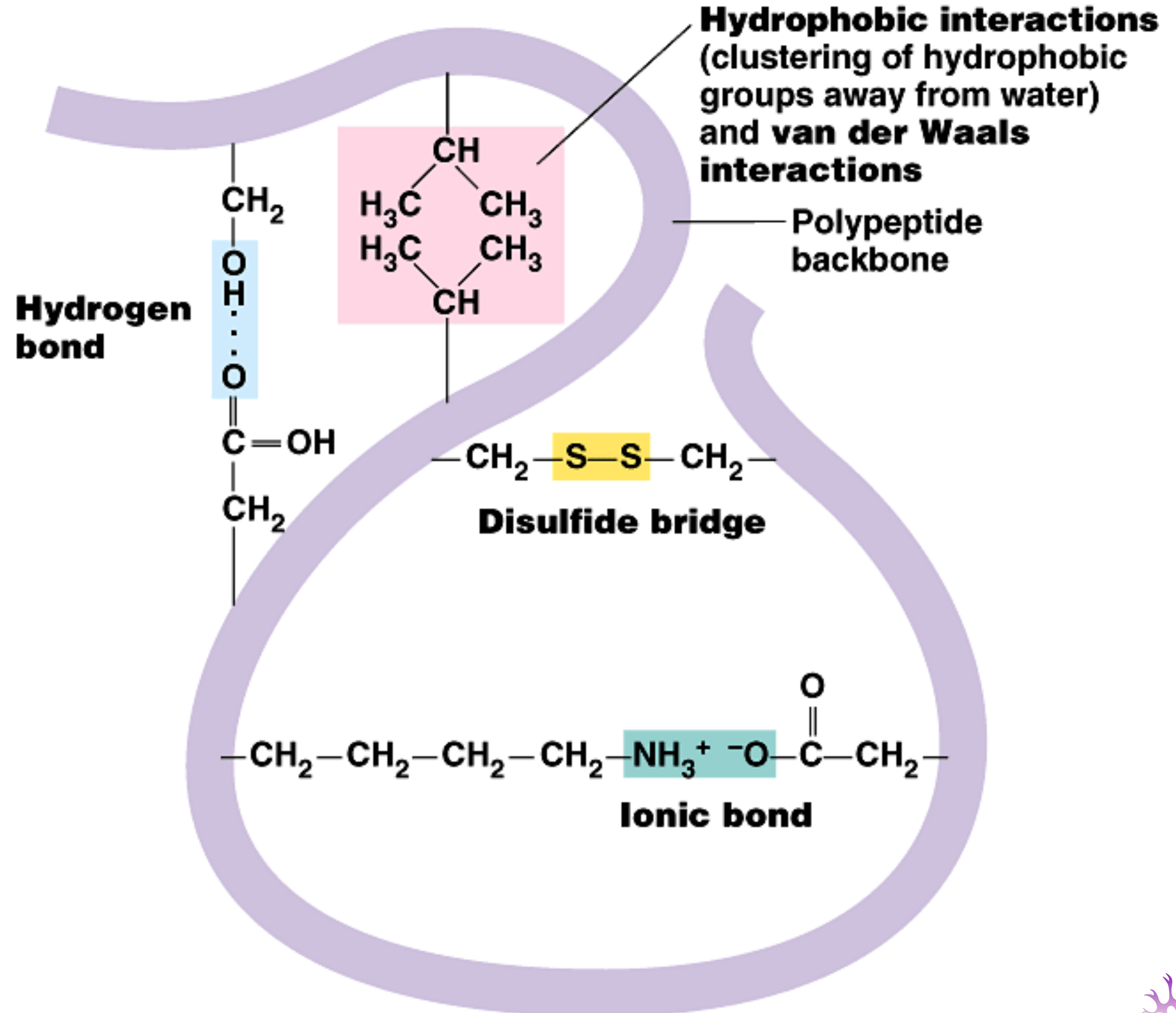
# Protein Secondary and Tertiary Structures

## Secondary structure



The polypeptide chain can hydrogen bond to itself, to produce the secondary structure

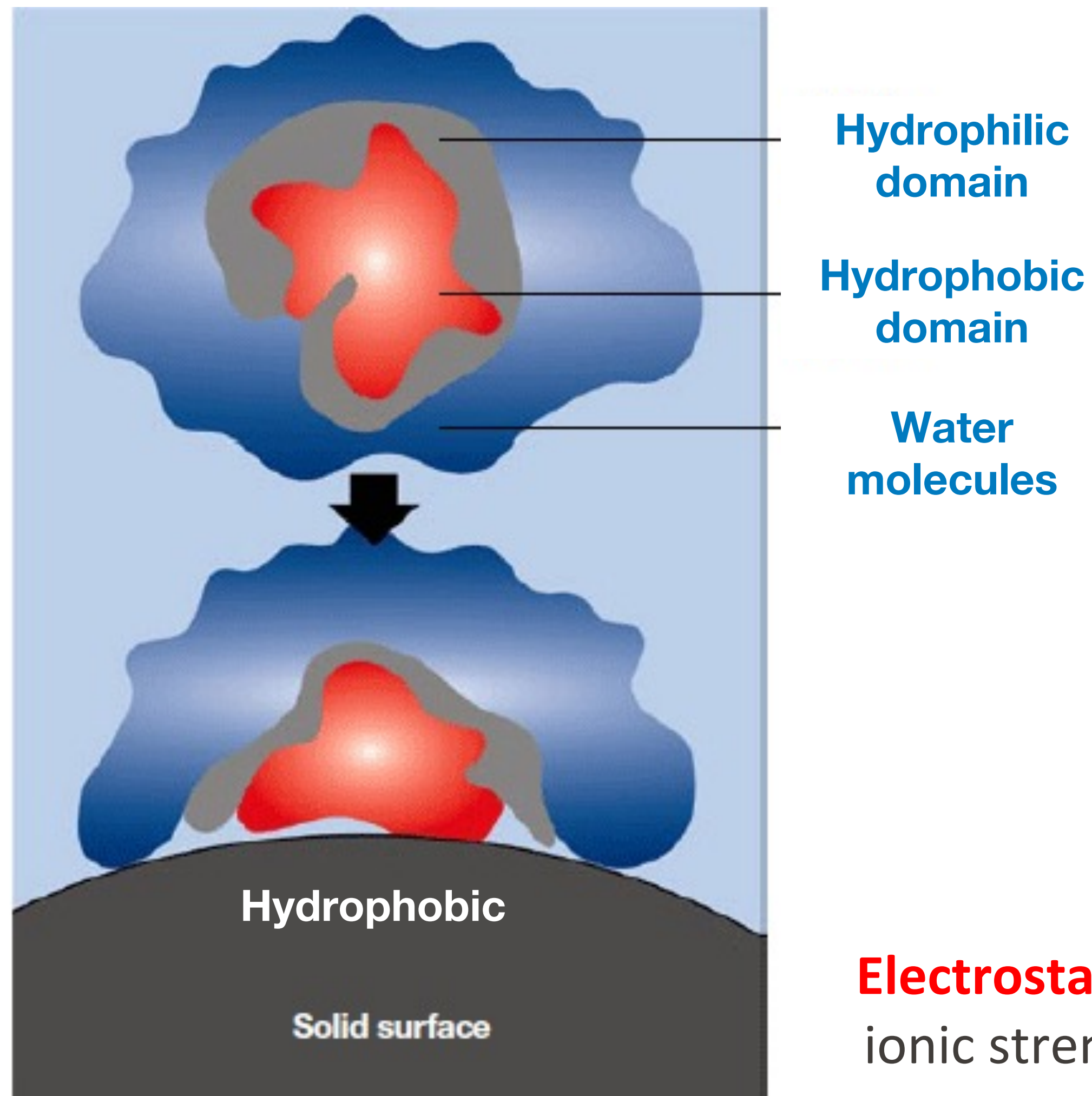
## Tertiary structure



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# Proteins on Surfaces – Adsorption and Denaturation

Proteins adsorb on surfaces to lower the free energy of the system



## Hydrophobic interactions

Hydrophobic groups on the protein: tend to be located inside of the overall protein structure



Shielded from aqueous surroundings

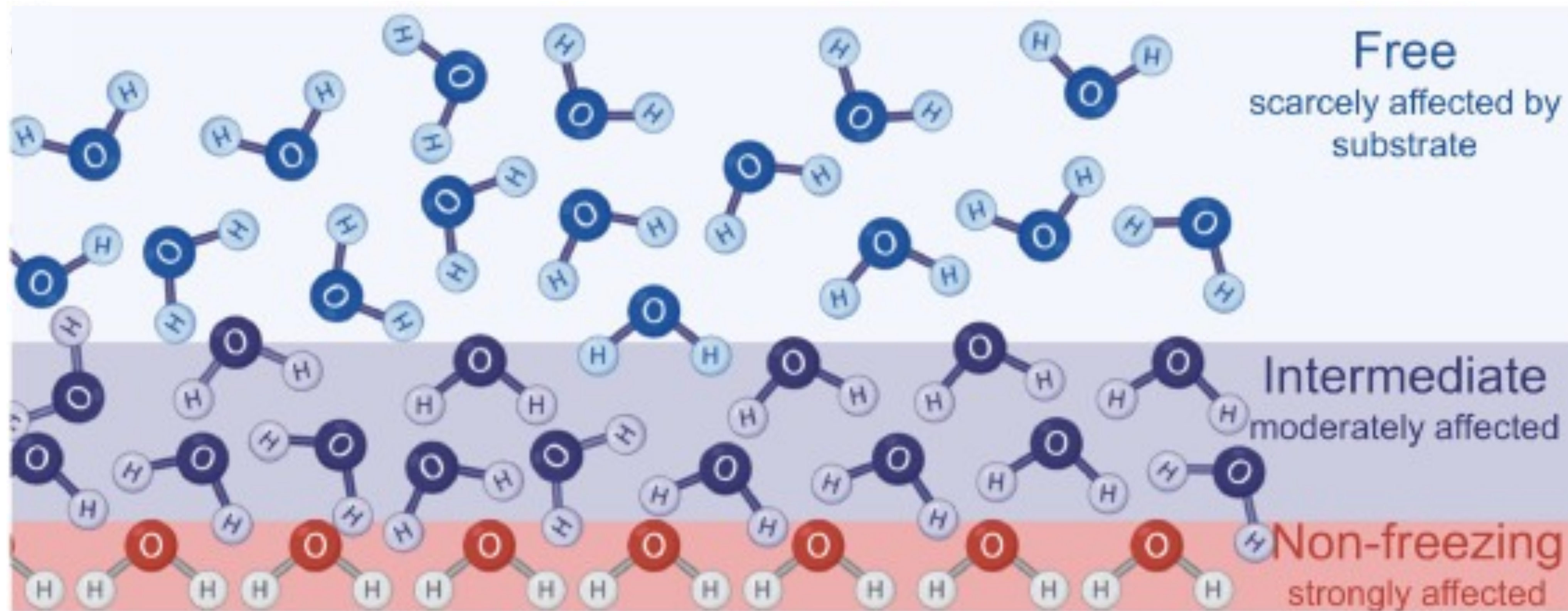
Upon encountering a hydrophobic surface, it is energetically favorable for hydrophobic amino acids to be close to the surface



Unfolding or denaturation of the protein

**Electrostatic interactions** with the surface are often important → pH and ionic strength of the solution can significantly influence protein adsorption

# Hydration Layer on the Surface Composed of Multiple Layers



“Bulk” water  
1-2 nm from  
surface

1-2 orders slower  
kinetics than  
“free” water

Tightly bound  
water with slow  
dynamics

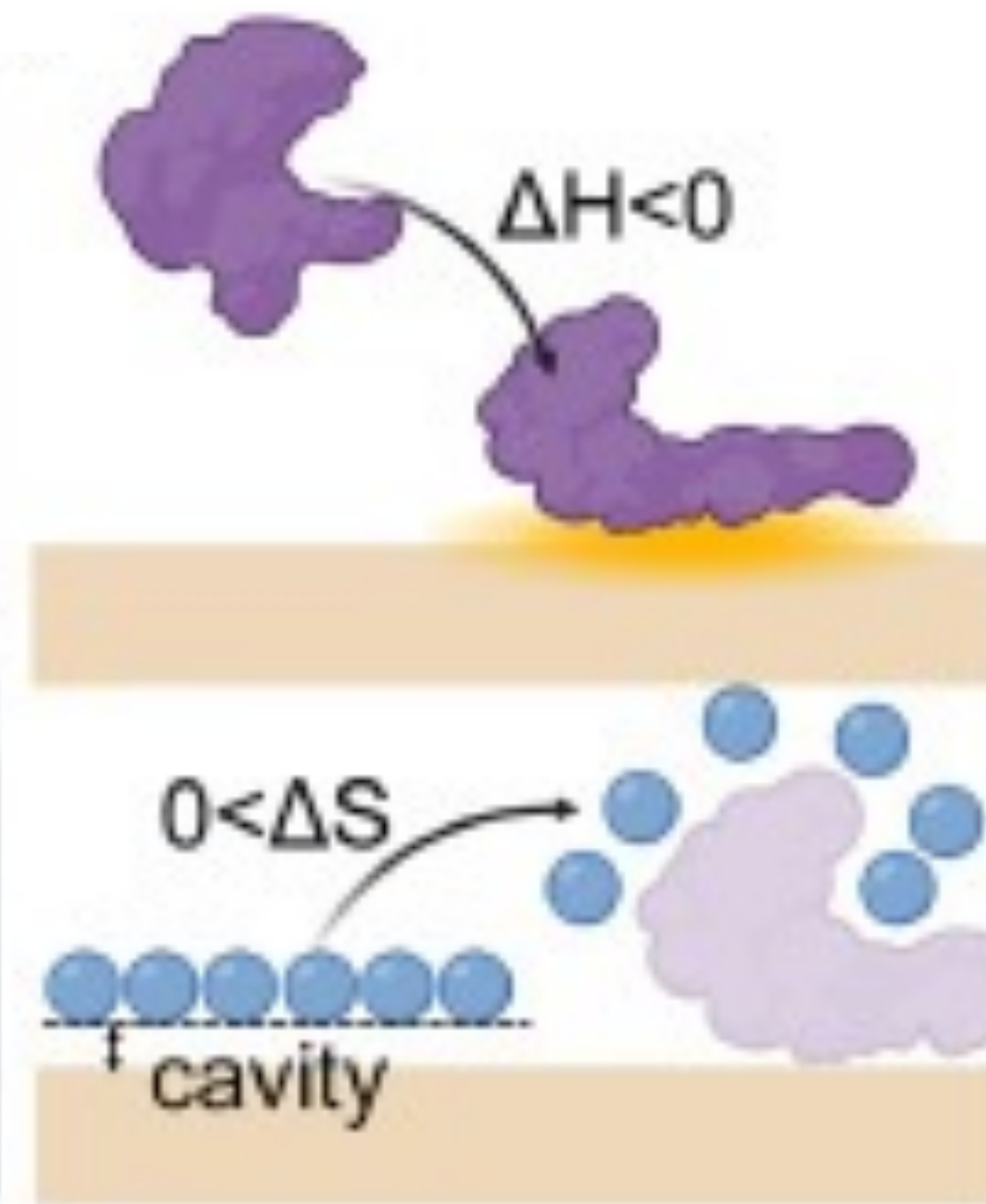
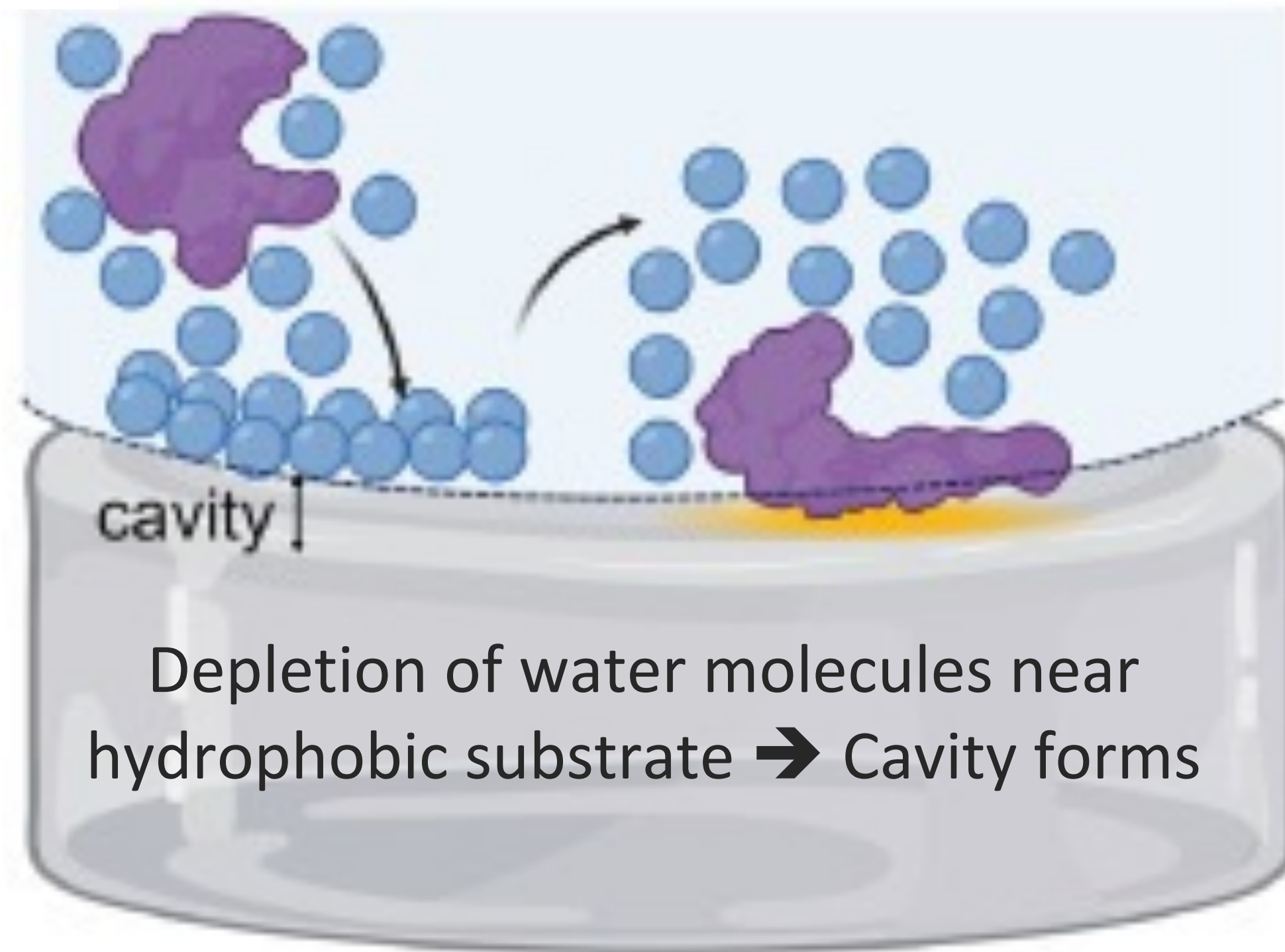
The interactions of water with the surface can drive how proteins stick to surfaces

# Thermodynamics of Protein Adsorption on Hydrophobic Surfaces

Hydrophobic surfaces don't have functional groups to form hydrogen bonding or strong polar or electrostatic interactions

H<sub>2</sub>O molecules stick together (H-bonds) since they avoid the surface

Reorganize into tight, ordered layer



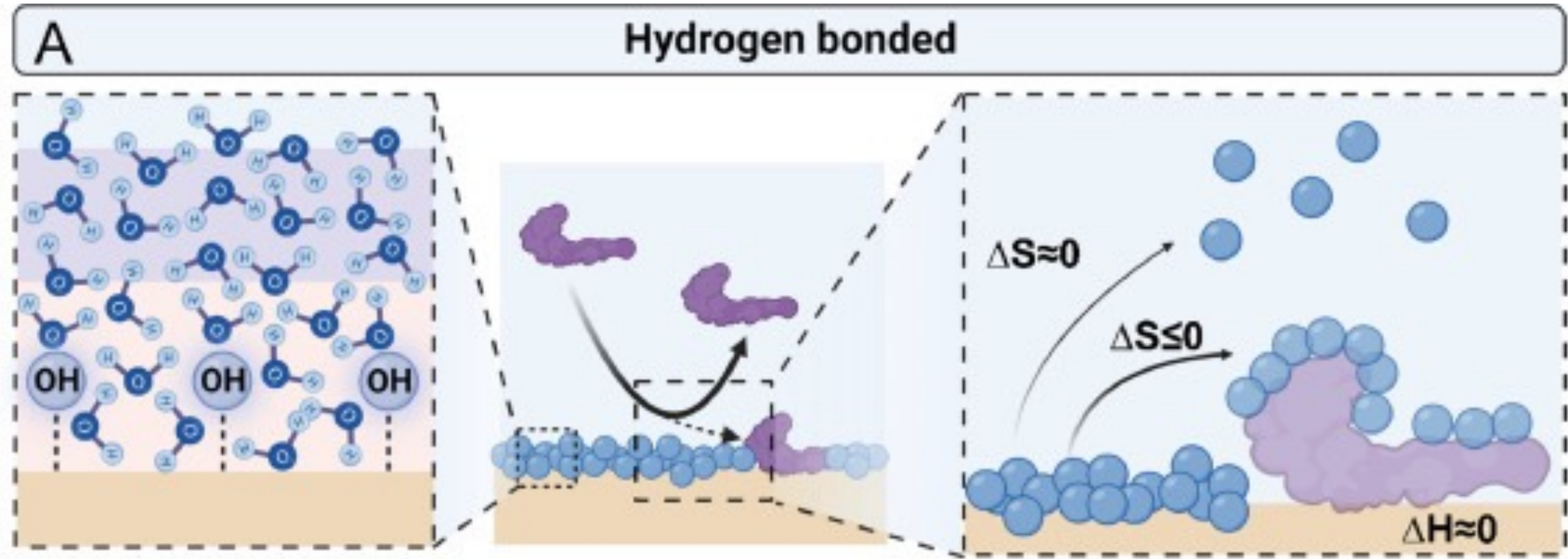
Hydrophobic bonds between hydrophobic amino acids and surface

Protein pushes bound water away

↓  
More freely moving water

# Thermodynamics of Protein Adsorption on Hydrophilic Surfaces

Hydrophilic surfaces can provide more energetically stable bonds with proteins and water molecules like polar and hydrogen bonding

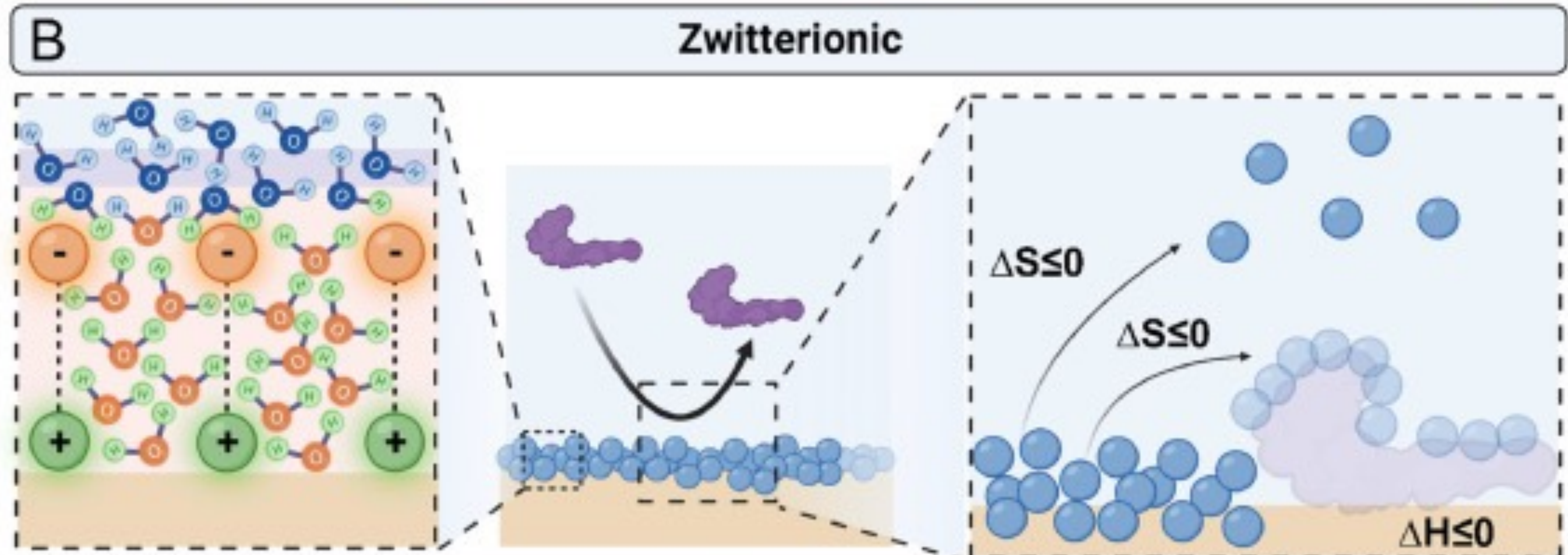


Water sticks to surfaces through H-bonds

To stick, protein must gain more H-bonds to the surface than the water-surface H-bonds being broken (protein adsorption not favorable)

# Thermodynamics of Protein Adsorption on Hydrophilic Surfaces

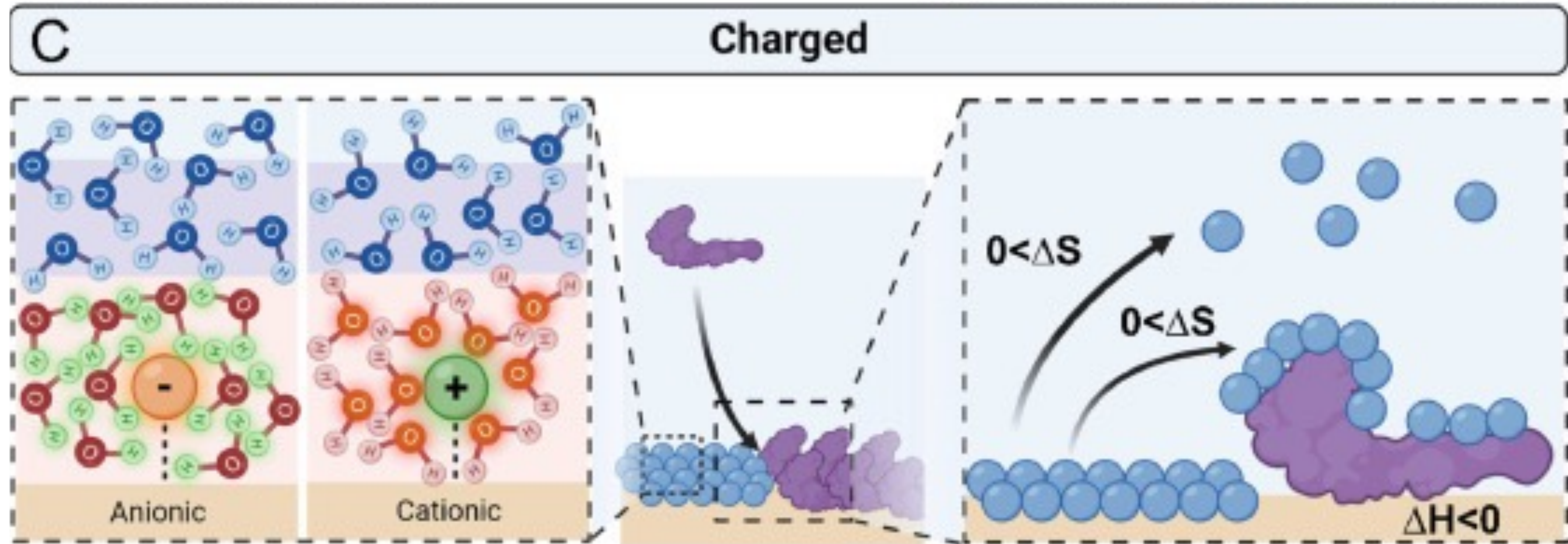
Zwitterionic surfaces have both + and – charges → surface is overall neutral  
This allows water near the surface to keep its natural H-bonding network



Water in happy state Zwitterionic surfaces resist protein adsorption because the water near the surface is already stable, so removing it costs entropy

# Thermodynamics of Protein Adsorption on Charged Surfaces

Charged surfaces (+ or – charges) make water near the surface very ordered → water must align around the electric field (restricted motion)



Ordered water

Ordered water release leads to entropy increase  
Protein forms electrostatic interactions with the surface

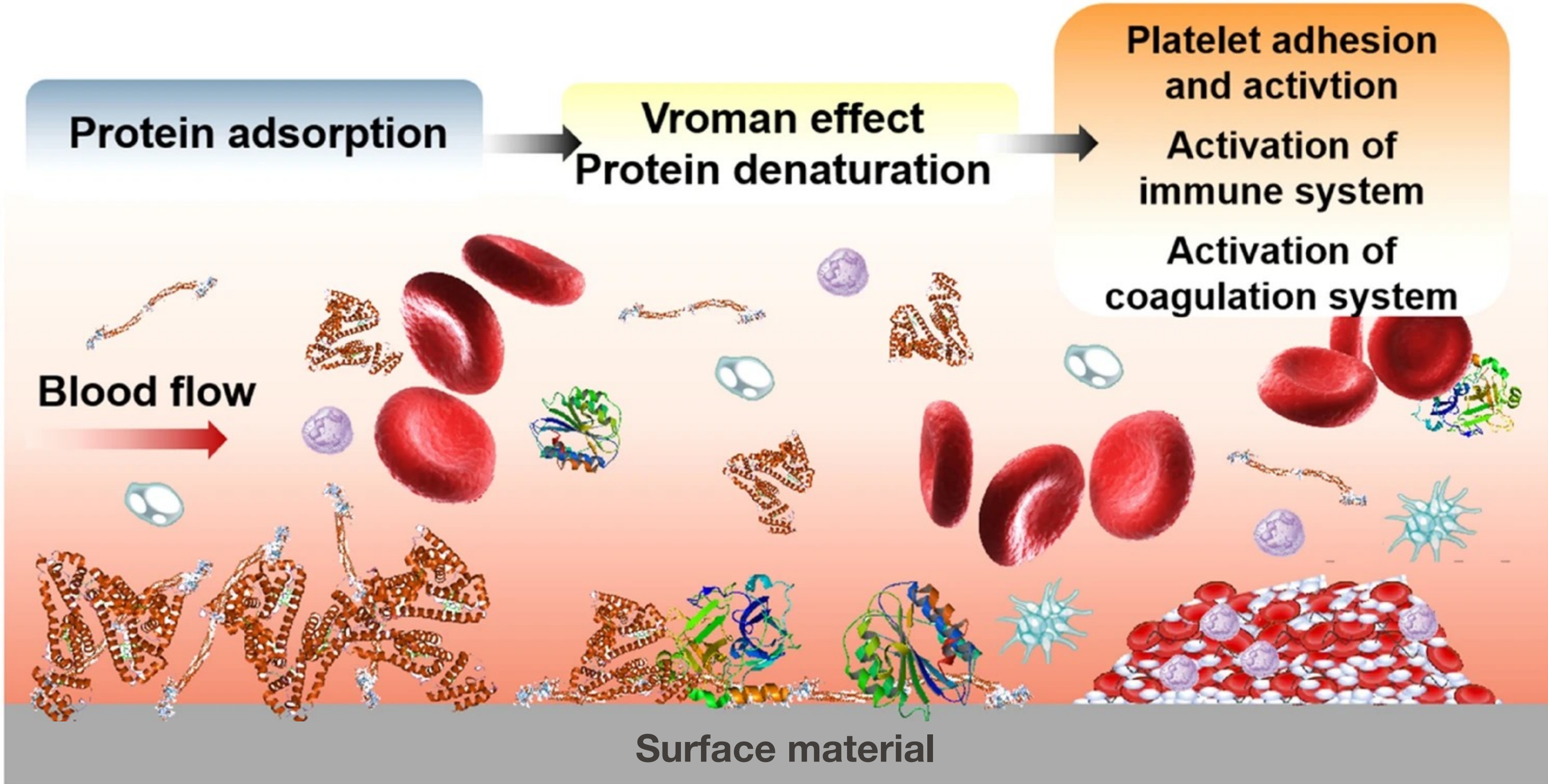
# Thermodynamics of Protein Adsorption Summary

Surface Property	Water at Surface	Entropy Effect	Enthalpy Effect	Protein Adsorption Strength
Hydrophobic	Water is highly ordered (unhappy)	$\Delta S > 0$	Hydrophobic contacts give $\Delta H < 0$	Strong (can denature protein)
Hydrophilic / Hydrogen-Bonding	Water is stable and hydrogen-bonded naturally	$\Delta S \approx 0$	Only favorable if protein forms extra H-bonds	Weak–Moderate
Zwitterionic	Water remains very stable, highly “bulk-like”	$\Delta S \leq 0$	Electrostatics small $\Delta H \leq 0$	Very Weak (Protein-Resistant)
Charged (Anionic / Cationic)	Water highly oriented by electric field (low entropy)	$\Delta S > 0$	Electrostatic binding gives $\Delta H < 0$	Strong

# Proteins on Surfaces – Vroman Effect

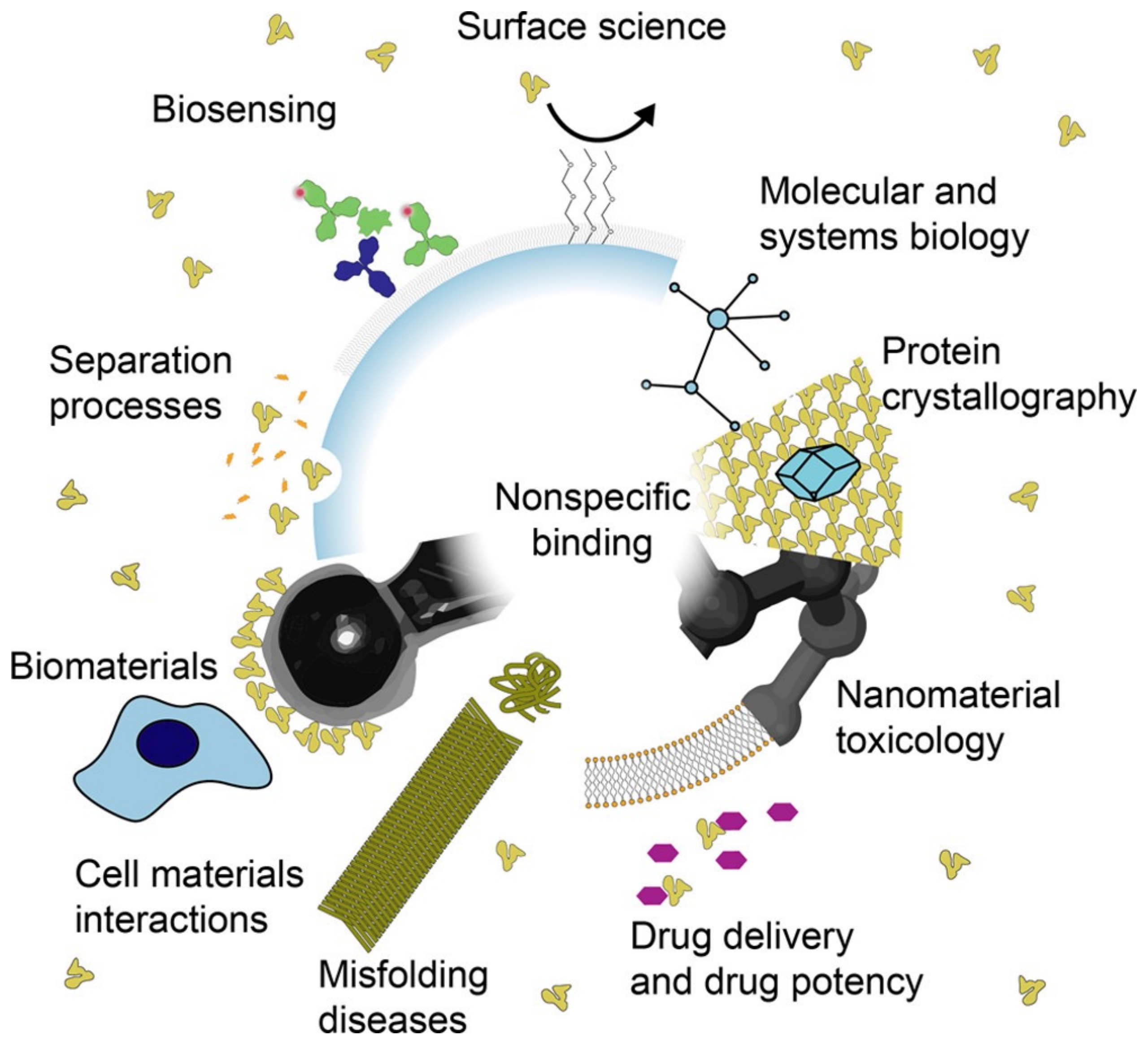
Small, abundant molecules (albumin, fibrin) will rapidly and reversibly coat a surface, gradually being replaced by larger molecules with higher affinity for the surface

Conformational change of many proteins upon nonspecific adsorption to the interface



Ji, et al. | *Adv. Fiber Mater.* | 2023

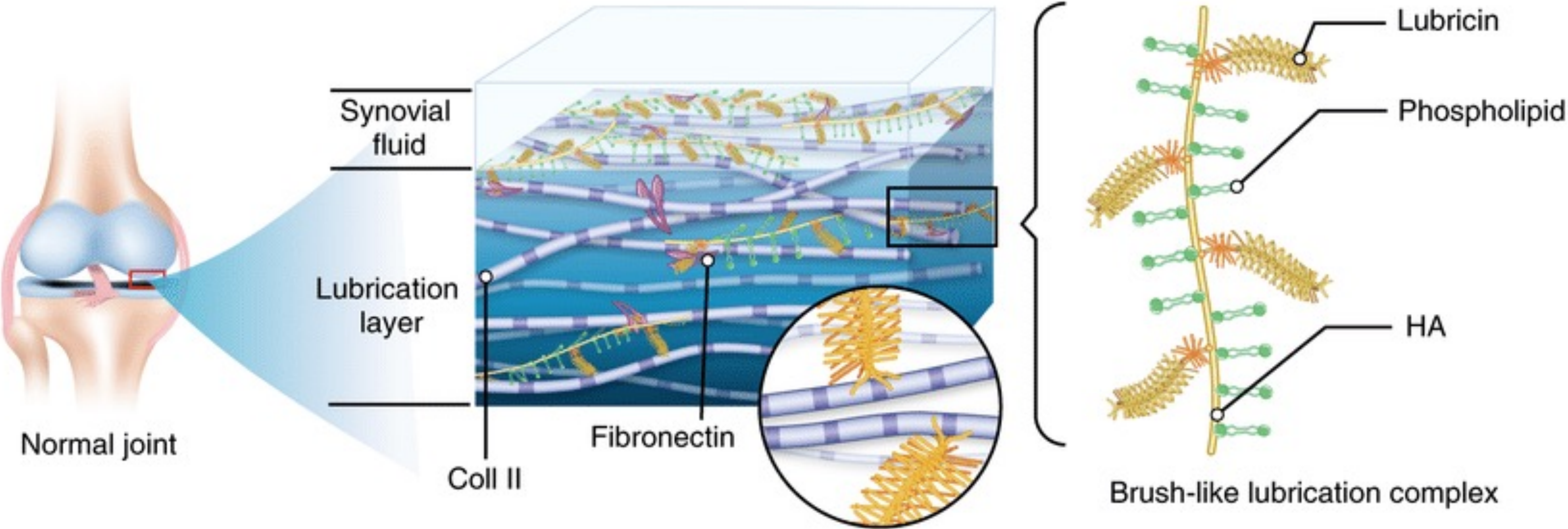
# Proteins on Surfaces has Major Implications in Many Fields



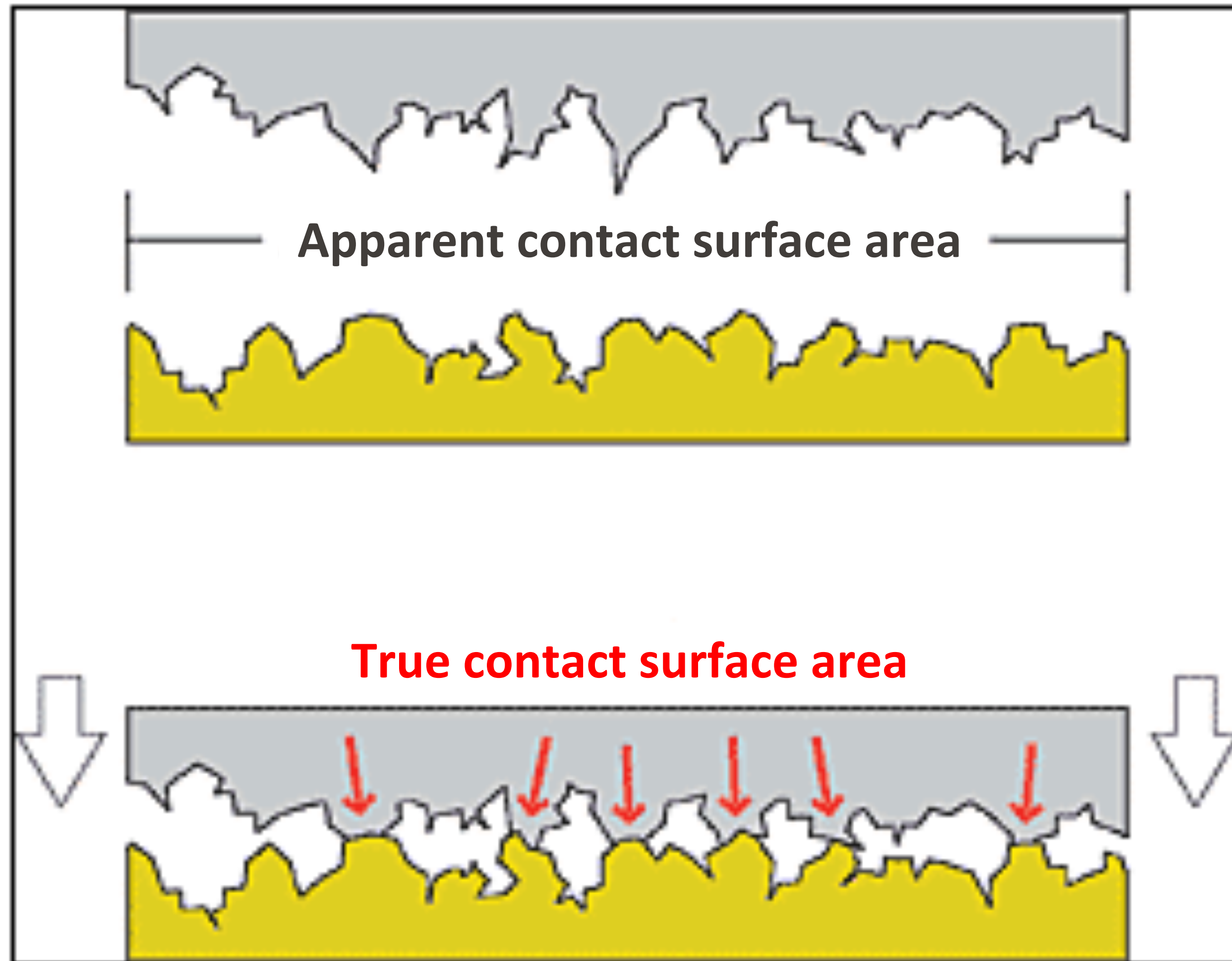
Frutiger, et al. | Chem. Rev. | 2021

# Case Study of Proteins on Surfaces: Synovial Fluid in Joints

Synovial fluid contains lubricin and hyaluronic acid that **adsorb** to cartilage surfaces → Hydrated brush-like layer reduces friction



# Tribology: Rubbing of Surfaces that are Never Perfectly Flat



The normal load is borne by the asperities on each surface coming into contact with each other

Contact areas are small (**asperities**)



Pressures are large



Asperities become elastically and plastically deformed upon contact with each other

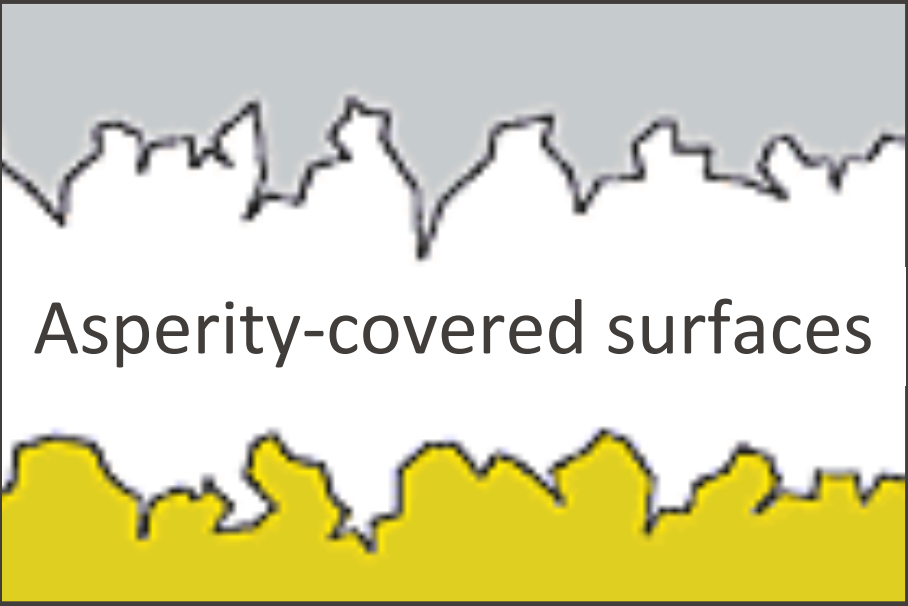


Contact areas increase

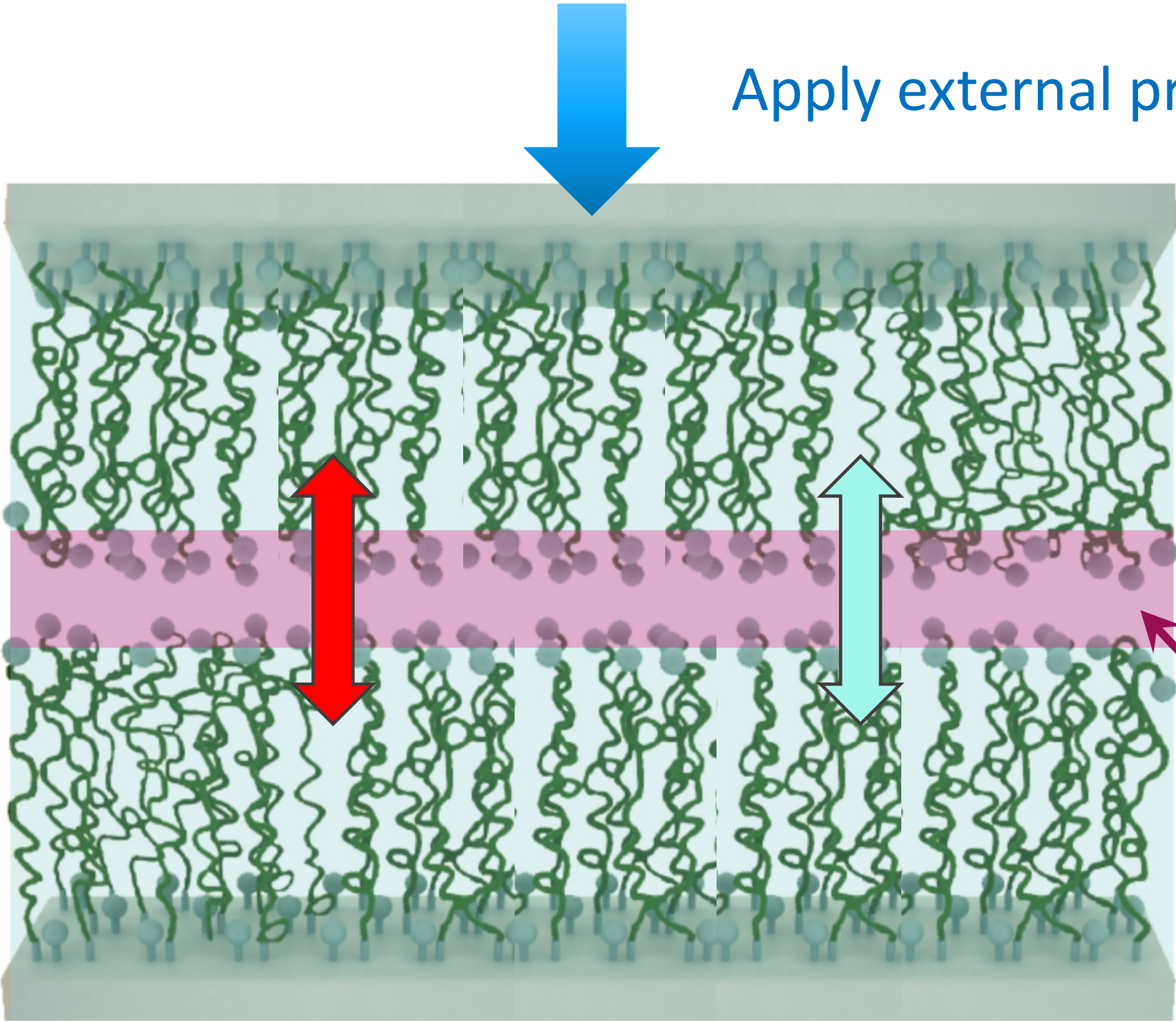


Contact pressure may eventually exceed the elastic limit of the material (wear)

# Polymer Brushes Reduce Friction by Acting as Lubricating Layers



Osmotic pressure  
Resists compression



Entropic forces  
Resists interpenetration

Hydrated fluid film  
enables smooth sliding

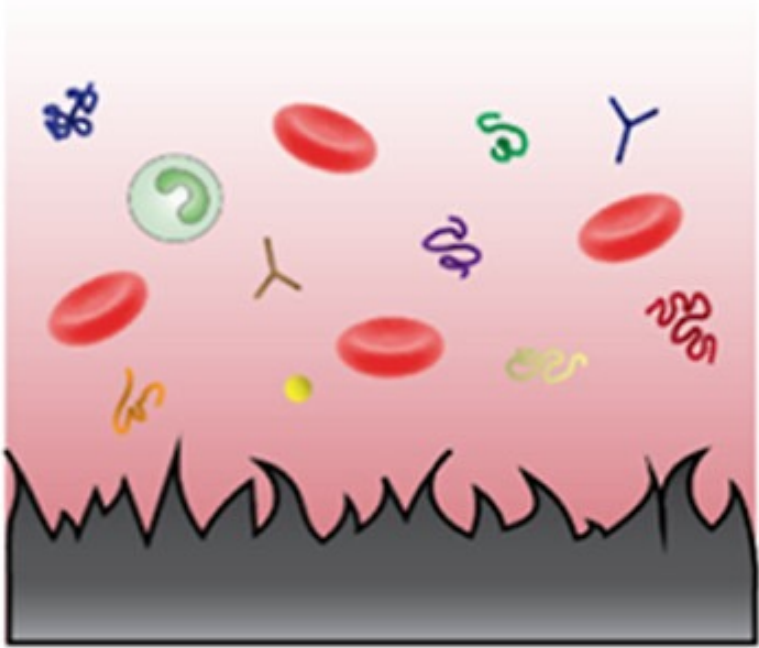
Polymer brush-coated surfaces

Polymer brushes lubricate by effectively reducing contact between the hard, asperity-covered surfaces, and replacing it with brush-brush contact

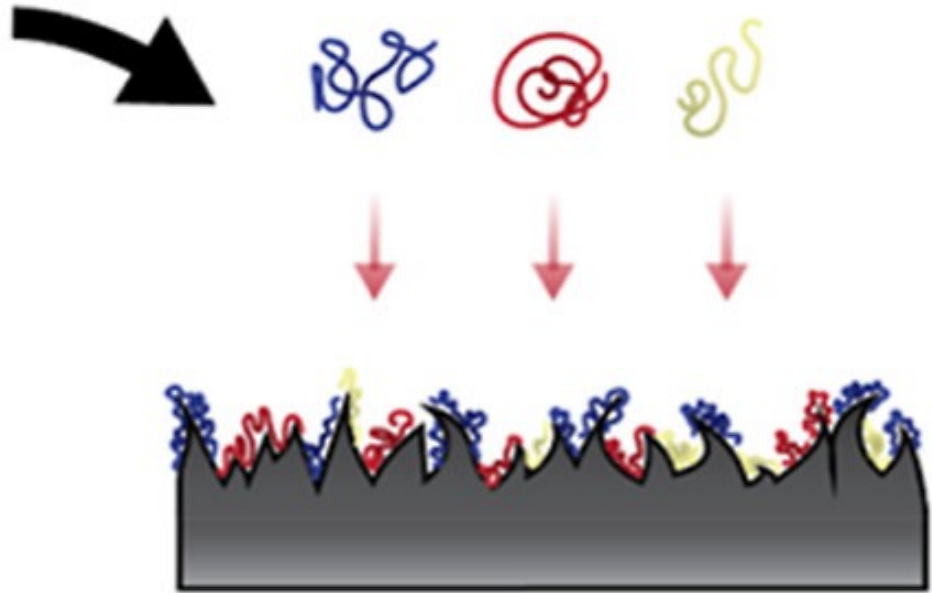
# Synovial Proteins Rapidly Adsorb to Implant Materials



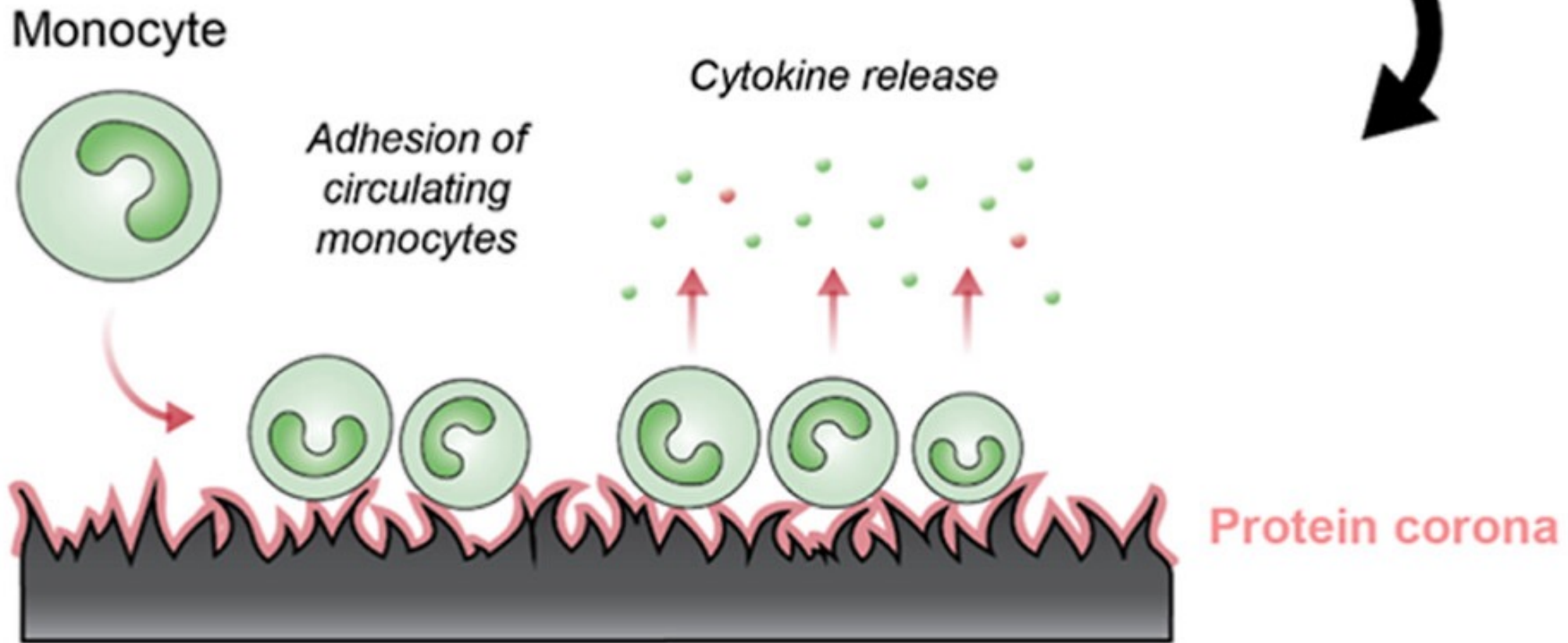
1) Implantation of titanium hip implant



2) Material interaction with blood and synovial fluid

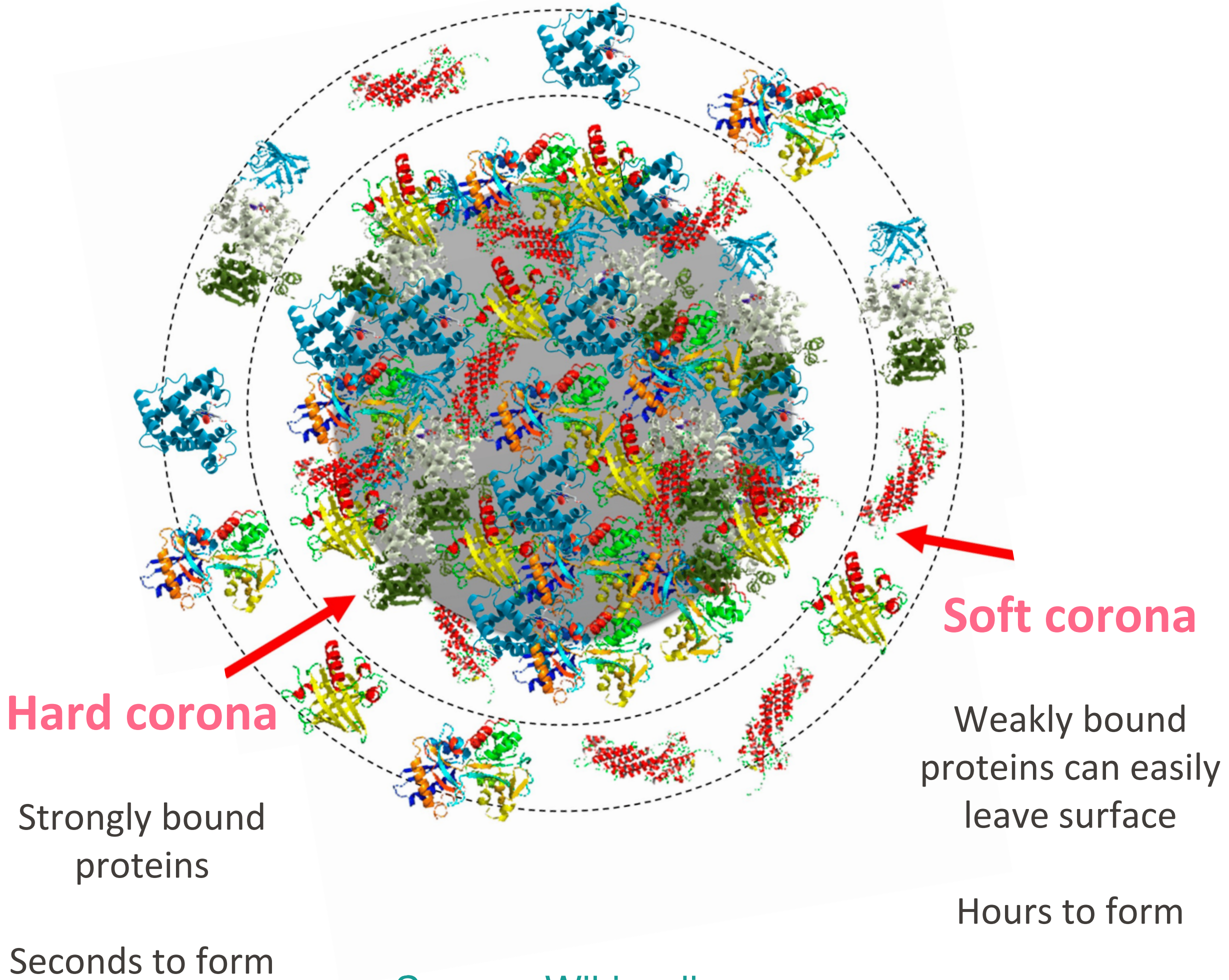


3) Protein adsorption and formation of a complex corona

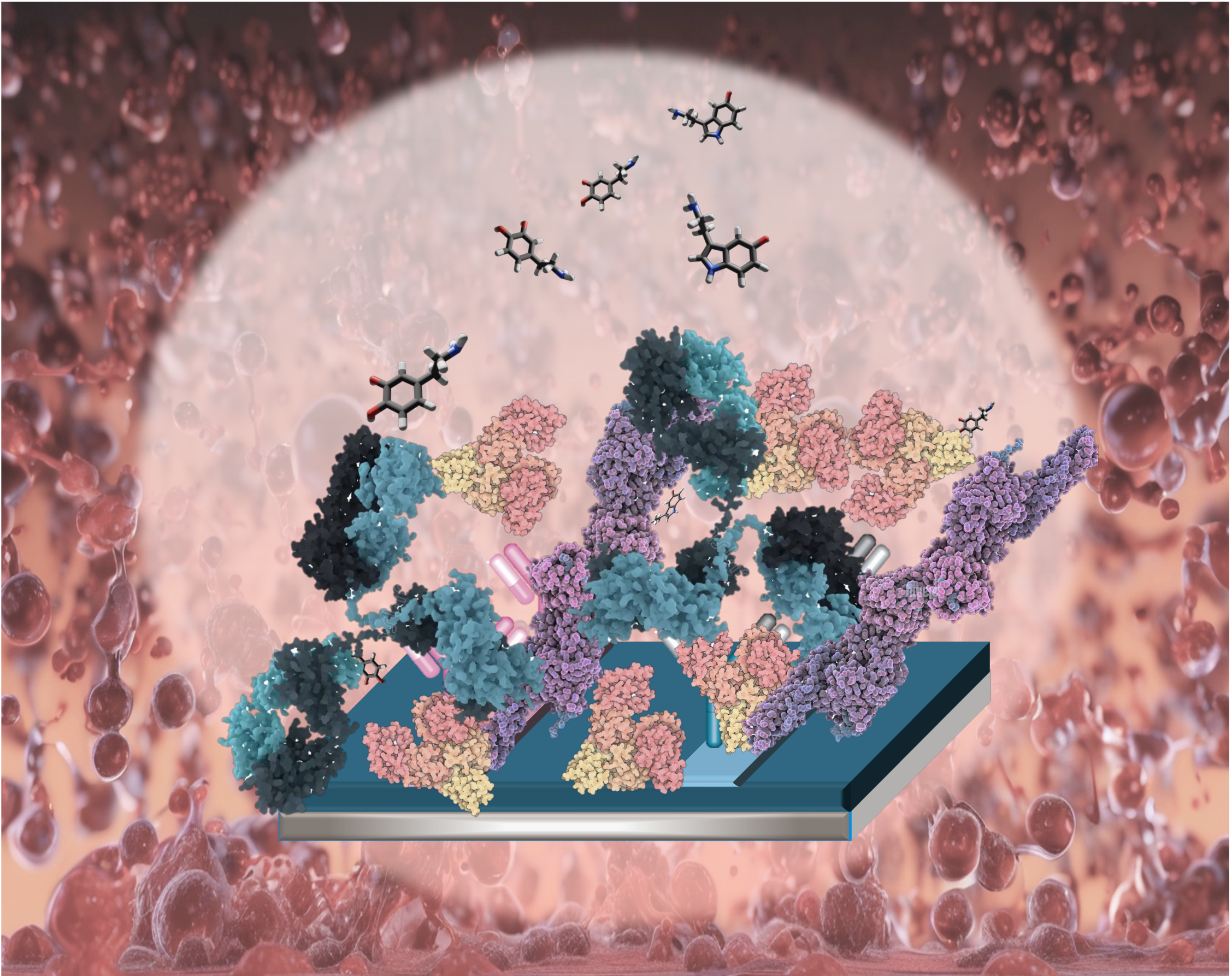


4) First cellular response

# Case Study of Proteins on Surfaces: Biosensors vs. Biofouling



Source: Wikipedia



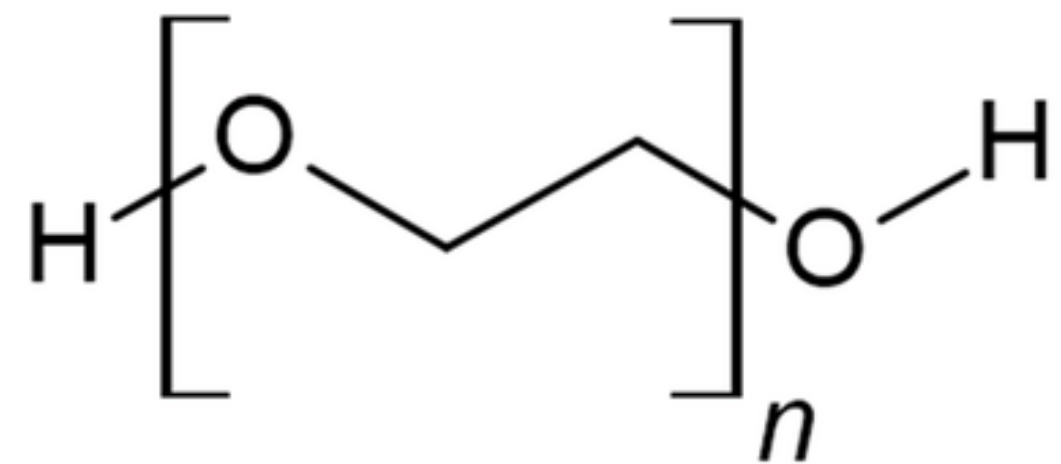
# Key Takeaways

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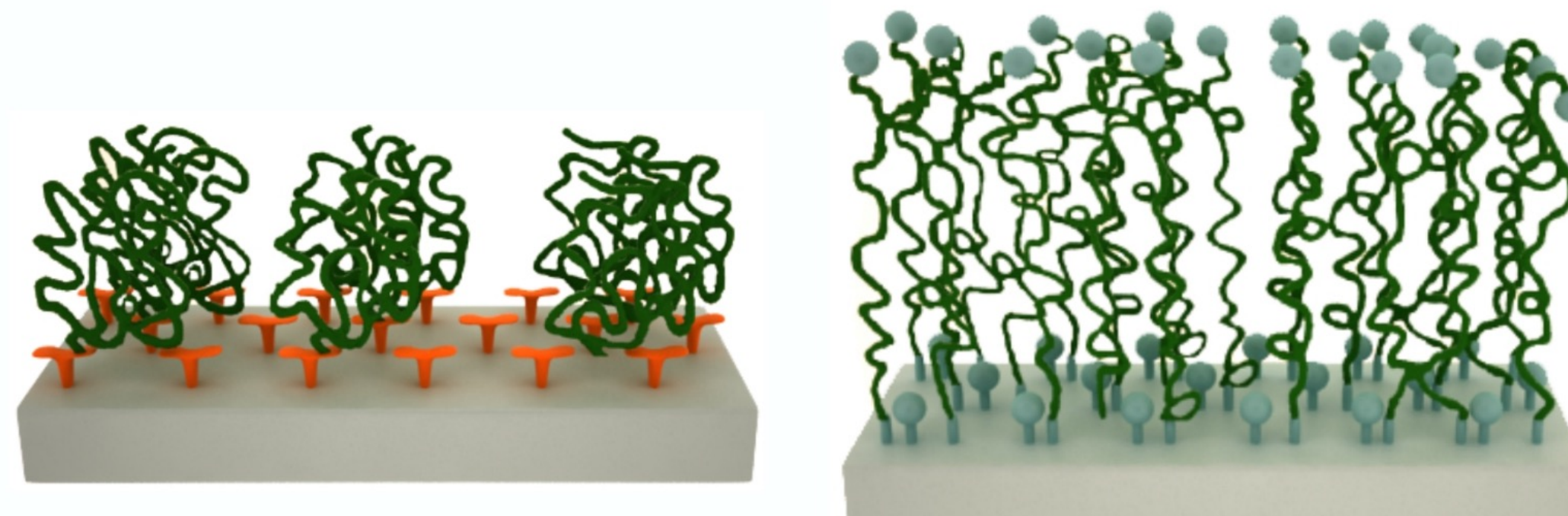
- One of the biggest challenges surfaces face in biological systems is nonspecific binding of proteins
  - Proteins stick to surfaces through intermolecular interactions
- Surface properties (hydrophilic/hydrophobic) drive the propensity for proteins to stick to surfaces
- The Vroman effect explains the dynamics of protein adsorption over time
  - Challenges and opportunities of protein interactions

# How To Control Protein Adsorption Using Polymer Brushes

Certain polymer brushes have been shown to be highly effective in preventing protein adsorption



**Poly(ethylene) glycol (PEG)**



Neutral brushes

Highly hydrated and flexible

Antifouling benchmark

Polymer layer prevent proteins reaching the solid surface beneath

non-charged (negligible electrostatic interactions with proteins)

hydrophilic (no hydrophobic interactions with proteins)

free-energy (penetration of a protein into the brush is unfavorable)

# What is a Polymer?

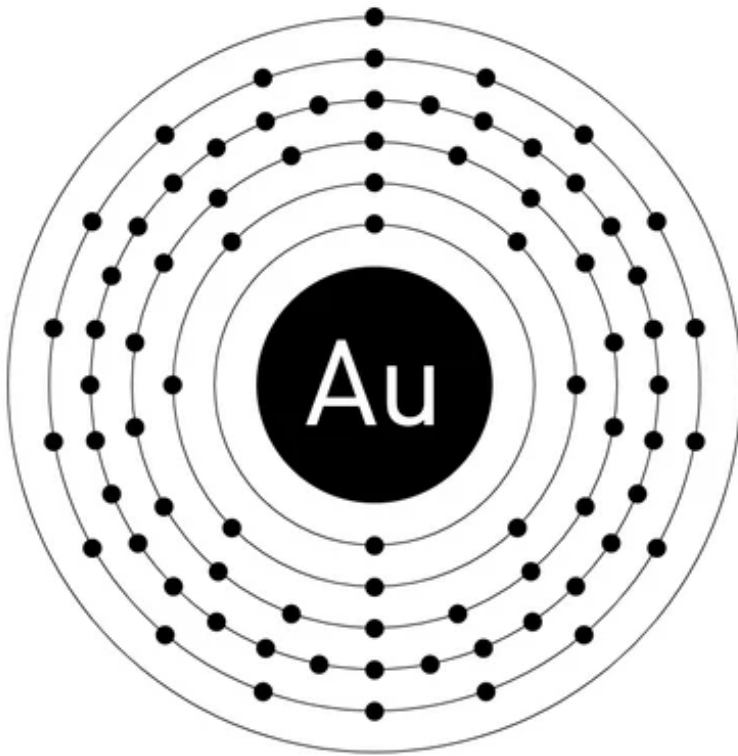
Metal



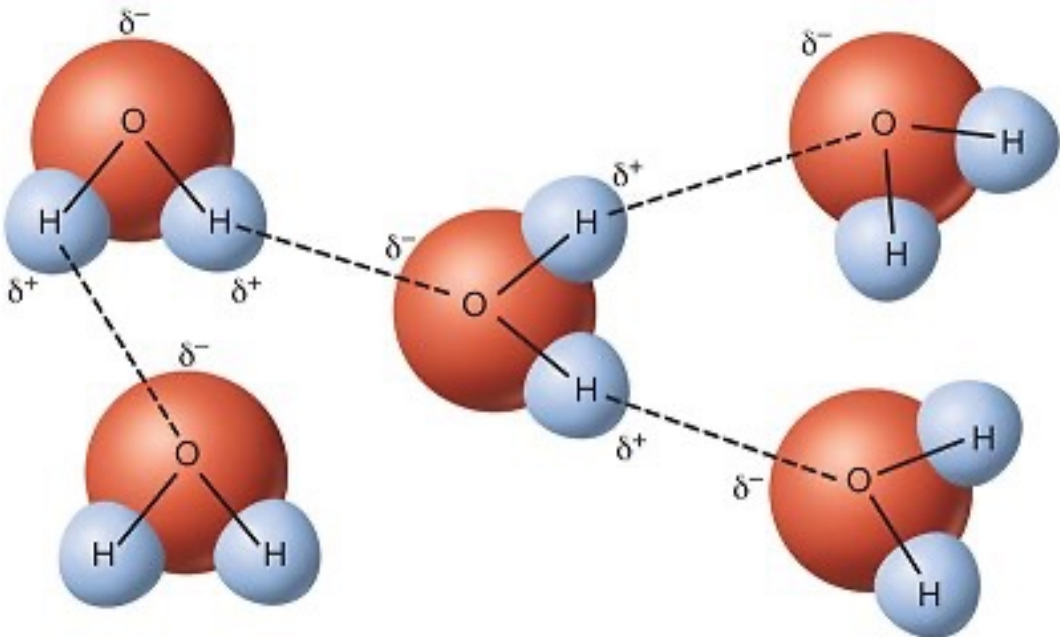
Water



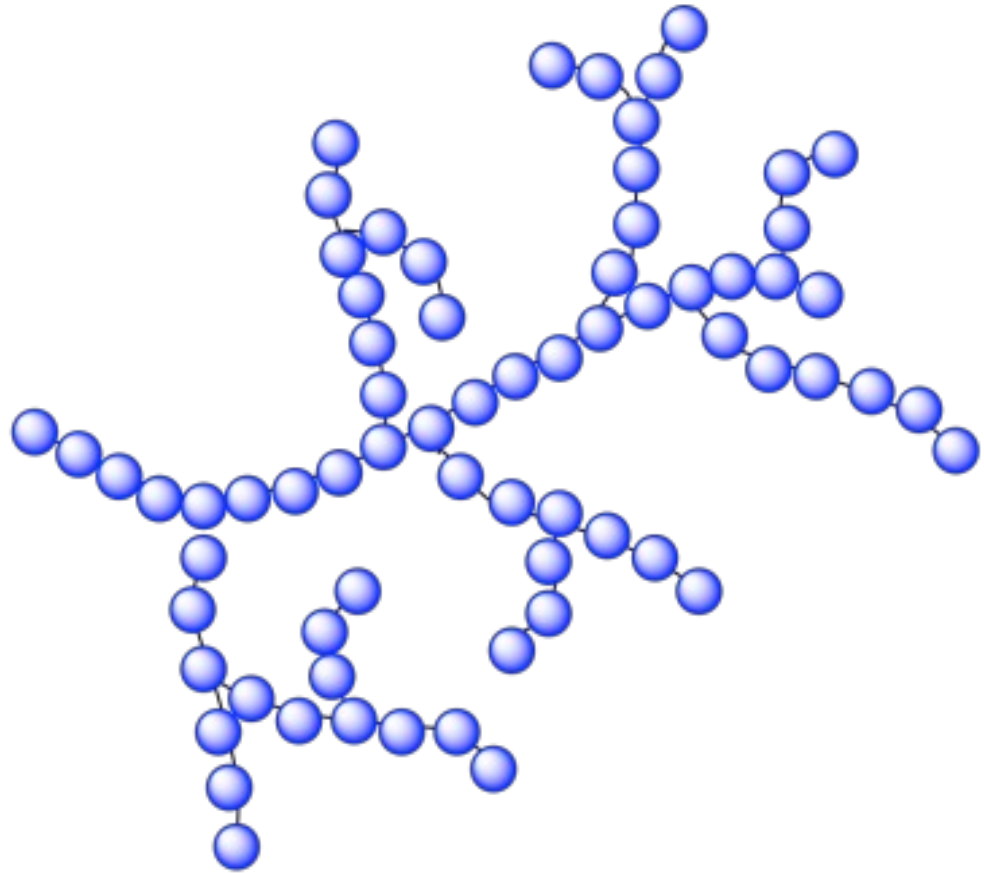
Polymer



Atoms



Molecules



Chains of molecules

# Polymers are All Around Us

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**Teflon**



**Rubber**



**Plastic**



**Polyester**



**Sugercane**



**DNA**



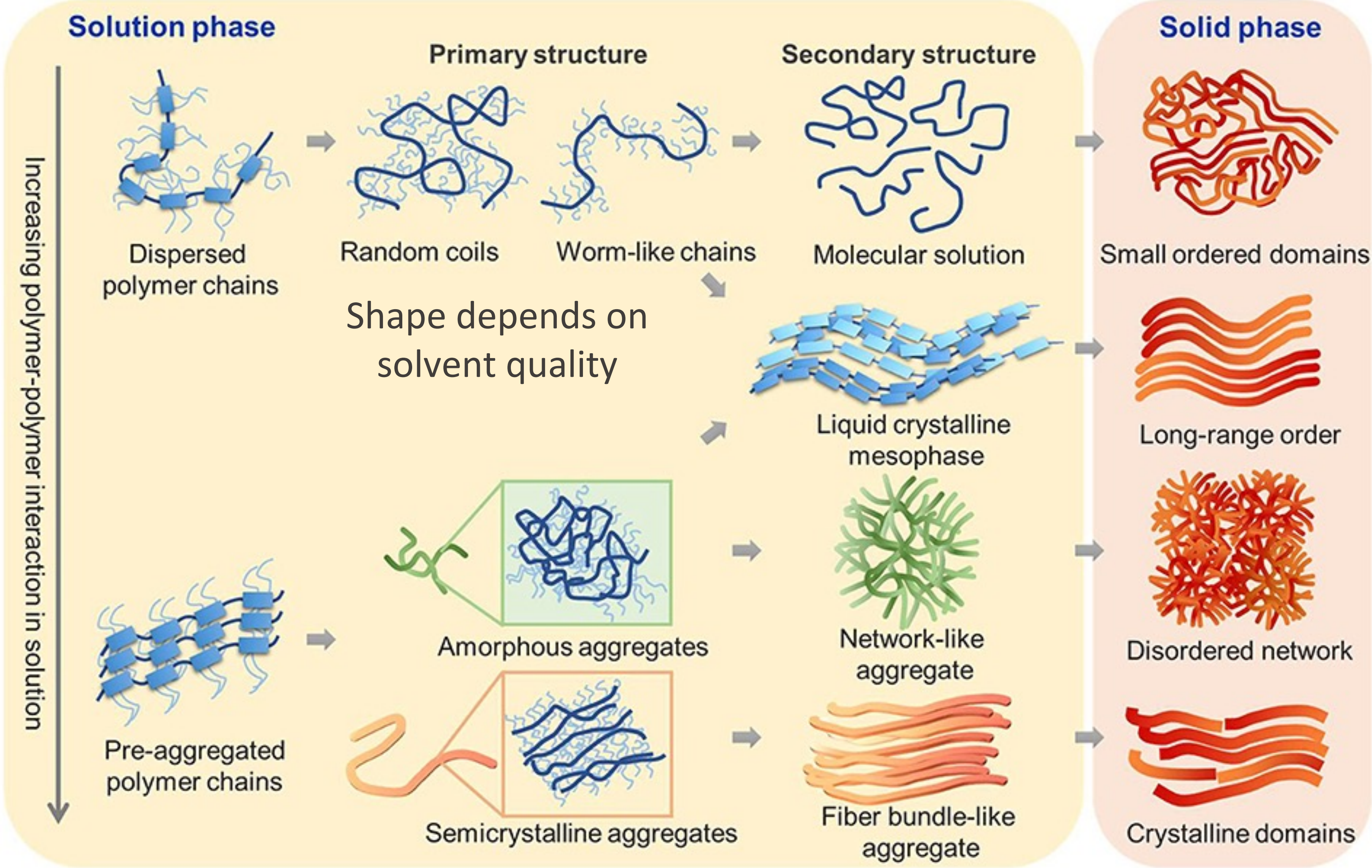
**RNA**



**Wool**



# Polymer Chain Conformation from Solution to Solid Phases

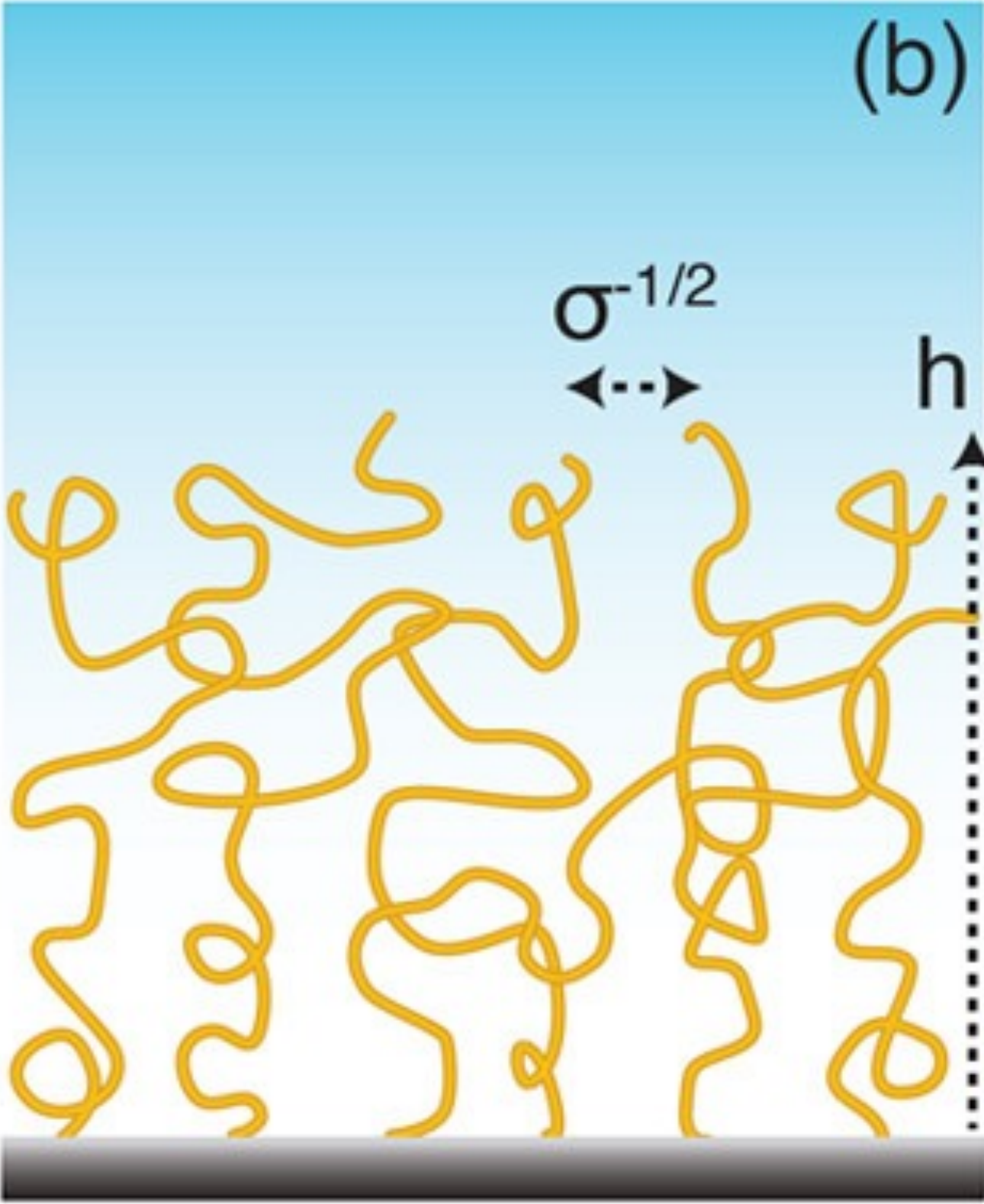


# Polymers on Surfaces – Grafting Density Influences Structure

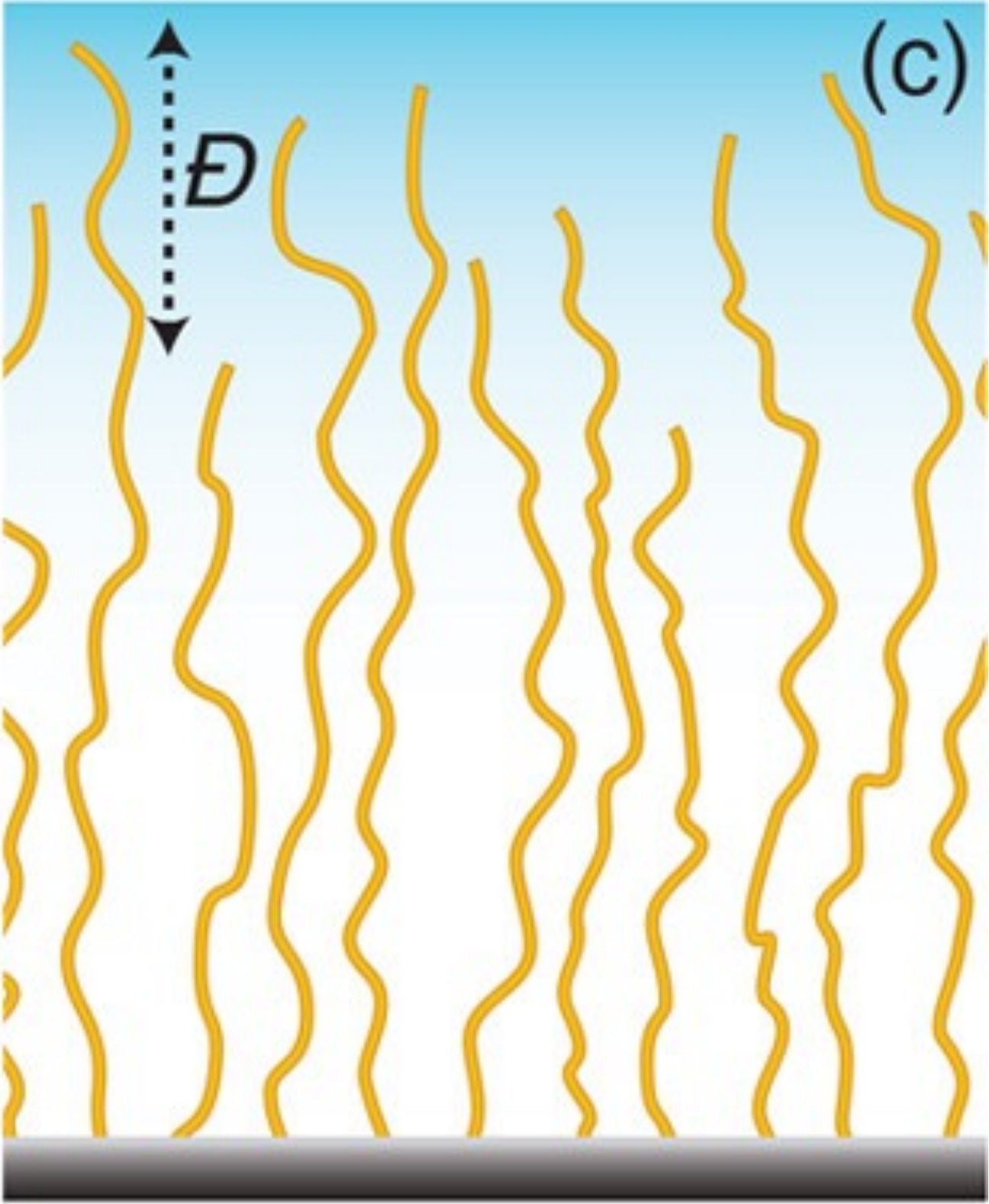
increasing grafting density  $\sigma$  .....



Low density:  
chains behave independently

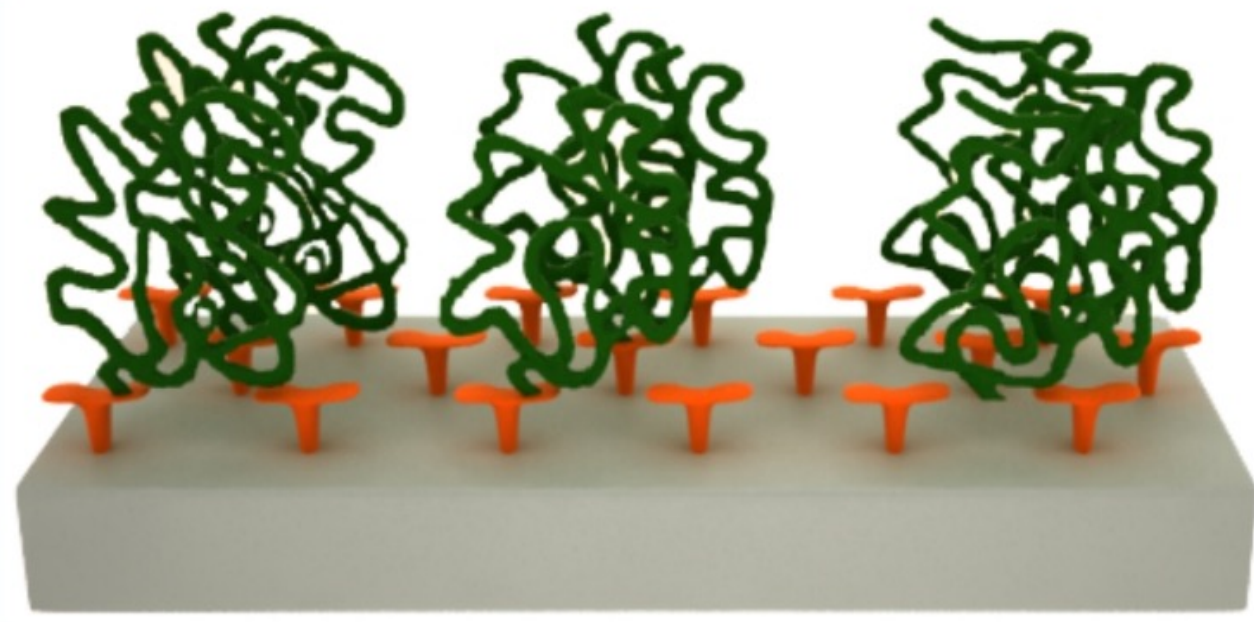
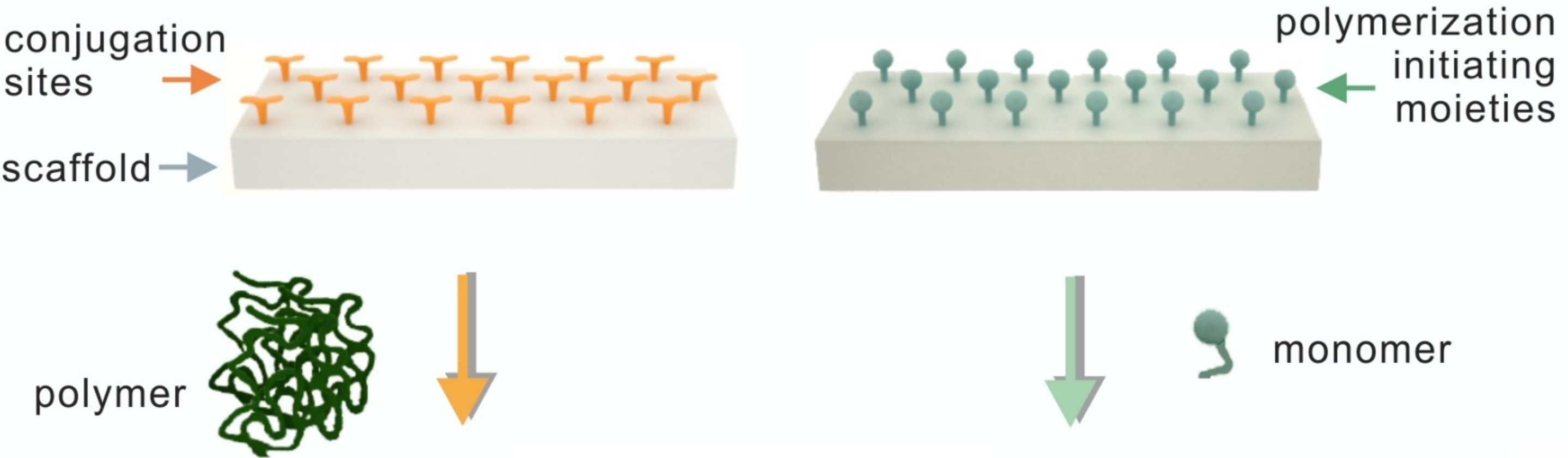


Middle density:  
Overlapping separated chains

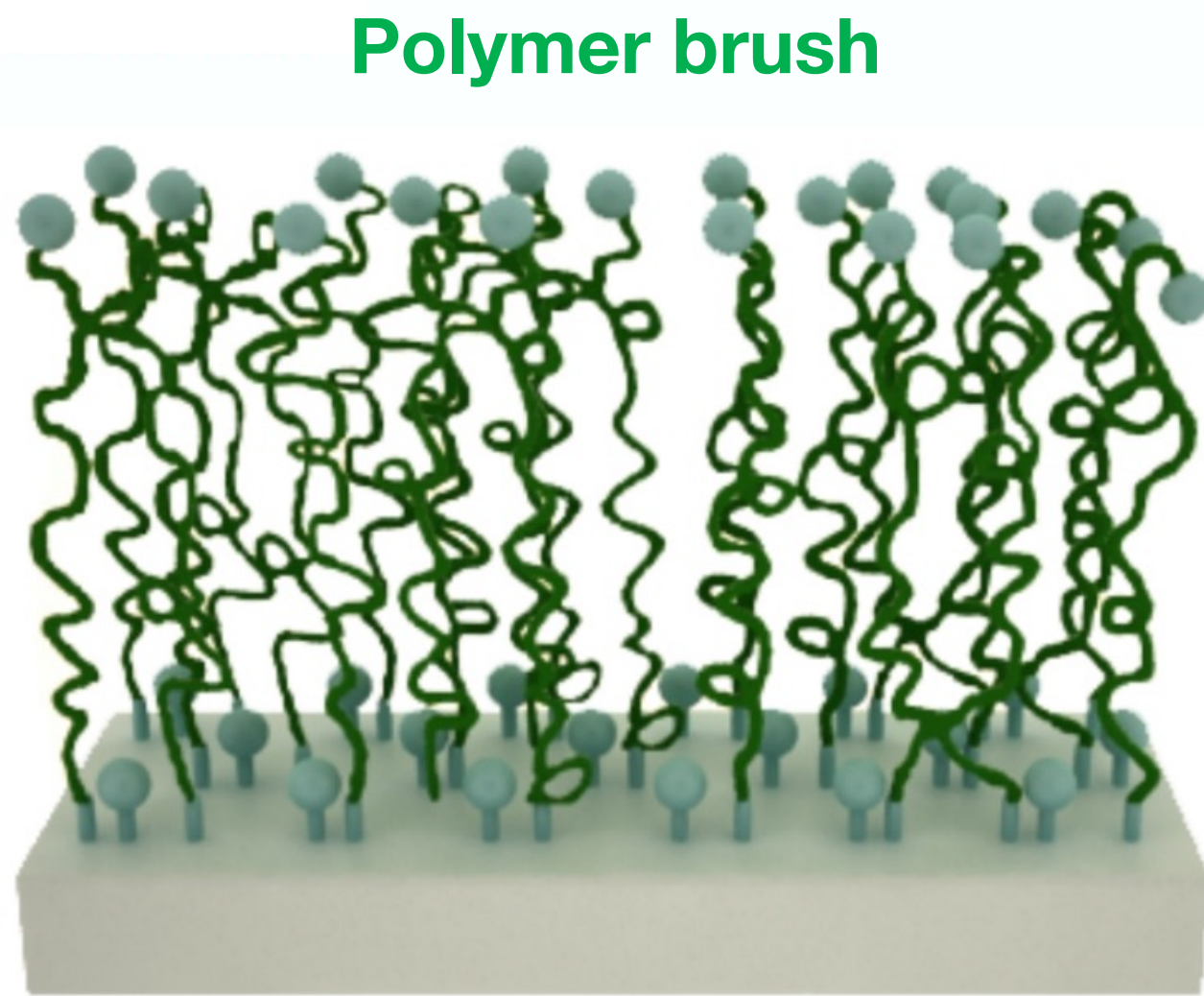


High density:  
crowding → chain extension

# Polymers on Surfaces – Grafting Density Influences Structure



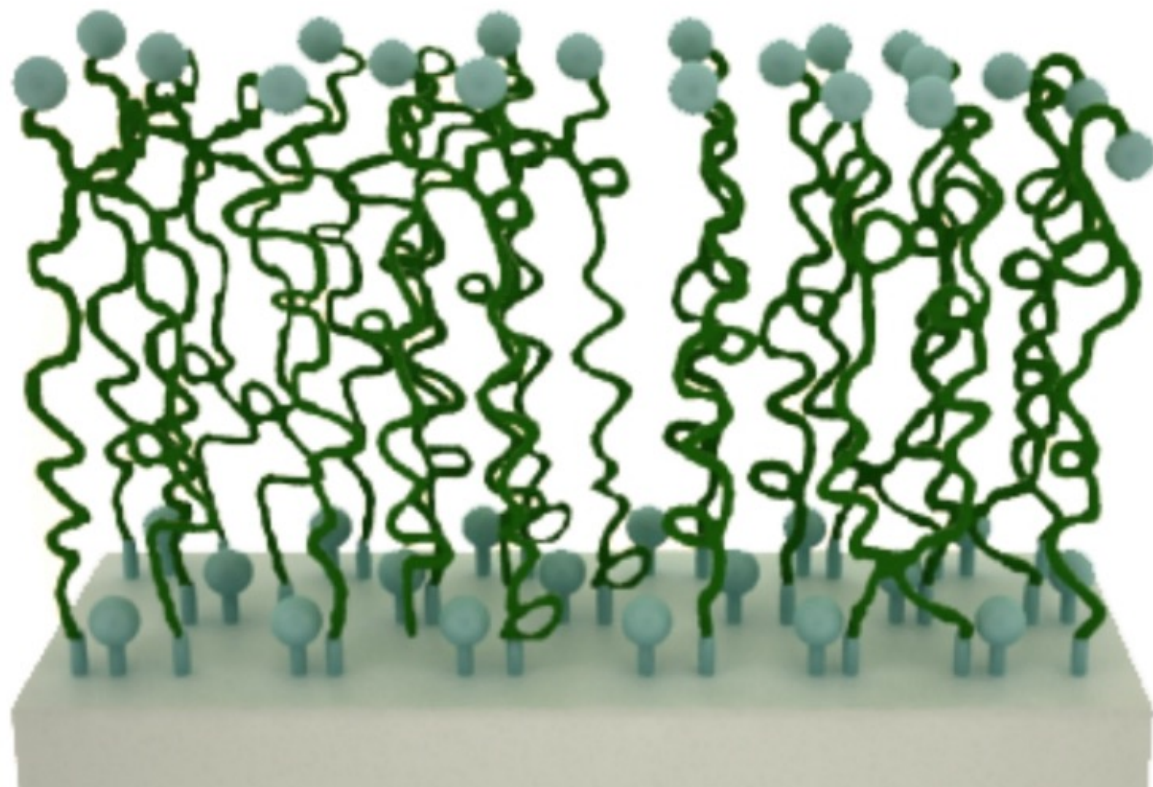
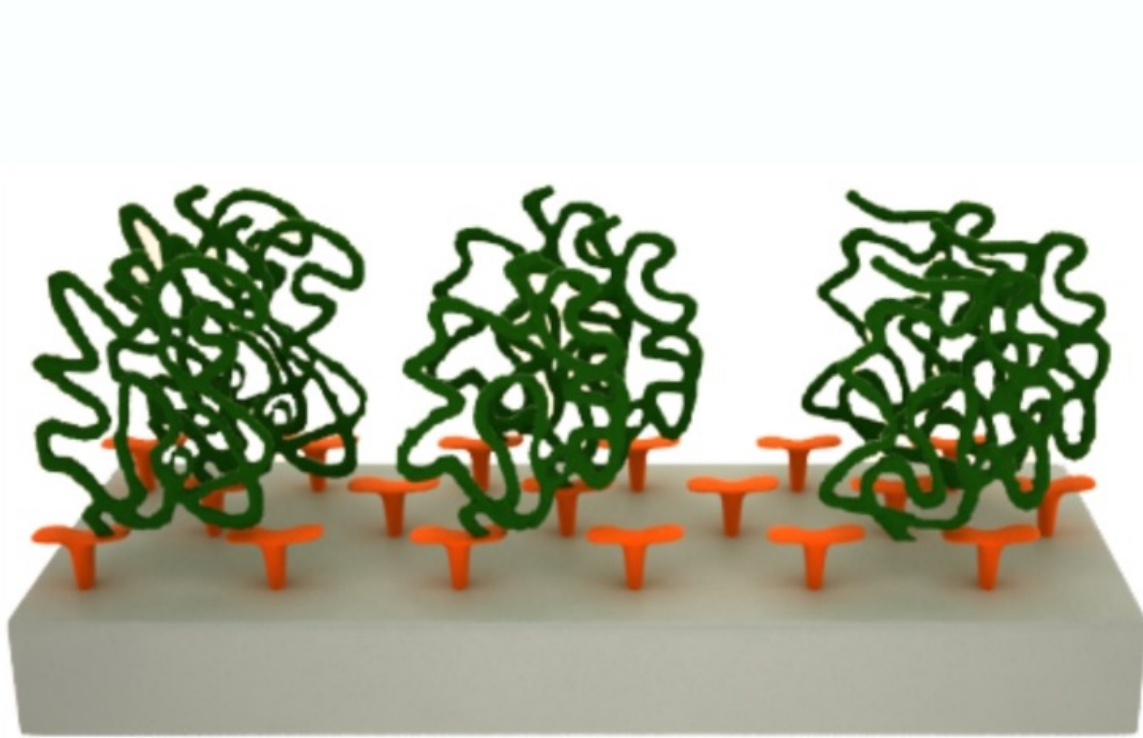
chemical conjugation between polymers and scaffold



surface-initiated polymerization



# Polymers on Surfaces – Grafting Density Influences Behavior



Polymer brush

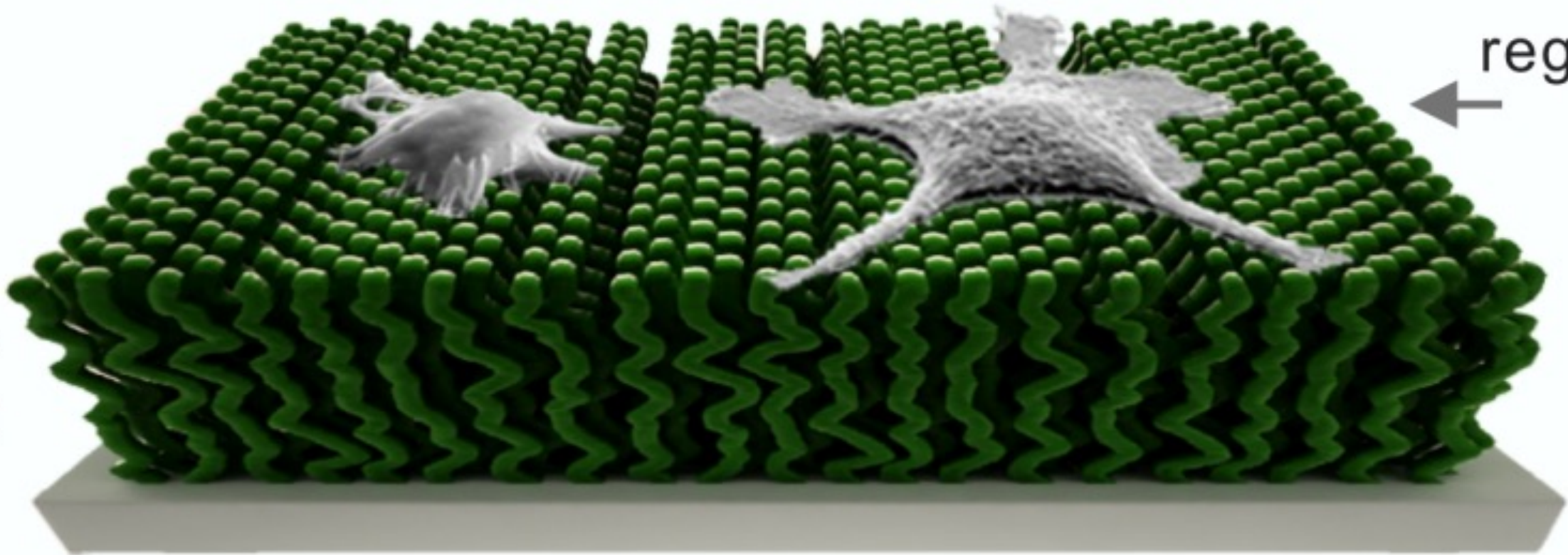
$$D < 2R_g$$

Distance between adjacent grafting points

Radius of gyration of polymer

stem cell → differentiated cell

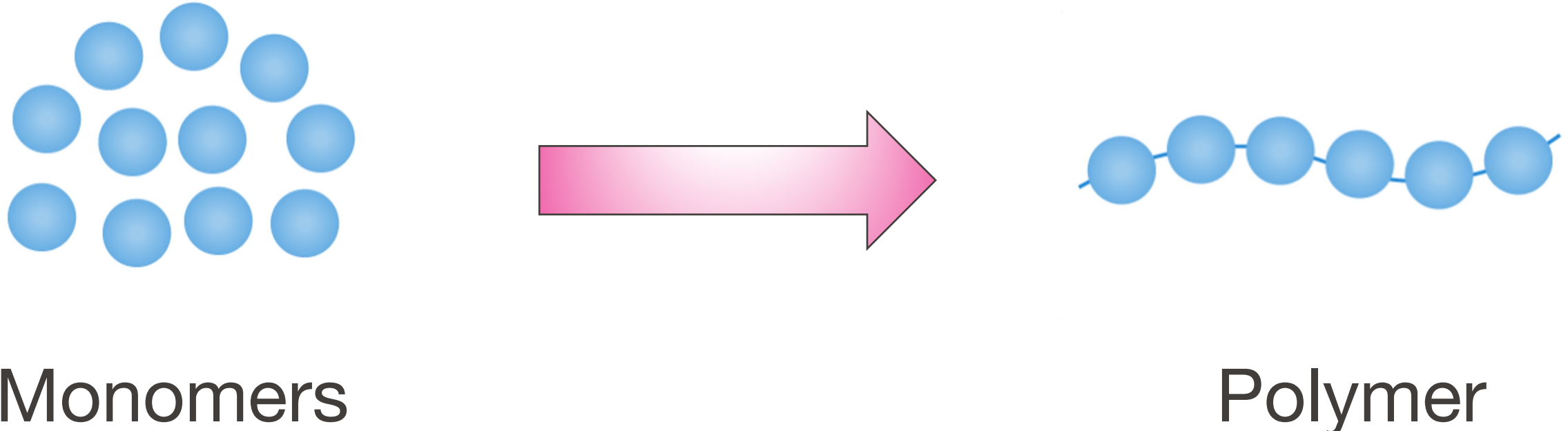
structured polymer brush →



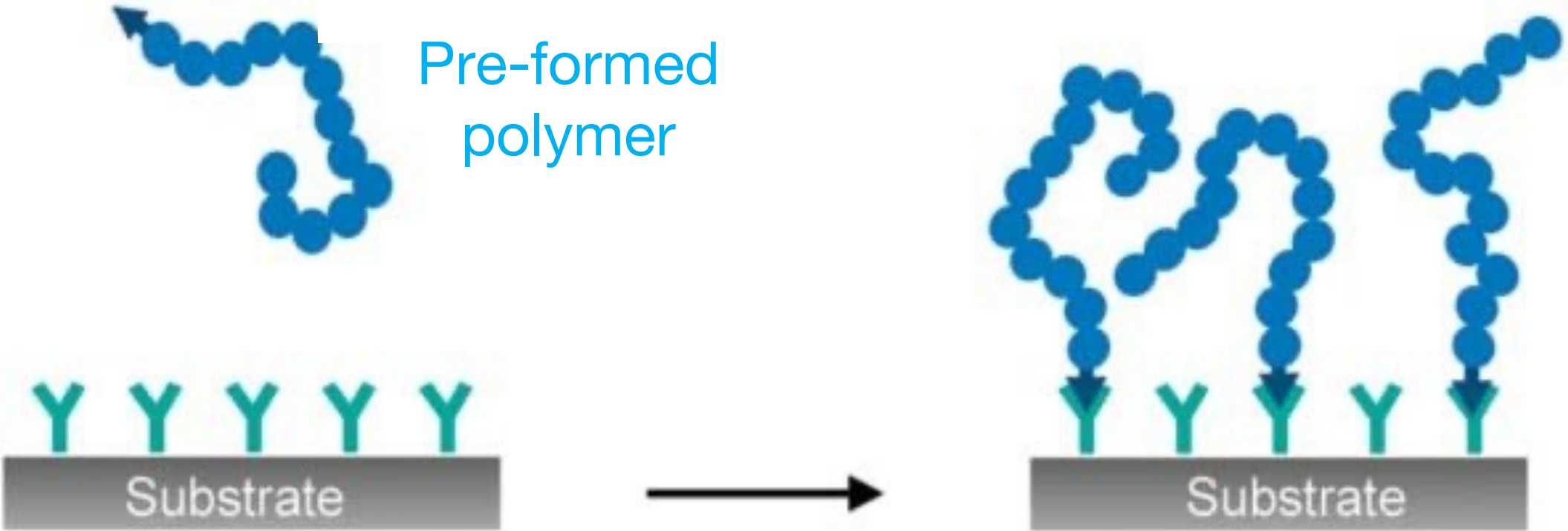
regulation of stem cell

Stem cell differentiation

# Surface Assembly of Polymers on Surfaces



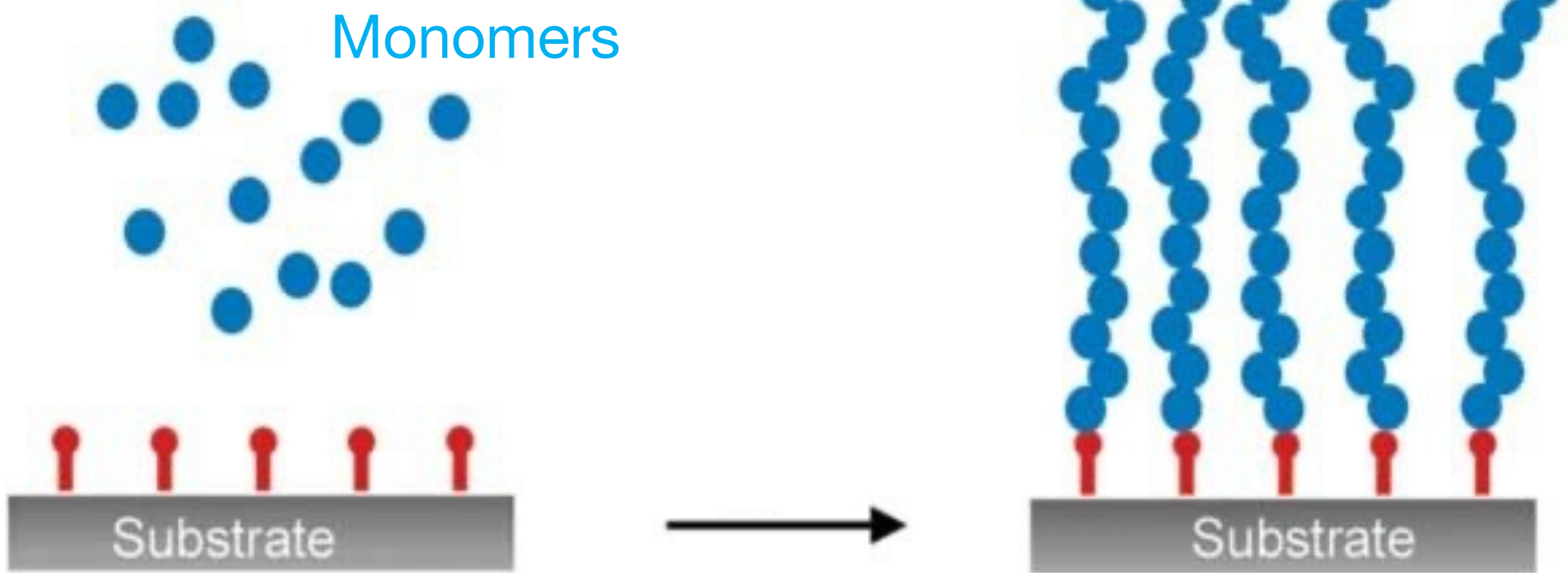
Top-down:  
**Grafting-to approach**



Control over polymer properties

Limited surface density  
steric hindrance

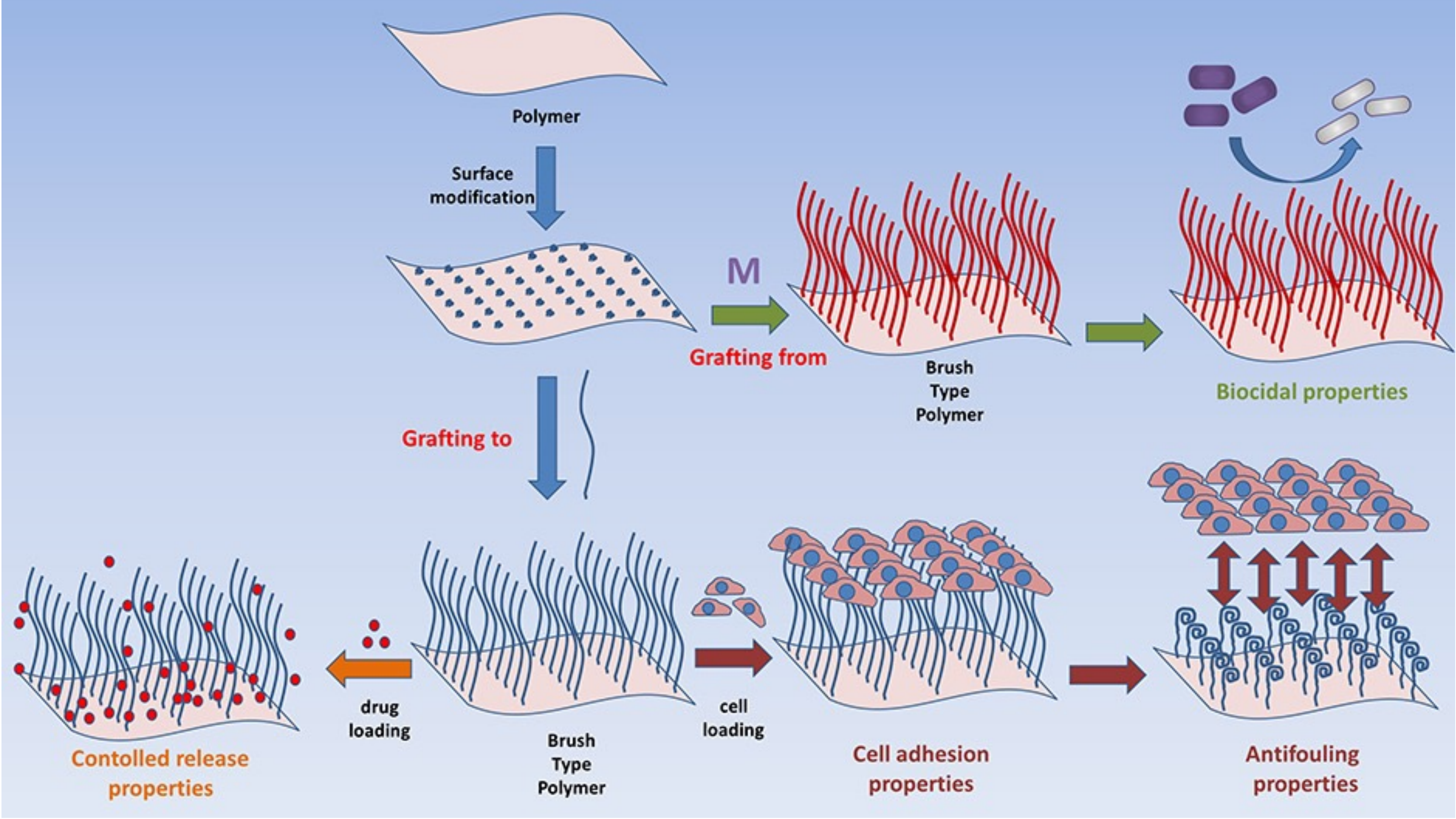
Bottom-up:  
**Grafting-from approach**



High grafting density  
and stronger bonding

Less control over  
polymer properties

# Different Grafting Routes for Different Applications

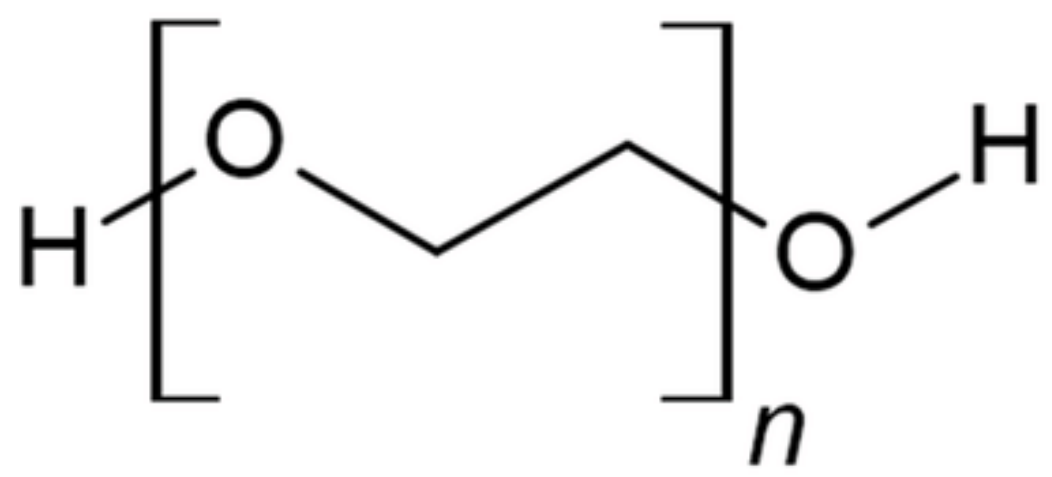


# Comparing Polymer Brushes to SAMs

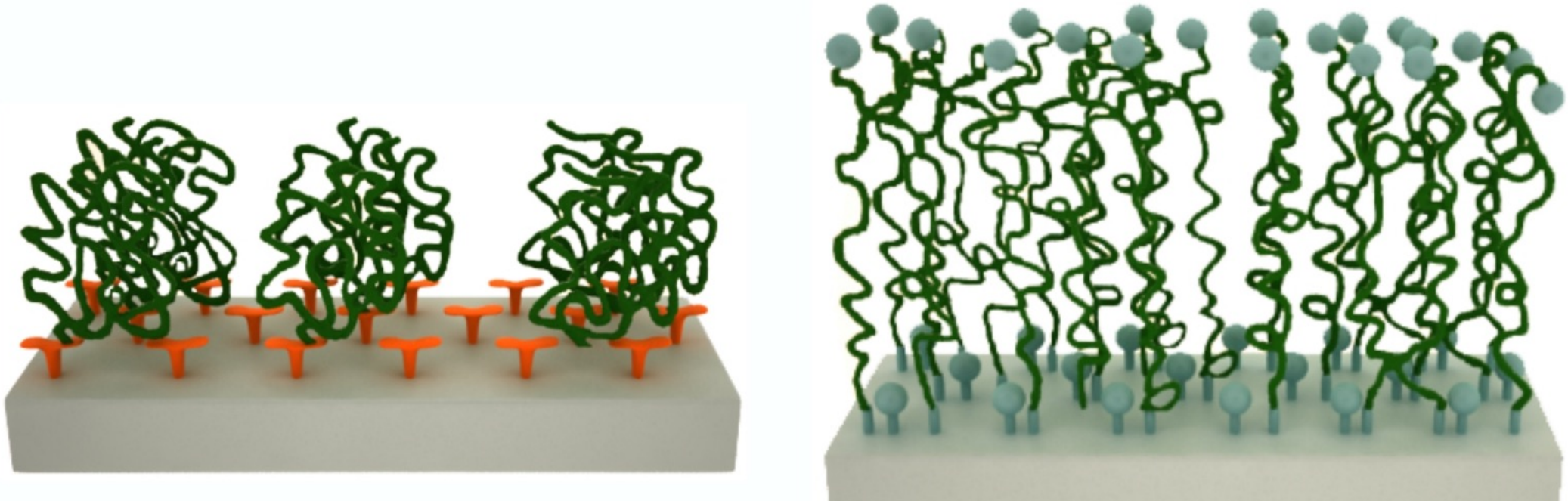
	Grafting-to Polymer Brush	Grafting-from Polymer Brush	SAM
Fabrication	Attach pre-made polymers	Grow polymers from surface	Molecules self-assemble
Density	Low	High	High
Stability	Good in biological media	Very good in biological media	Lower in biological media
Defects	More gaps	Fewer (self-repair)	Can have pinholes
Thickness	Medium	Thick, tunable	Very thin (1 layer)

Kim & Jung, *Phys. BMB Rep.*, 49, 12, 2016

# Different Types of Polymer Brushes: Neutral

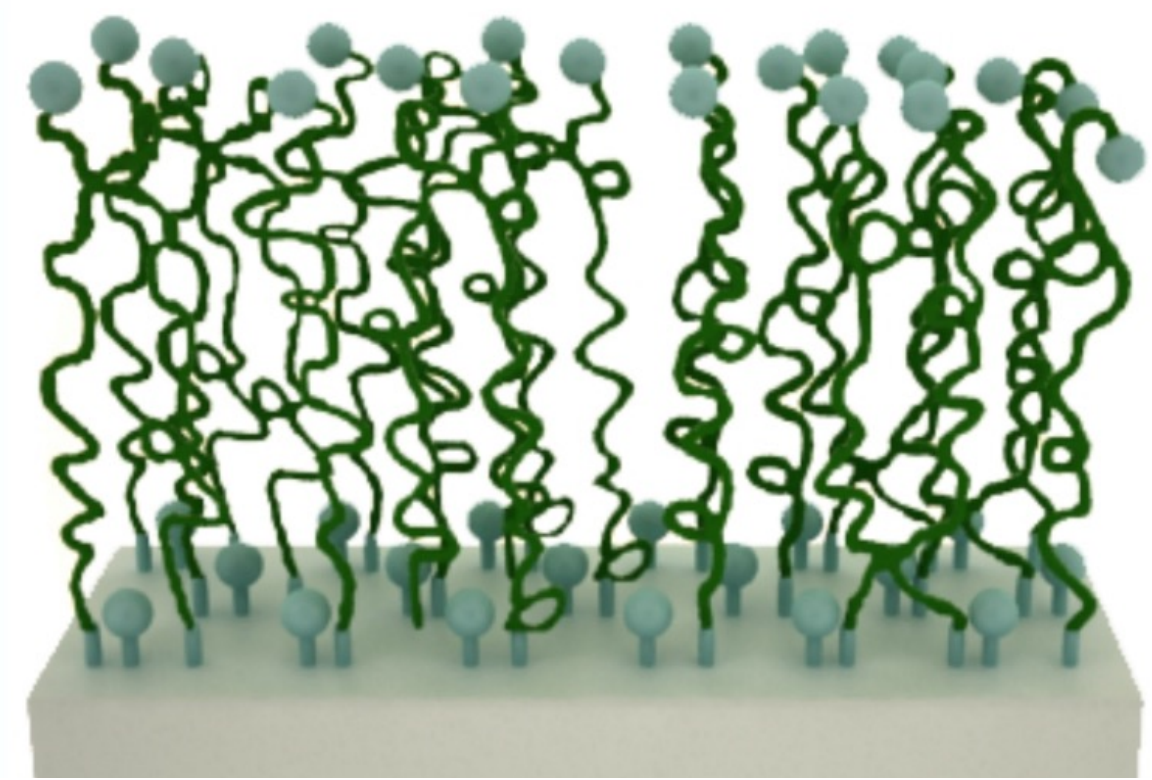


Poly(ethylene) glycol (PEG)



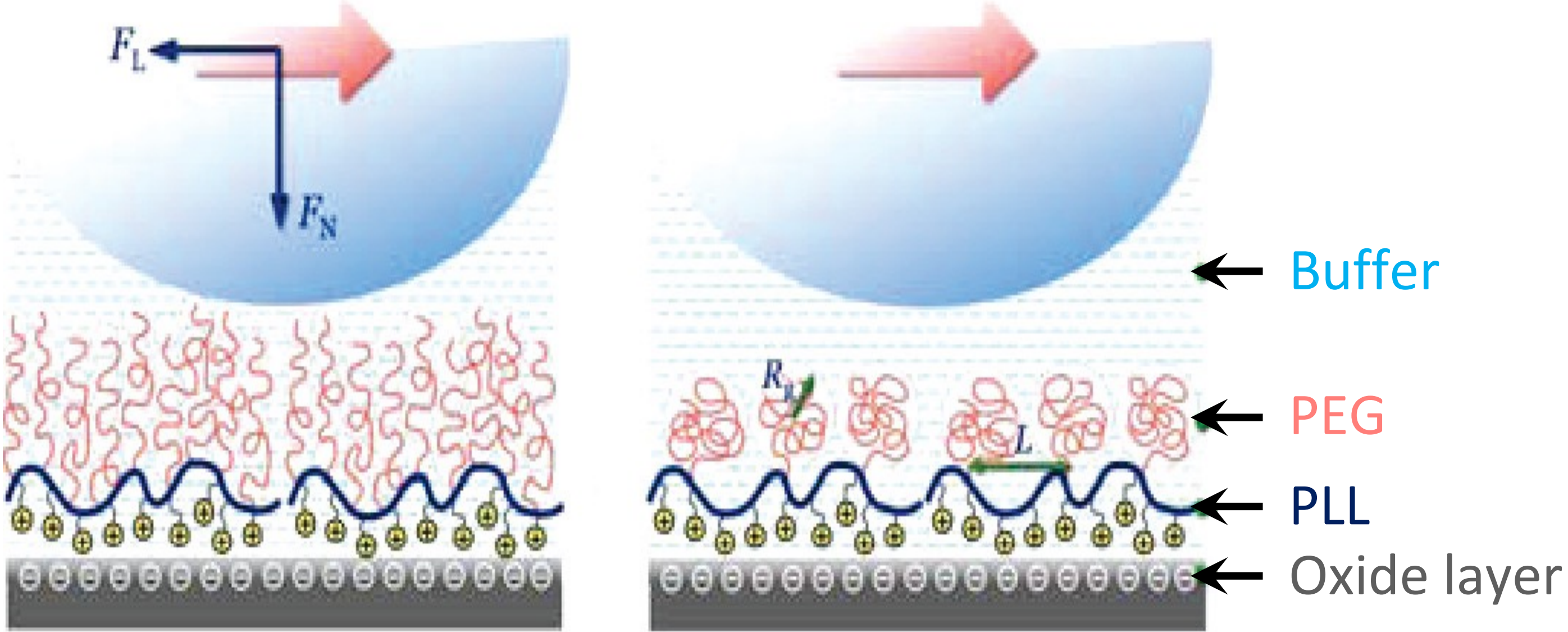
Neutral brushes  
 Highly hydrated and flexible  
 Antifouling benchmark

## 1. Dense PEG layer = steric barrier



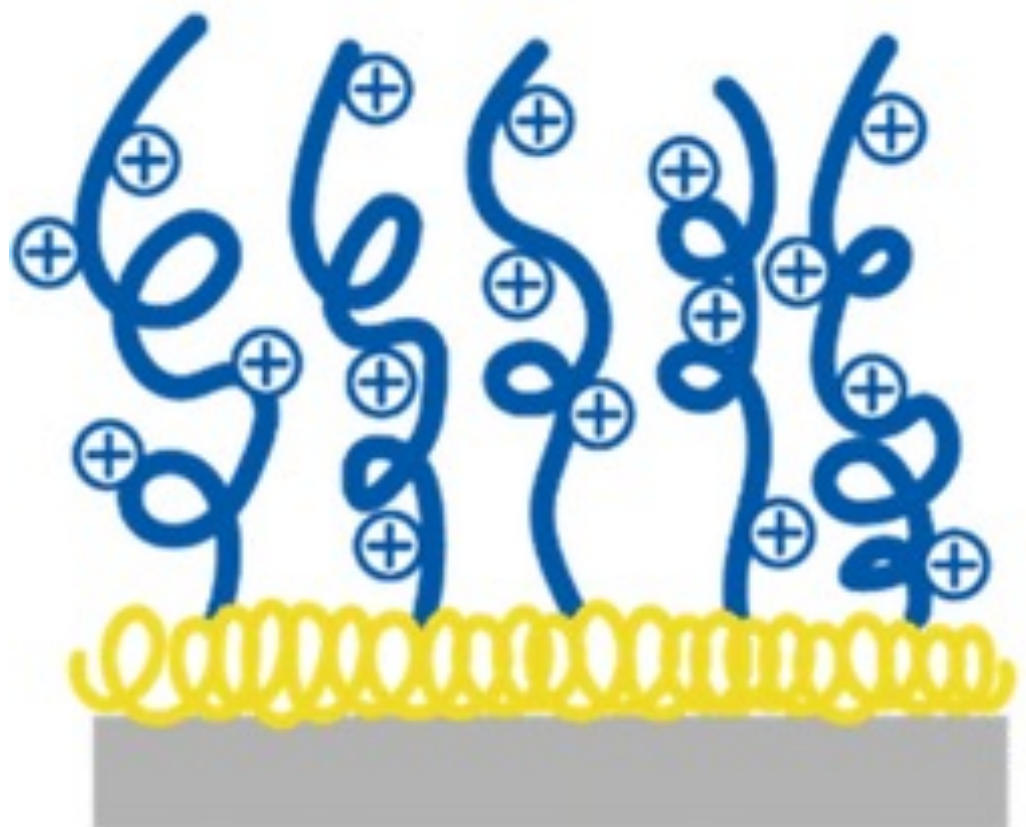
Durmaz et al. | ACS Appl. Polymer Mater. | 2021

## 2. Hydration and dynamic flexibility of PEG brush

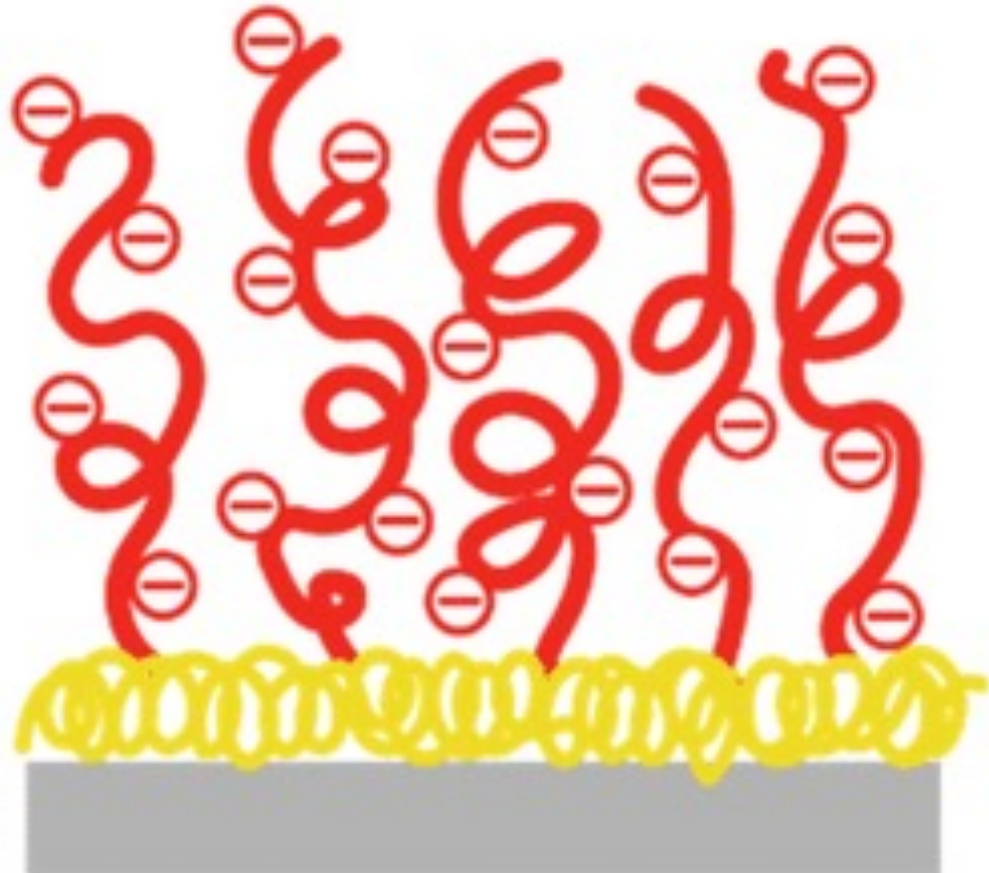


Wei et al. | ACS. Appl. Mater. Interf. | 2009

# Different Types of Polymer Brushes: Charged



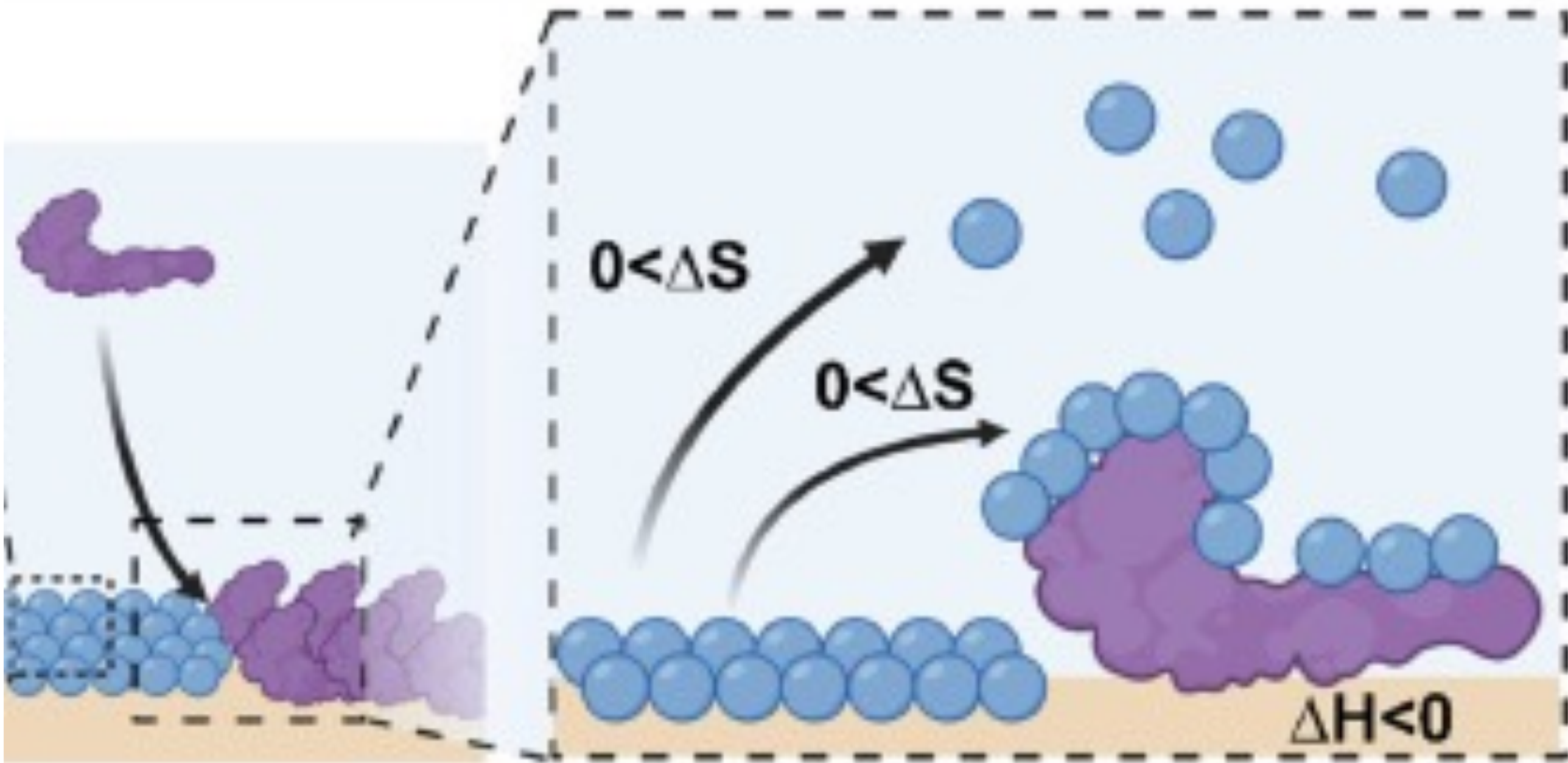
Cationic polymers



Anionic polymers

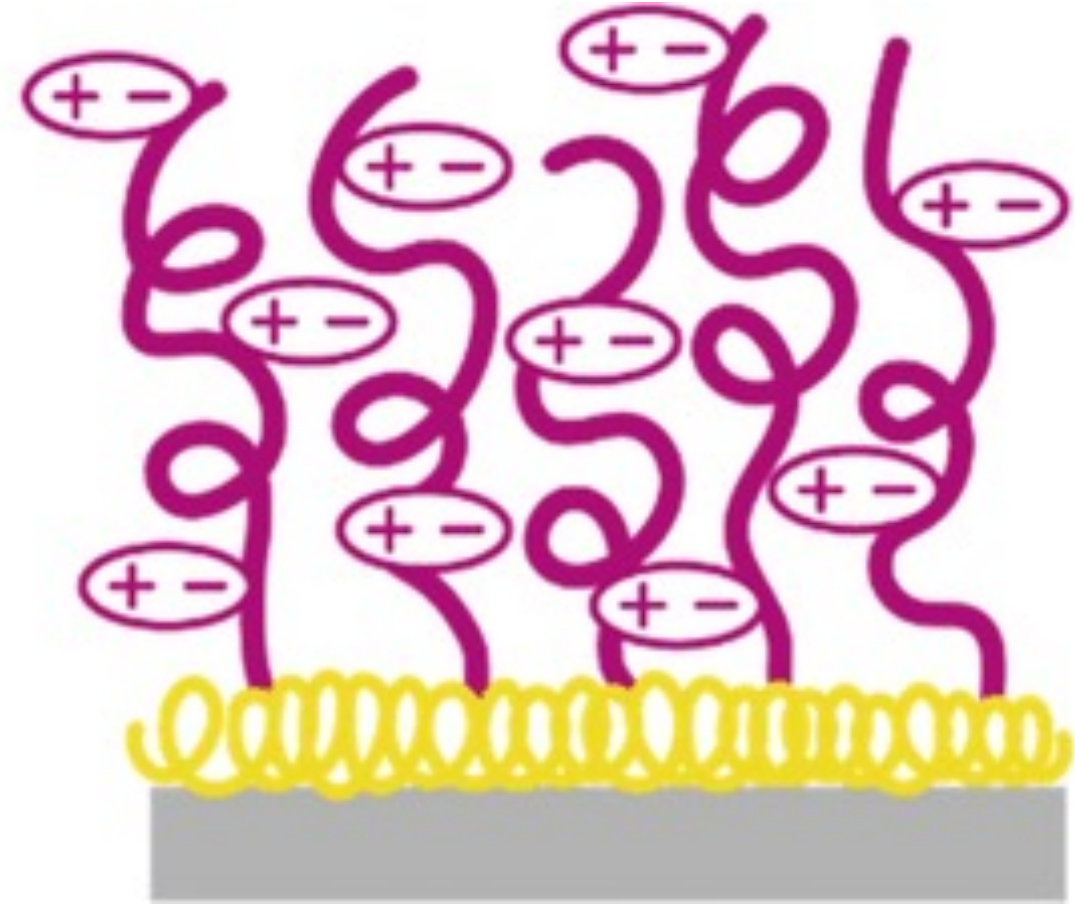
Charged polymers promote electrostatic adhesion

At physiological pH, majority of proteins are negatively charged



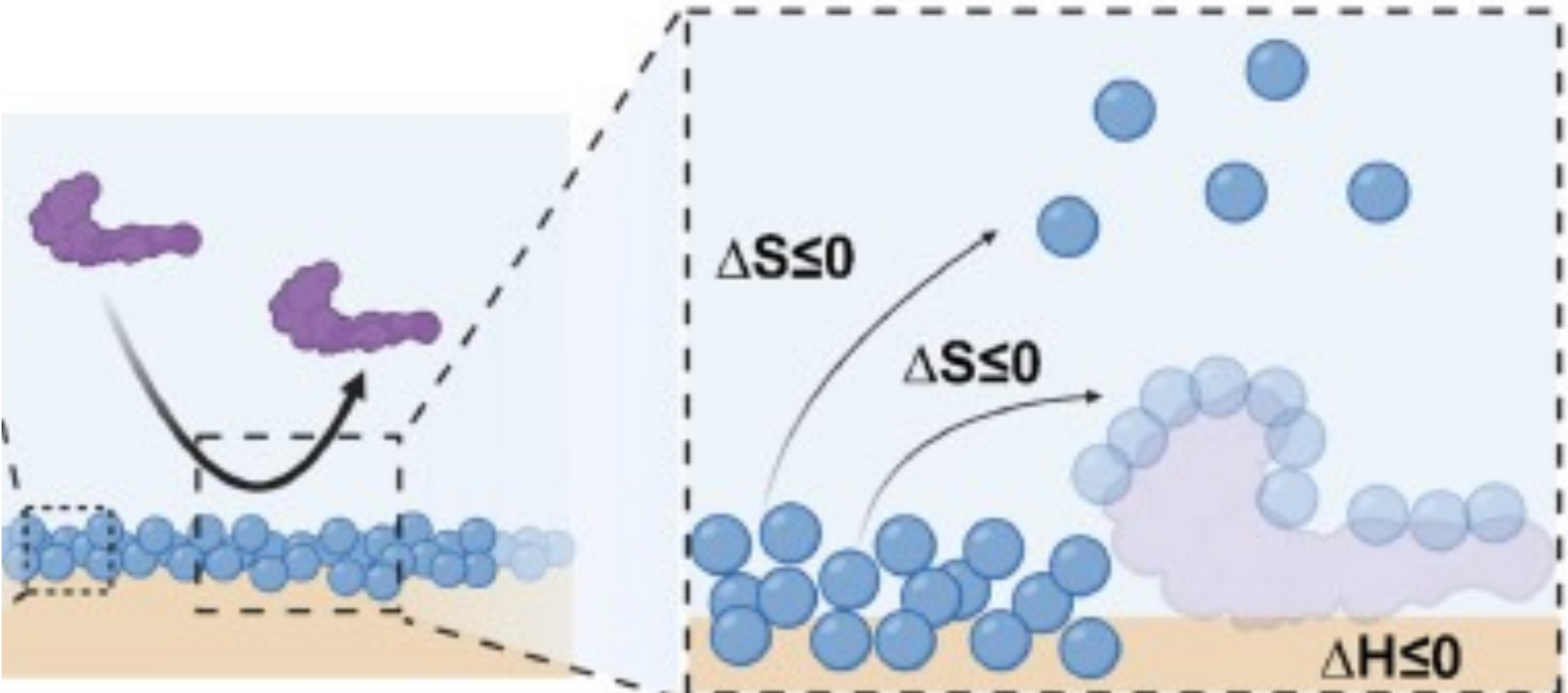
Ordered water release leads to entropy increase  
Protein forms electrostatic interactions with the surface

# Different Types of Polymer Brushes: Zwitterionic Polymers



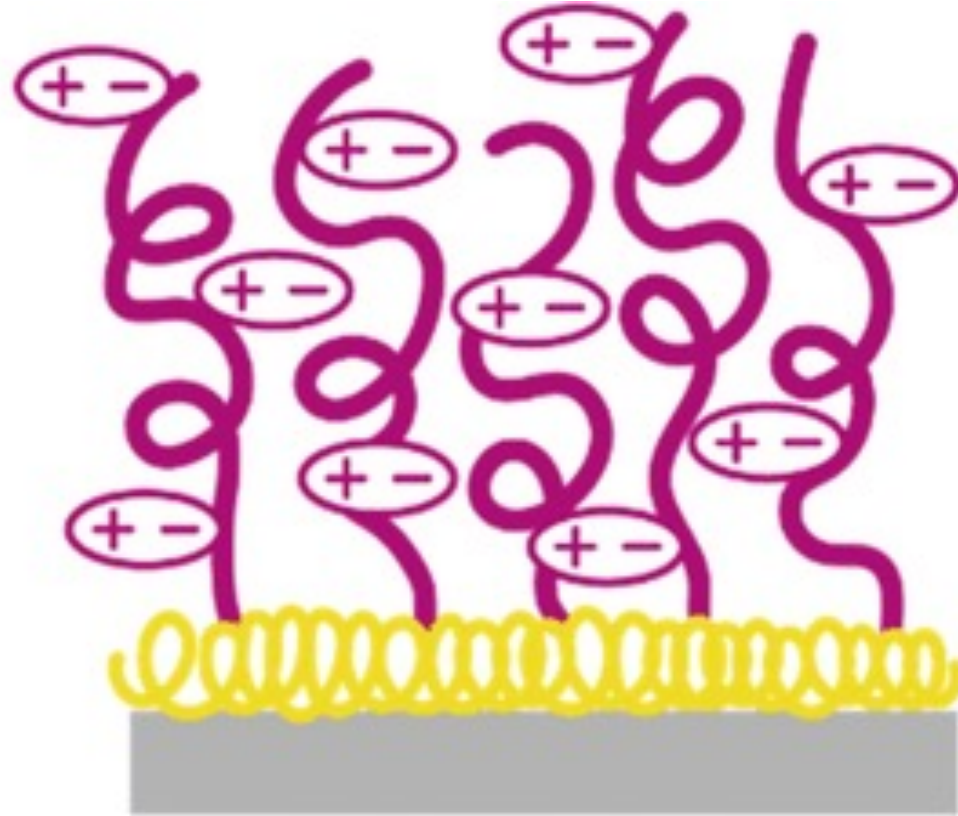
Zwitterionic polymers

Stronger hydration layer than PEG – better at resisting protein adsorption

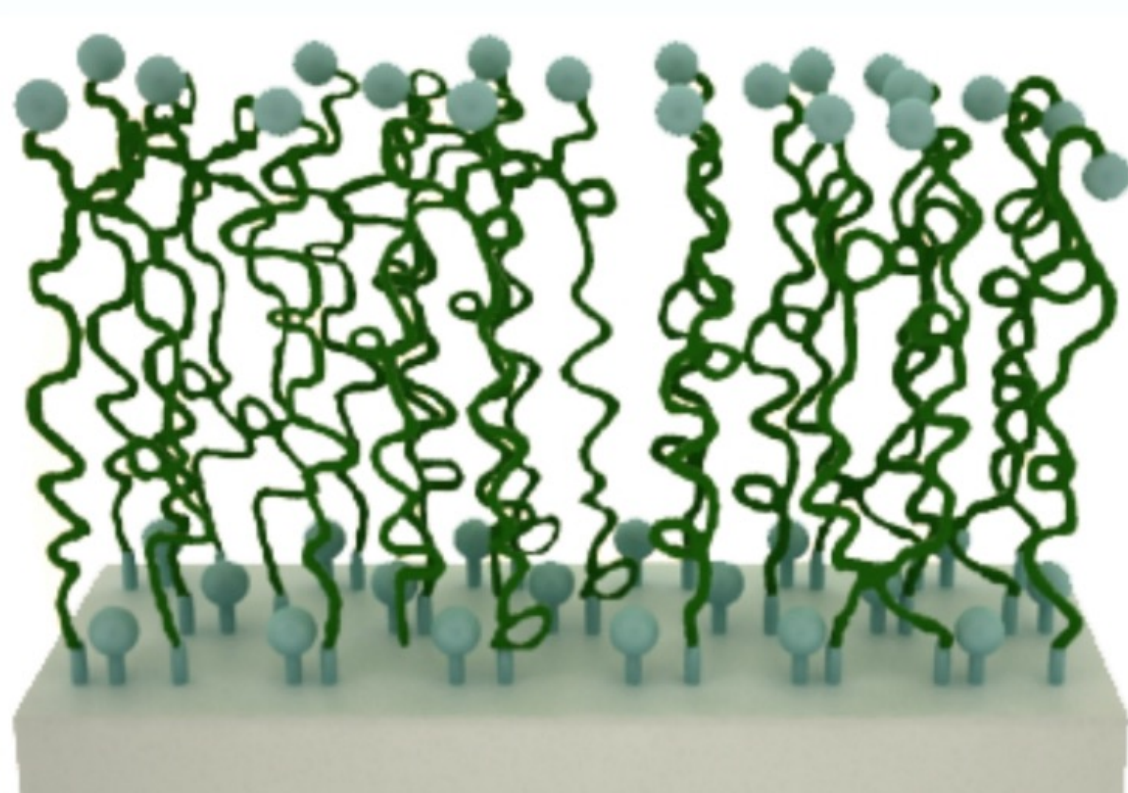


Zwitterionic surfaces resist protein adsorption because the water near the surface is already stable, so removing it costs entropy

# Different Types of Polymer Brushes: Zwitterionic Polymers



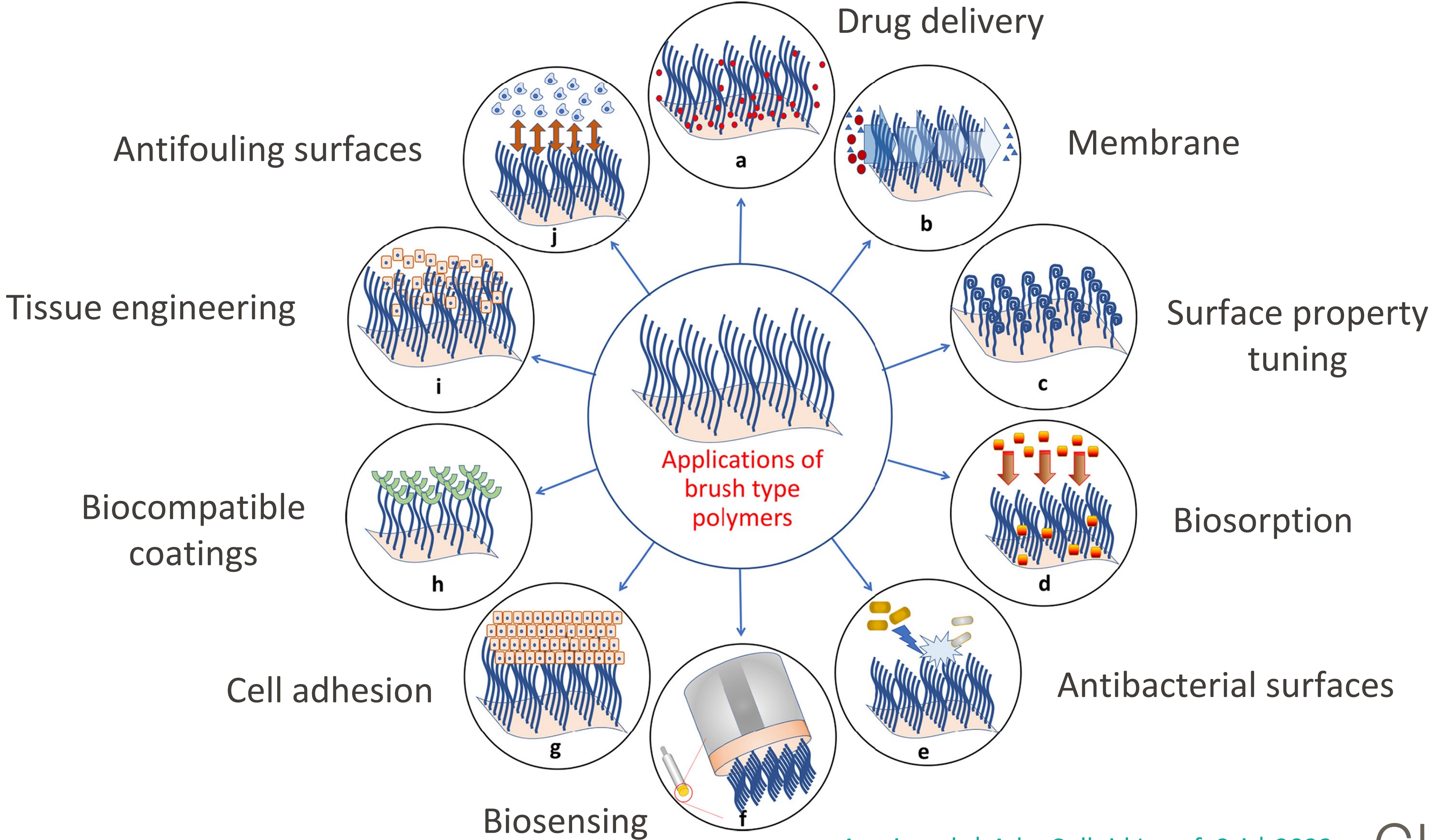
**Zwitterionic polymers**



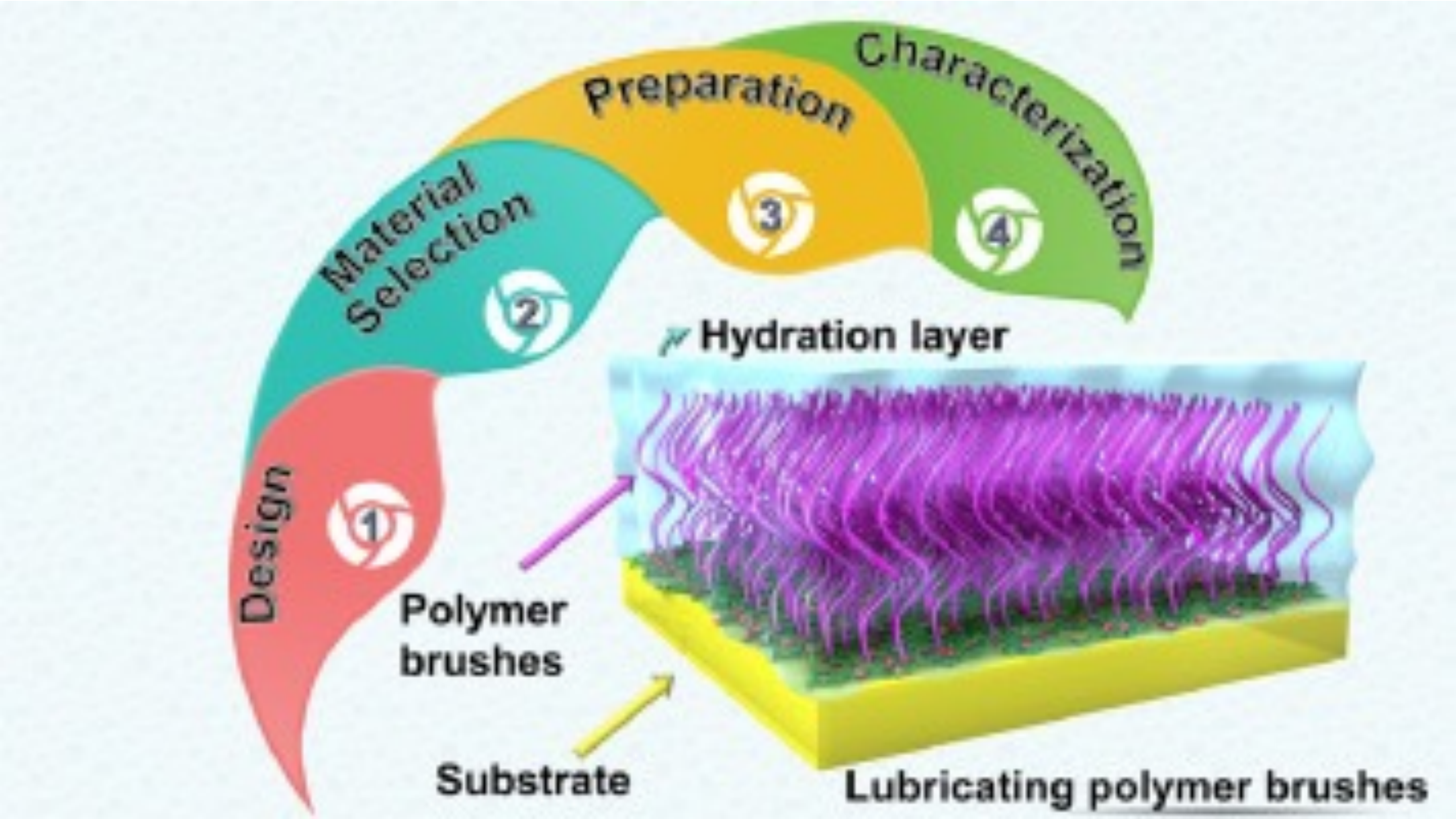
**Neutral PEG**

<b>Type of interaction</b>	Ion–dipole (electrostatic)	Hydrogen bonding (dipole–dipole)
<b>Hydration layer strength</b>	Strong, stable	Moderate, dynamic
<b>Water exchange rate</b>	Slow (tight binding)	Fast
<b>Ionic strength sensitivity</b>	Robust (maintains hydration)	Sensitive (can collapse)
<b>Antifouling performance</b>	Often superior in biological fluids	Excellent

# Biomedical Applications of Brush-Type Polymers

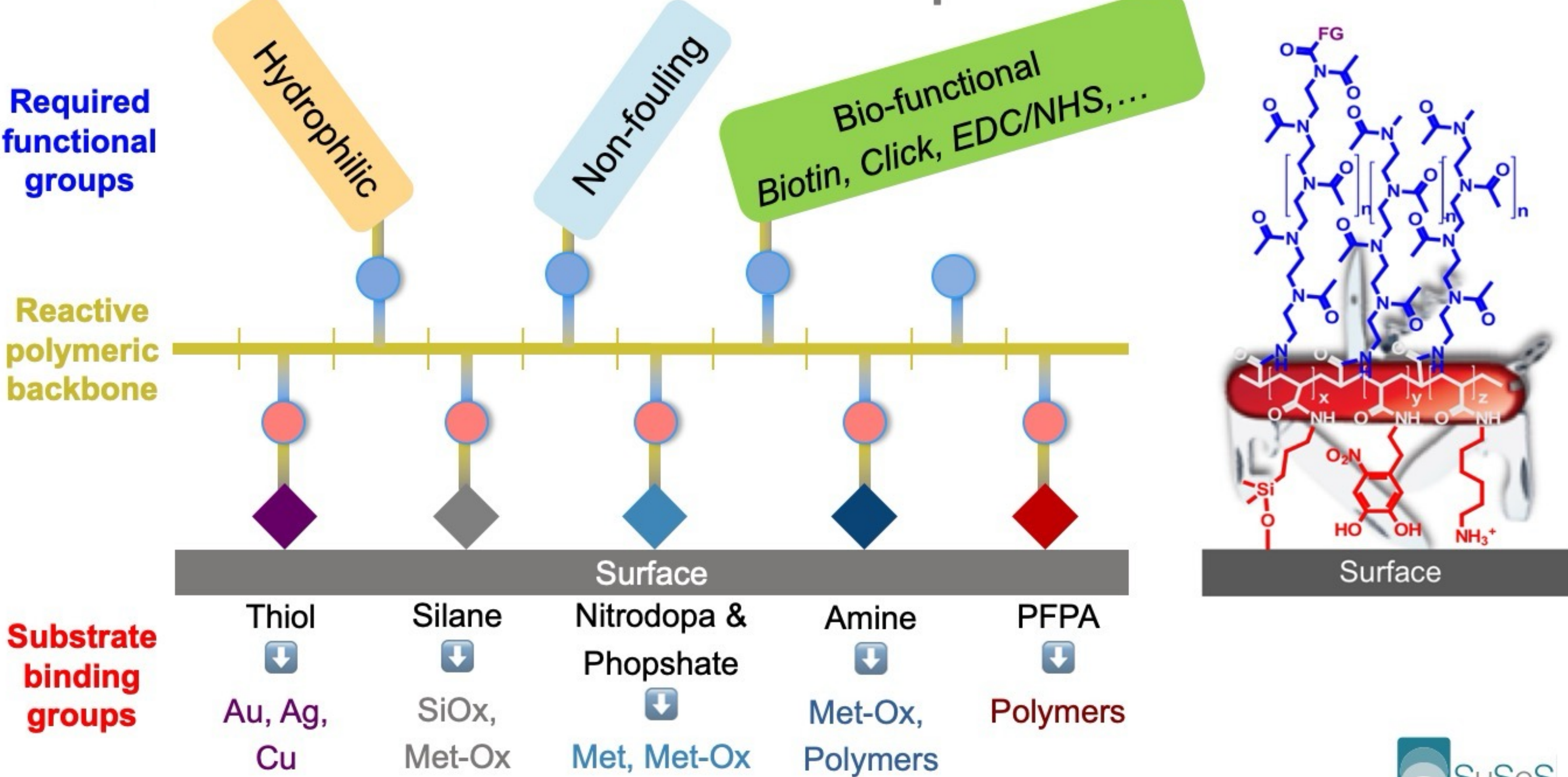


# From Design to Characterization of Polymer Brushes



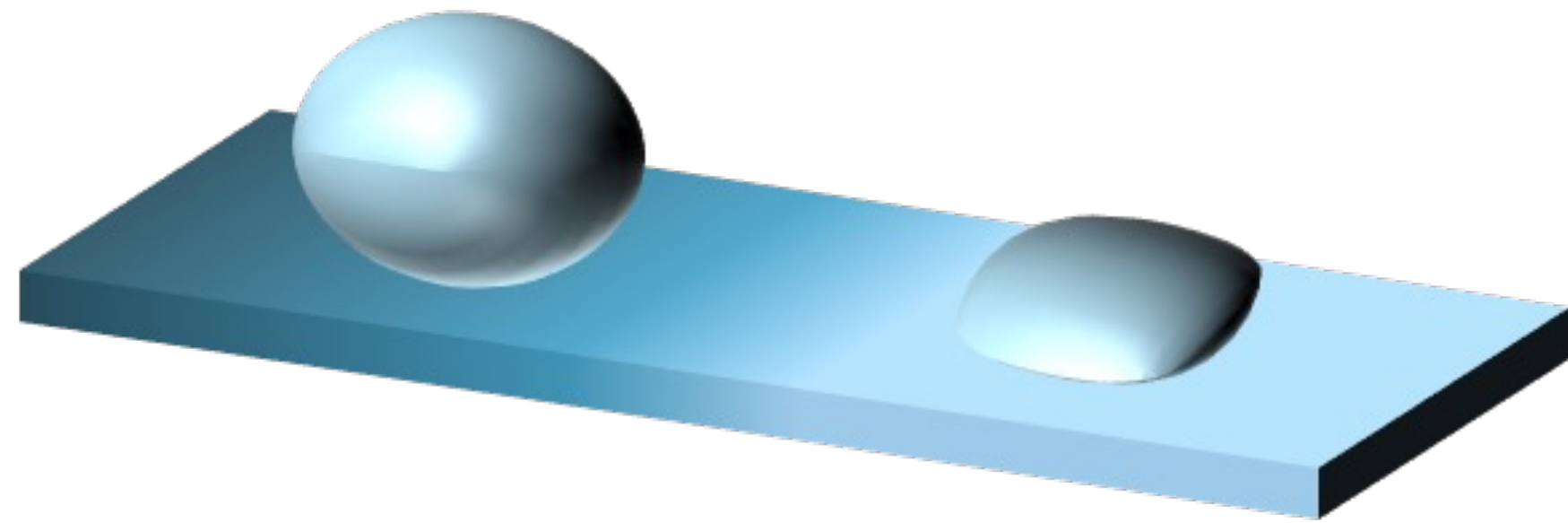
# The “Swiss Army Knife” of Surface Chemistry with Polymers

## PAcrAm™ – molecular pocket-knife

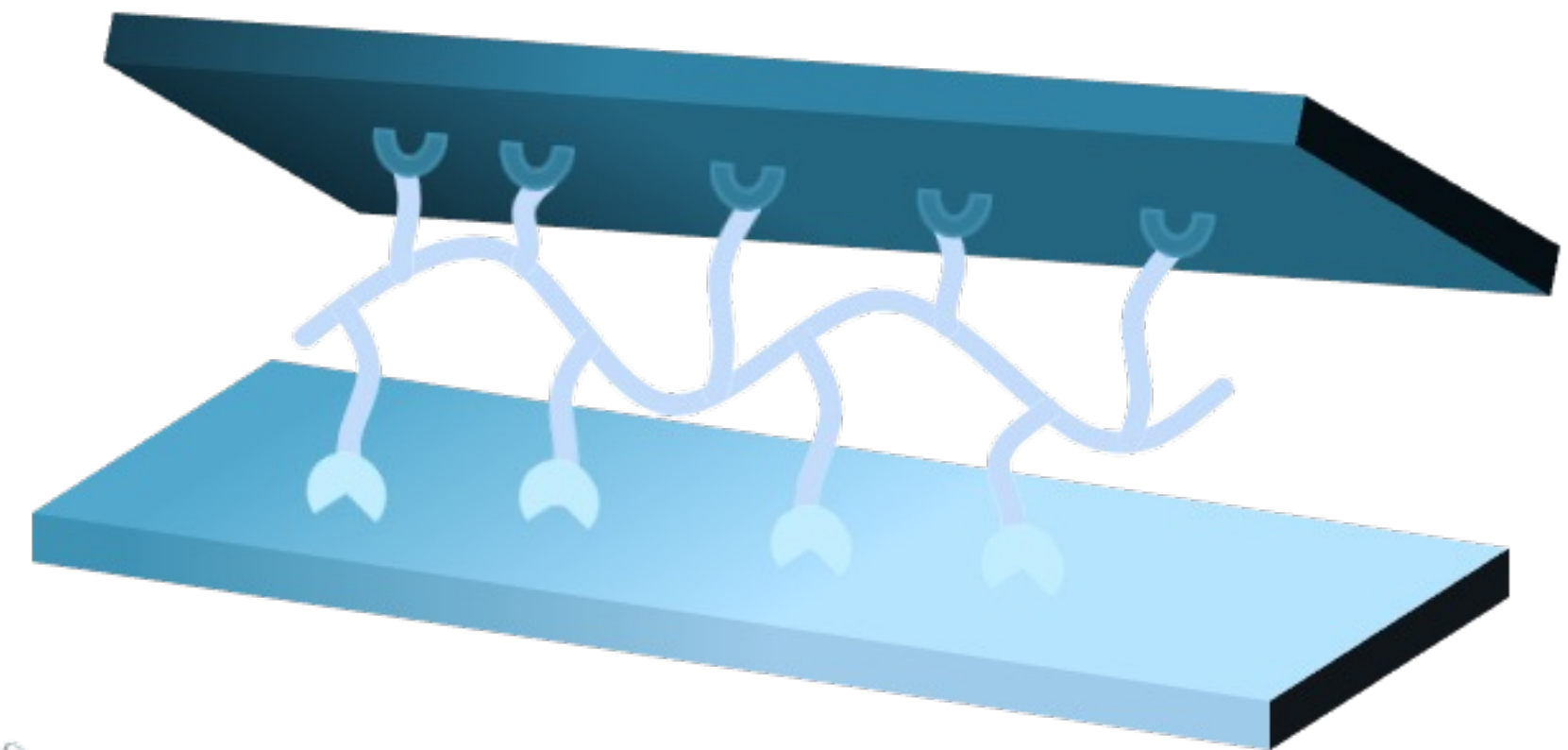


# Core Applications for “Swiss Army Knife” Polymers

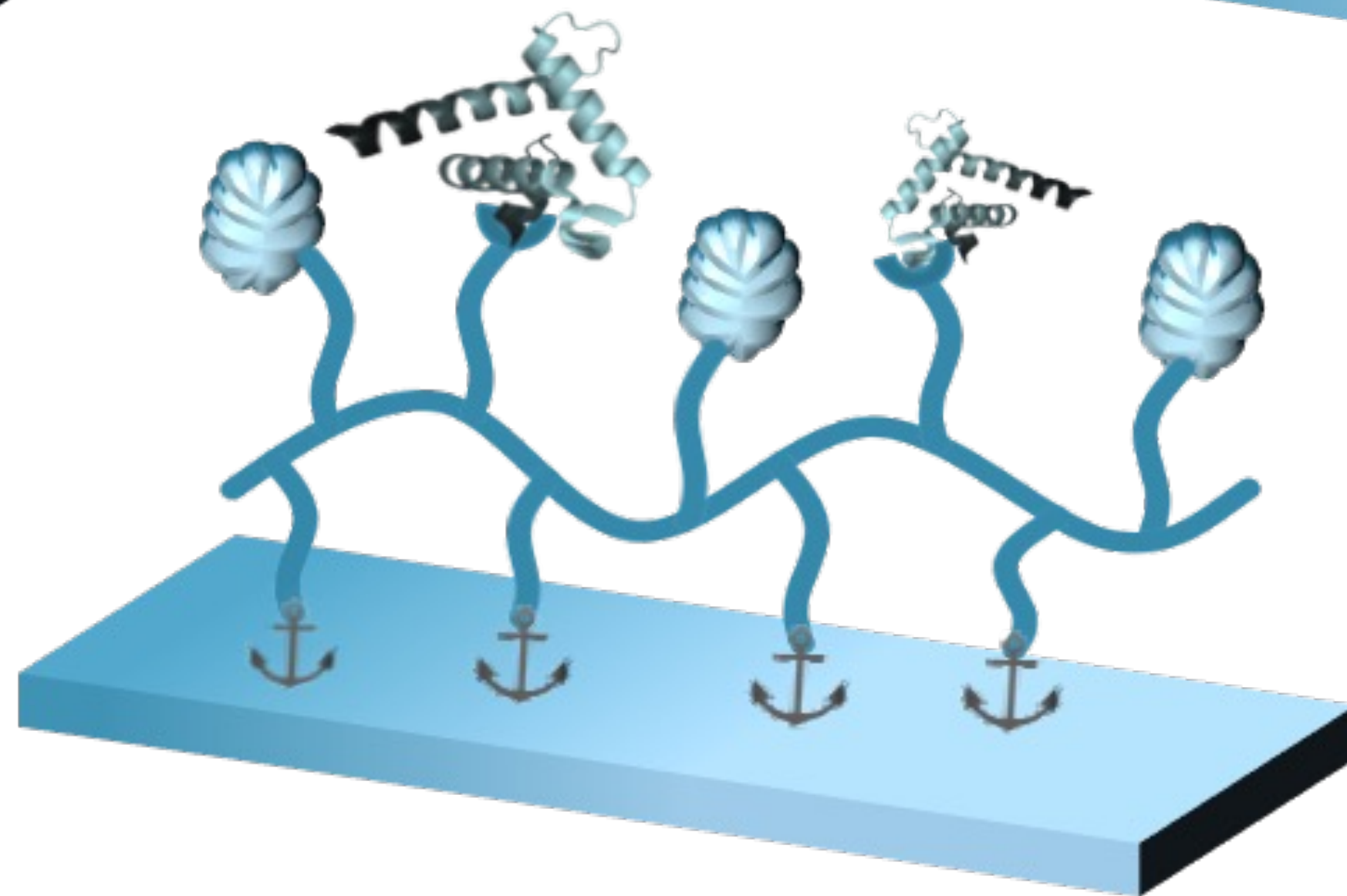
Tuning surface energy using functional molecular layers allow control of the wettability of surfaces



Molecular “super glue” bonding two surfaces together with polymer layers



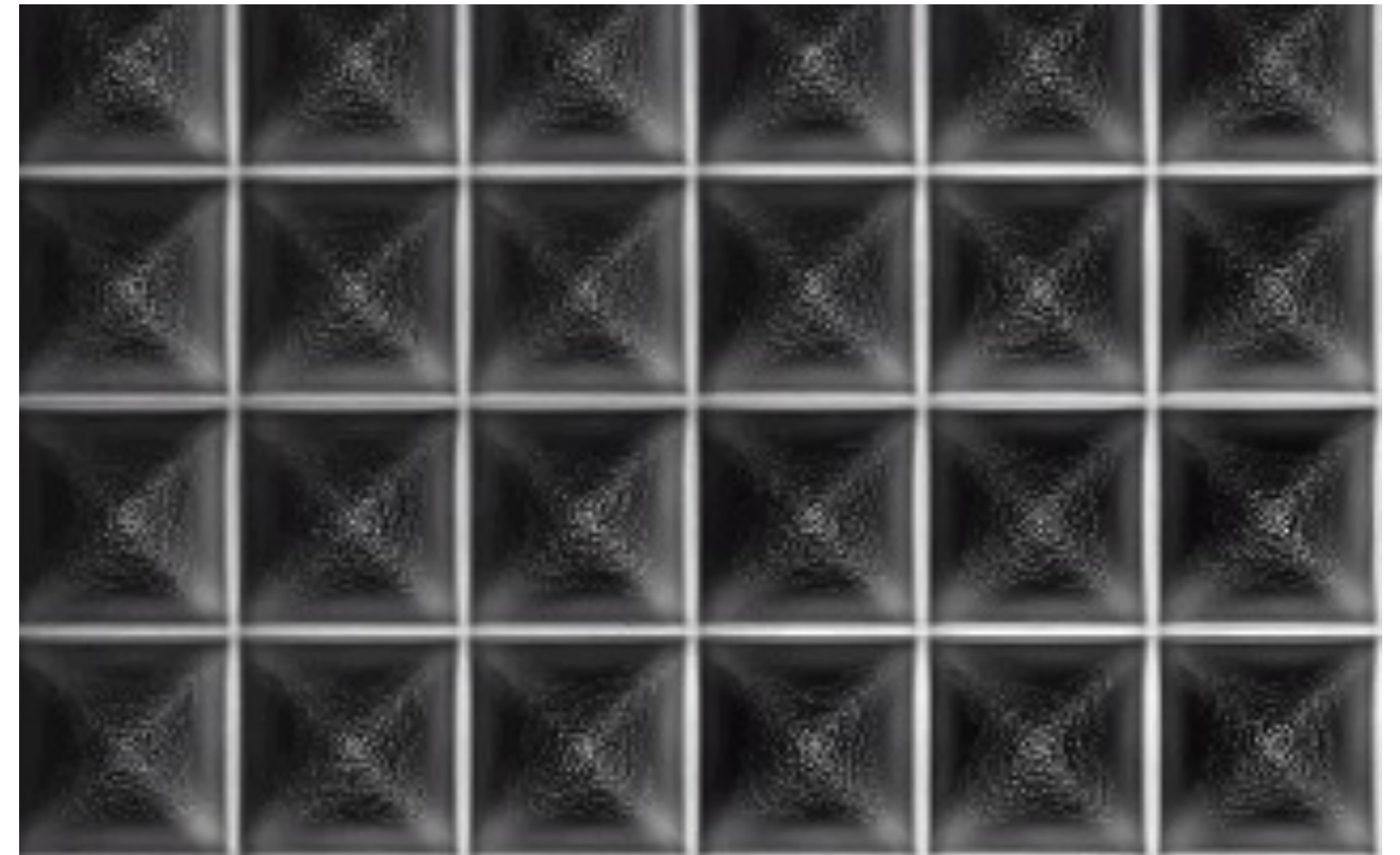
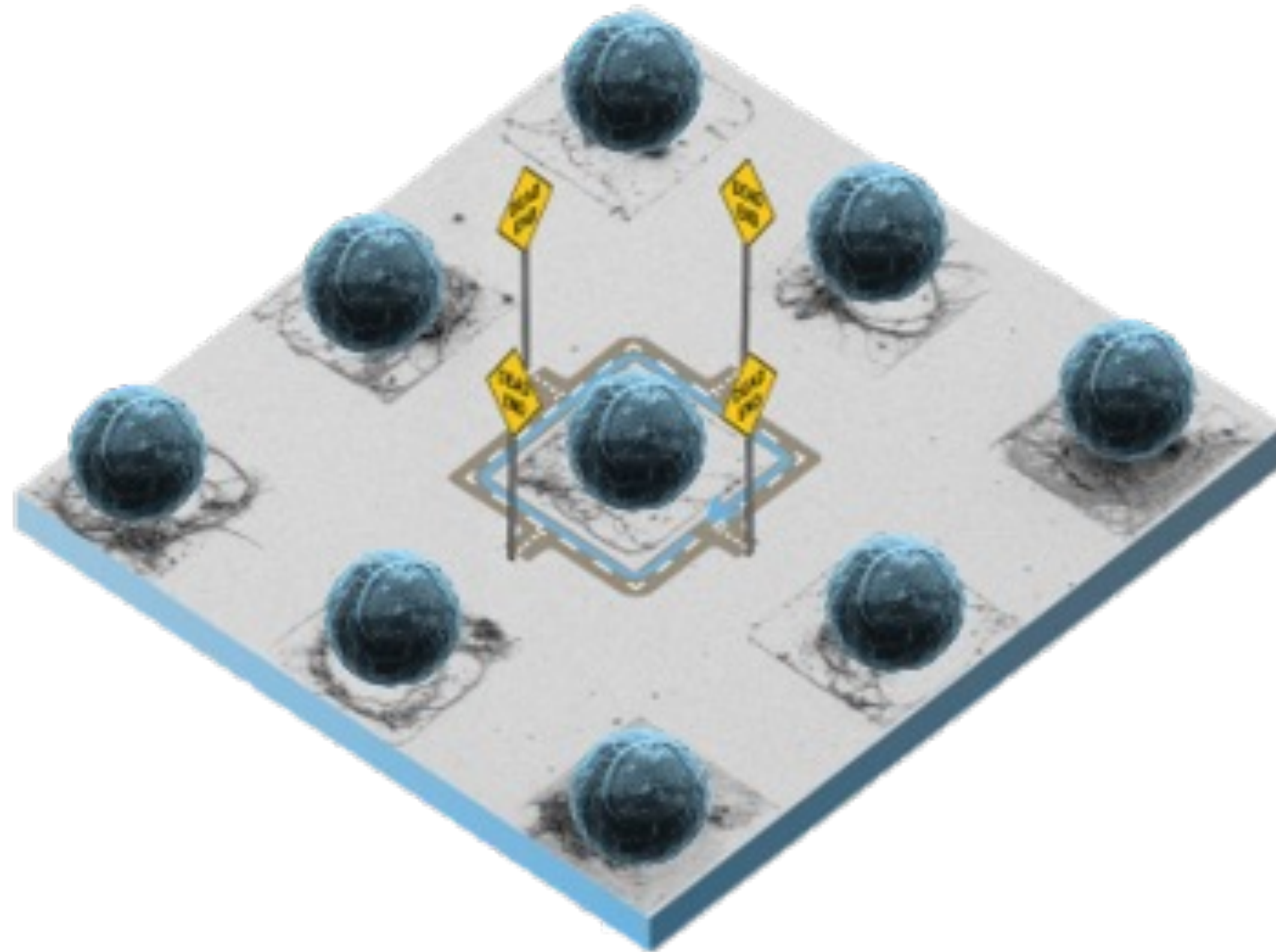
Specific bio-functionalization with specific binding sites surrounded by non-fouling groups



# Core Applications for “Swiss Army Knife” Polymers

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Formation of spheroids (3-D cell aggregates) as *in vitro* models in wells coated with polymers



# Key Takeaways

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- Polymer brushes (neutral, zwitterionic) minimize protein adsorption
  - Polymer chains on surfaces – grafting density and structure
  - Polymers on surfaces can influence biological behavior
    - Assembly routes for polymers on surfaces
  - Multifunctional polymers for diverse applications

# Summary of Today's Class

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- Coupling chemistries at surface
- Molecular patterning on surfaces from micro to nanoscale
- Nonspecific protein binding on surfaces – thermodynamic mechanisms
- Controlling protein adsorption through polymer brushes with specific properties
- Tuning surface properties using surface chemistry and molecular assembly

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